

RESEARCH ARTICLE

DETERMINATION OF PROXIMATE COMPOSITION AND ANTIOXIDANT PROPERTIES OF SELECTED UNDERUTILIZED FRUITS (NAMNAM, LOVI, ROSEAPPLE, PUMELLO, AND SAPOTA) GROWN IN SRI LANKA

Baddegama ST^{1*}, Ranaweera KKDS², Sarananda HS³, Wijesundera BLDLD¹, De Silva RN¹

¹Fruit Research & Development Institute, Department of Agriculture, Sri Lanka

²Department of Applied Science, University of Sri Jayewardenepura, Sri Lanka

³Faculty of Agriculture and Plant Management, Wayamba University of Sri Lanka

Received: 12 December 2023, Accepted: 28 May 2024, Published: 30 June 2024

ABSTRACT

The present experiment was conducted at the Fruit Research and Development Institute Horana, Sri Lanka, from 2020 to 2021 to evaluate the proximate composition, antioxidant activity, ascorbic acid and total phenolic content of five underutilized fruits, namely Namnam, Lovi, Roseapple, Pumello and Sapota using Association of Official Agricultural Chemists (AOAC) methods. The parameters determined were moisture, crude fiber, crude fat, sugar, antioxidant activity, total phenolic content, titratable acidity, pH, total soluble solids and ascorbic acid. Rose apple and Namnam fruits recorded the highest (91.47%) and the lowest (68.87%) moisture content, respectively. The Pumello fruit exhibited the lowest fat, crude fiber, total sugar, non-reducing sugar and reducing sugar contents with the values of 0.01%, 1.33%, 0.05%, 0.03%, and 0.02%, respectively. Sapota reported the highest fat, total soluble solids and pH values of 1.33%, 16%, and 4.74%, respectively. Roseapple recorded the highest crude fiber (5.59%) and the total sugar content (0.81%), while Lovi showed the lowest pH (2.29) and highest titratable acid content (2.01%). Pumello had the highest ascorbic acid concentration, whereas Lovi had the lowest value. Pumello and Namnam demonstrated the lowest IC₅₀ value (0.03). Sapota and Roseapple had the highest (20.49) and lowest (2.16) total phenolic content. The study emphasizes the diverse nutritional elements in these underutilized fruits, highlighting their potential as valuable sources of supplementary nutrients.

Keywords: Antioxidant activity, ascorbic acid, moisture, phenolic content, underutilized fruits.

INTRODUCTION

Fruits contain vitamins and minerals in large quantities (Khan *et al.* 2021) and many other beneficial compounds required for human health (Ibrahim *et al.* 2017). In recent years, increasing attention has been paid to studying the role of fruits and vegetables in diet and human health. Fruits and vegetables have a direct correlation with healthy lifestyles, and higher fruit and vegetable consumption is related to a healthy dietary pattern (Dhendevi and Jeewon 2015). Frequent consumption of

fruits and vegetables has been associated with a lowered risk of diabetes, hypertension, coronary heart disease, cancer and strokes (Ibrahim *et al.* 2017).

Fruits and vegetables are very good sources of antioxidants (Ravimannan and Nisansala 2017). Antioxidants are substances that protect human cells from damage caused by free radicals such as superoxide, peroxy, and hydroxyl radicals. Antioxidants can be naturally obtained from fruits, vegetables, nuts, legumes, grains, and cereals. Fruits

Corresponding author: samanthafirdi@gmail.com

exhibit different antioxidant activities due to the presence of various types of antioxidants (Conneorly 2008).

Fruits and vegetables contain not only high levels of antioxidants, but also nutrients such as moisture, ash, fat, protein and carbohydrate. The moisture content of the fruit plays a vital role in determining its nutritional level, shelf-life, microbial stability and quality. Whereas ash content means the total amount of minerals (Sadulech *et al.* 2012 and Untalan 2015). Crude fiber content is the estimate of the indigestible fiber in food. Fruits give carbohydrates in the form of soluble sugars and cellulose and serve as a source of nutrients. A few underutilized fruits are rich in antioxidants and phytochemicals, besides necessary nutritional compounds such as vitamins, minerals and dietary fiber (Mallawaarachchi *et al.* 2015). According to the World Health Organization (WHO), the daily requirement for fruits is 200 g per day, but this requirement is not fulfilled by most people in Sri Lanka. There are a lot of underutilized fruits available in Sri Lanka (Ranawake, 2021). Those fruits can be used for the fulfilment of nutritional deficiency problems. Recent studies have suggested that underutilized fruits, such as cancer-fighting antioxidants and phenolic compounds, possess many health benefits.

Underutilized resources in Sri Lanka, including fruit crops, play a crucial role as lifelines for poor individuals in regions facing significant challenges in food and nutrition security and ayurvedic medicine (Ranawake and Pathirana, 2024). These fruits offer a solution to nutritional deficiencies. The levels of phytochemicals and macronutrients in these crops can be influenced by varieties, climatic conditions, cultural practices, and maturity at harvest and storage conditions (Tiwari *et al.* 2013). Proximate analysis, which includes assessing nutrients like moisture, ash, crude fiber, crude fat, crude protein and carbohydrates is essential for understanding the nutritional composition (Untalan *et al.* 2015). Additionally, physicochemical characteristics serve as vital, qualitative indicators for the fresh consumption of fruits (Zaman *et al.* 2006).

However, detailed studies on underutilized local fruits to establish their usefulness as fruit crops are limited with special reference to their nutrient content and phytochemical compounds are not available. Therefore, the main objective of the present study was to determine the antioxidant activity, total phenolic content ascorbic acid content and proximate composition (moisture, ash, crude fat, sugar and crude fiber) of five locally available underutilized fruits.

MATERIALS AND METHODS

Chemicals and equipment

All the chemicals used in the study were pure analytical-grade materials. The chemicals include gallic acid, 2,2-diphenyl-1-picrylhydrazyl (DPPH), quercetin and folin phenol reagent, Fehling's solution A and B, Sulphuric acid, NaOH, Na₂CO₃ and other required reagents, which were purchased from Sigma Aldrich USA. Experiments were designed with a complete randomized design (CRD) with three replicates.

Samples

Five underutilized fruits were analyzed for their proximate composition and antioxidant properties. The English name "Lovi" corresponds to the local name "Lovi," with the scientific name *Flacourita inermis* and belonging to the family Flacourtiaceae. The English name "Roseapple" is known locally as "Jambu," with the scientific name *Syzygium jambos* and is part of the Myrtaceae family. "Namnam" is locally called "Nami-nam," with the scientific name *Cynometra cauliflora* and falls under the Fabaceae family. The fruit "Pumello" is known locally as "Jambola," with the scientific name *Citrus maxima* and is in the Rutaceae family. Finally, "Sapota" is locally called "Sapadilla," with the scientific name *Manilkara zapota* and belongs to the Sapotaceae family.

Samples of underutilized fruits were collected from their natural habitats (fruits are naturally grown without introduction) in well-grown areas in wet, intermediate, and dry zones. Mature ripe fruits without visible damage were selected as samples at the edible stage. The collected fruits were cleaned and washed

with tap water to remove dirt and adherent particles. Subsequently, the fruits were kept in a clean, dry place until further use.

Sample preparation for analysis.

The seeds were removed from the selected fruits, and for the Pummelo fruit the peel was also removed. The remaining parts were cut into small pieces and homogenized using a blender. (The number of fruits depends on the size of fruits and sample weights; fruits are selected from a few trees which are in the same location and conditions).

This study used one hundred fruits of Lovi, Naminam, and Roseapple from three trees, fifty fruits from three trees of Sapota, and ten fruits of Pummelo from three trees.

Analytical methods

Determination of moisture

Moisture content was determined as prescribed by (Ranganna, 2010) AOAC for fruits. It was determined by heating 5 g of each sample to a constant weight in an air oven maintained at 105° C. This process involved heating, cooling in a desiccator, and measuring the weight, until a constant weight gain was achieved. Moisture was determined in triplicate using the following formula.

Moisture % =

$$\frac{\text{fresh weight(g)} - \text{dry weight(g)}}{\text{fresh weight(g)}} \times 100$$

Determination of titratable acidity

Titratable acidity was determined using AOAC method (Ranganna, 2010). The sample was titrated against 0.1N NaOH solution using phenolphthalein as an indicator. The acidity was calculated using the following formula. The three replicates and the average readings were reported.

Titratable acidity (%) =

$$\frac{\text{titre} \times \text{normality} \times \text{volume made up (ml)} \times \text{equivalent weight} \times 100}{\text{volume of extract (ml)} \times \text{weight of the sample (g)}}$$

Determination of total soluble solids (TSS)

The TSS of pulps was determined using a

hand-held Refractometer. A drop of juice sample was squeezed onto the surface of the refractometer's prism, and the percentage of total soluble solids was recorded directly. TSS was measured at room temperature (Ranganna, 2010).

Determination of crude fat

The dried sample in a thimble was covered with fat-free cotton and placed in a soxhlet apparatus. The flask was filled with 150 ml petroleum ether and extraction was carried out for 16 hours. The sample was then dried at 100°C in an oven for 1 hour, cooled to room temperature, and weighed. The difference in the weights gave the fat-soluble material present in the sample. Determinations were conducted in triplicate, and the averages were recorded (Ranganna, 2007).

Crude fat% =

$$\frac{\text{weight of flask + oil (g)} - \text{weight of flask (g)}}{\text{weight of sample (g)}} \times 100$$

Determination of crude fiber

The crude fibre was determined from the residue after the crude fat determination. Boiling sulphuric acid (200 ml) was added to the 2 g of the residue in a digestion flask and heated for 30 minutes. The wetted material was then filtered and washed thoroughly with boiling water until the washing was no longer acidic. A NaOH solution was added to the washed material, and the mixture was boiled under reflux for 30 minutes. The material was filtered and washed thoroughly with water, followed by 15 ml of alcohol. The contents were dried at 110° C to constant weight. The material was then burnt in the muffle furnace at a dull red heat at 550°C for 20 minutes, then cooled and weighed. The loss in weight represented the crude fiber content (Ranganna, 2010).

Crude fibre% =

$$\frac{\text{weight of crucible with fiber (g)} - \text{weight of crucible with ash (g)}}{\text{weight of sample (g)}} \times 100$$

Determination of reducing sugar content

The method described by Ahmmed (2015), with some modifications, was performed to determine the reducing sugar. Five grams of

homogenized sample was taken in a 250 ml conical flask, and water was added to make the volume up to 100 ml. The solution was stirred well until all the soluble matters were dissolved and then filtered through filter paper (Whatman no 01). The filtrate was used for analysis, 10 ml of mixed Fehling's solution (1:1) was taken into a conical flask and titrated against the filtrate sample using methylene blue as an indicator. The end point of titration was a brick red colour.

Reducing sugar % =

$$\frac{\text{Factor} \times \text{volume made up}}{\text{titer volume} \times \text{weight of the sample}} \times 100$$

Determination of total sugar

Total sugar was determined following the same procedure as the reducing sugar estimation. The sample was taken into a conical flask and 10 ml of diluted (1:1) HCL was added. The mixture was boiled for five minutes and then cooled. The sample was neutralized using 20% NaOH with phenolphthalein as an indicator, and the volume was made up to 250 ml in a volumetric flask. Ten milli litres of Fehling's solution (A & B) was taken into a conical flask, and two drops of methylene blue were added as an indicator. The solution was titrated against the sample, and the end point of the titration was brick red.

Calculation:

Total sugar % =

$$\frac{4 \times \text{factor} \times \text{volume made up} \times 100}{\text{titer volume} \times \text{weight of sample} \times 100}$$

Non-reducing sugar content

Non-reducing sugar =

Total sugar – Reducing sugar

Determination of ash

Five grams of sample was kept in a muffle furnace and burnt at a temperature not exceeding 500°C for 6 hours. The ash was then cooled in a desiccator and weighed. The ash content was recorded as a g per 100 g fresh weight (g/100 g fw) (AOAC 1990).

Ash % =

$$\frac{\text{weight of ash with crucible (g)} - \text{weight of crucible (g)}}{\text{weight of sample}} \times 100$$

Determination of ascorbic acid

Ascorbic acid content was determined using the titration method involving 2,6-dichloro indophenol dye method described by Sharma (2018) with some modifications.

Ascorbic acid content (mg /100 g) =

$$\frac{(\text{titer} \times \text{dye factor} \times \text{volume made up (ml)}) \times 100}{\text{volume of extract (ml)} \times \text{weight of the sample (g)}}$$

Determination of total phenolic content

Total phenolic content was determined using the folin ciocalteu method (Yu *et al.* 2002) and TPC was expressed as mg of Gallic acid equivalents (GAE) per gram of fresh weight.

Determination of antioxidant activity

Antioxidant activity and IC₅₀ values were determined using 2, 2-diphenyl -1-picrylhydrazyl (DPPH) assay described by Su *et al.* (2007).

Antioxidant activity % =

$$\frac{(1 - \text{absorbance of the sample})}{(\text{absorbance of the control})} \times 100$$

IC₅₀ value was calculated using a concentration vs. antioxidant activity graph.

Data analysis

Data were analyzed using computer-based R software (Kassambara, 2016).

RESULTS AND DISCUSSION

Moisture content

Among the tested fruits, the moisture values ranged from 91.47 % to 68.87%, where Rose apple exhibited the highest moisture content, while Namnam displayed the lowest moisture content (Figure 1). The moisture content in selected fruits is significantly different. The moisture content of fruits depended on climatic conditions, the maturity stage, and the type of fruits. The pulp was very high in moisture content, and this may underscore its

high perishability and susceptibility to microbial infections. This is indicative of low solid matter in the pulp. High moisture content characterizes the freshness of fruit since fruits kept for some time tend to lose moisture (Sri *et al.* 2022). The low moisture content indicated its high dry matter content and possible long shelf-life (Adepoju 2009). Moisture content is one of the properties that are important for nutritional labeling, food quality, value added products and microbial stability.

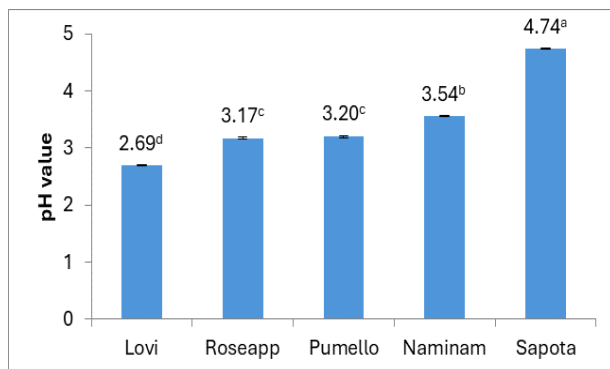


Figure 1: Moisture % of selected underutilized fruits

pH value

Sapota fruit had a high pH (4.74), and Lovi fruit had a lower pH (2.69) (Figure 2). The pulps' pH showed an acidic character and the low pH enhanced microbiological and physicochemical stability. Due to their organic acid content, fruits' pH generally varies between 2.5 and 4.5. The pH values of the tested fruits were within the range of general values.

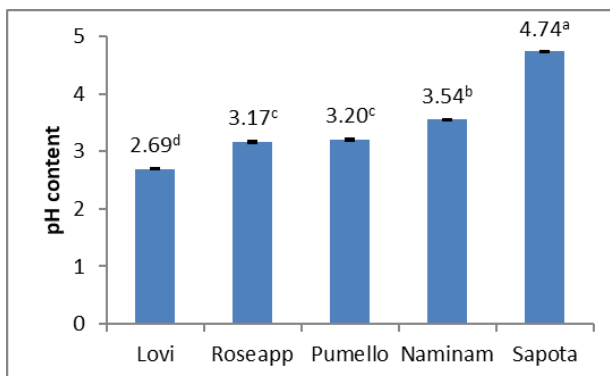


Figure 2: pH Values of selected underutilized fruits.

Titrateable acidity

The tested fruits' titrateable acidity levels ranged from 0.03 % (Sapota) to 2.01 % (Lovi) (Figure 3). The pulp's titrateable acidity, which contributes to the acidity of the aroma, mainly used citric acid as the prominent acid.

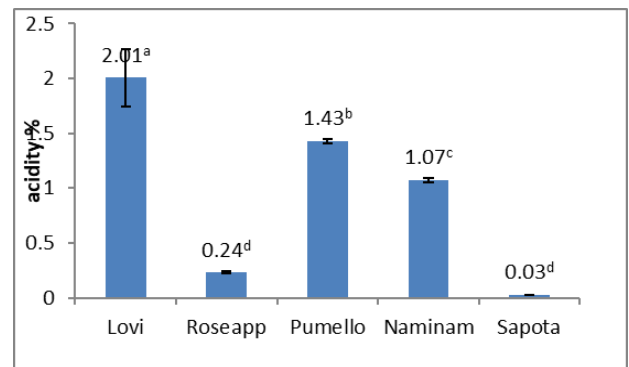


Figure 3: Titrateable acidity of selected underutilized fruit crops

Total soluble solids (TSS)

Results revealed that among the tested fruits, the highest TSS content (brix value) was reported in Sapota (16%) fruit, and the lowest value was shown in Rose apple (3.97%) (Figure 4). Brix is a unit used to describe the percentage of soluble solids in pulp. The soluble solids are mainly sugar, but there are also smaller or larger amounts of acids and other materials (Karunasena *et al.* 2018).

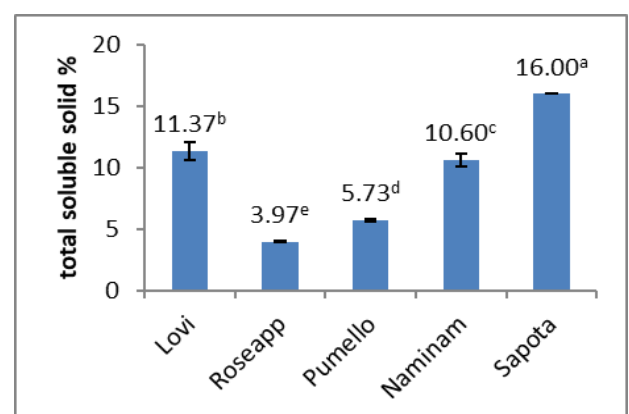


Figure 4: TSS content of selected fruit crops

Crude fat content

The lowest fat content was found in Pumello fruit (0.01%), and the highest was in Sapota fruit (1.33%) (Figure 5).

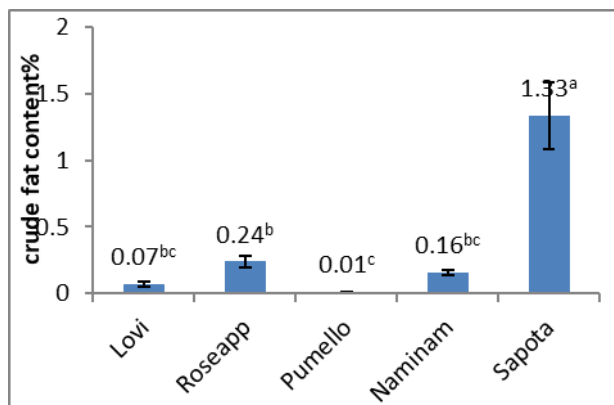


Figure 5: crude fat content of selected underutilized fruit crops

Ash content

The results revealed that the fruit with the highest ash content was Pumelo, with a value of 0.7% (Figure 6). Conversely, the fruit with the lowest ash content was identified as Lovi, with a recorded value of 0.03%. Ash content indicates mineral value, especially macro minerals (Adepoju 2009).

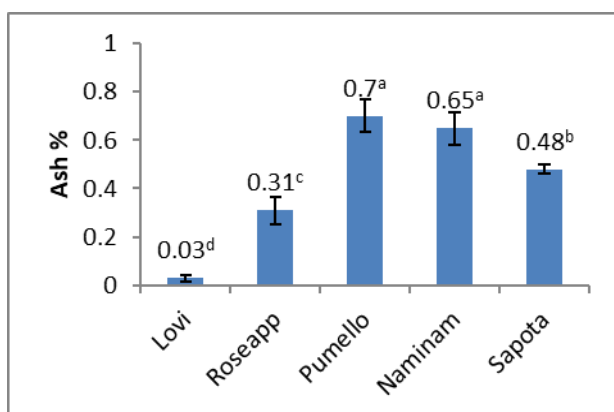


Figure 6: Ash content of selected underutilized fruit crops

Crude fiber content

Crude fiber content ranged from 1.33 g /100 g - 5.59 g /100 g among the tested fruits with the highest crude fiber content determined in

Rose apple fruit and lowest in Pumello fruit (Figure 7). Fiber contents of fruit samples were relatively low (1.5-2.9 g /100 g). Crude fiber is an essential nutrient. Which means it must be eaten in the diet. Crude fiber consists largely of cellulose, hemicellulose (60-80%), lignin (4-6%), and some mineral matters. These fibers are beneficial in treating or preventing constipation, hemorrhoids, diverticulosis, coronary heart disease, and some types of cancer (Madhu *et al.* 2017). Soluble dietary fiber has health-promoting properties as it has been implicated in lowering plasma and liver cholesterol concentrations (Behall and Resier 1986). Fiber helps to maintain the health of the gastrointestinal tract, but in excess, it may bind trace elements, leading to deficiencies in iron and zinc in the body (Siddhuraju *et al.* 1996).

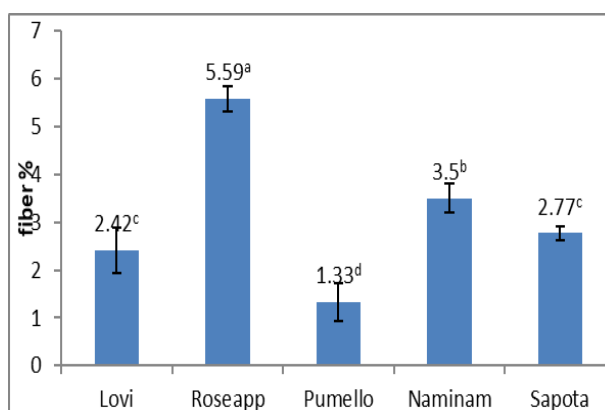


Figure 7: Crude fiber content of selected underutilized fruit crops

Sugar content

Among the tested fruits, the highest reducing sugar (0.18 %), non-reducing sugar (0.65%) and total sugar (0.81%) content were showed in Naminam and Rose apple fruits, respectively, and the lowest levels (0.02%, 0.03 % and 0.05% respectively) were showed in Pumello fruit (Figure 8). The most abundant sugars in fruits are glucose, fructose, and sucrose, which are in various proportions depending on the species. Sugar content, pH data and titratable acidity are essential characteristics indicating the potential for future use of fruits. Sugar content is important to find a good balance between pH, sugar and

titratable acidity to receive an optimal taste (Magaia *et al.* 2013).

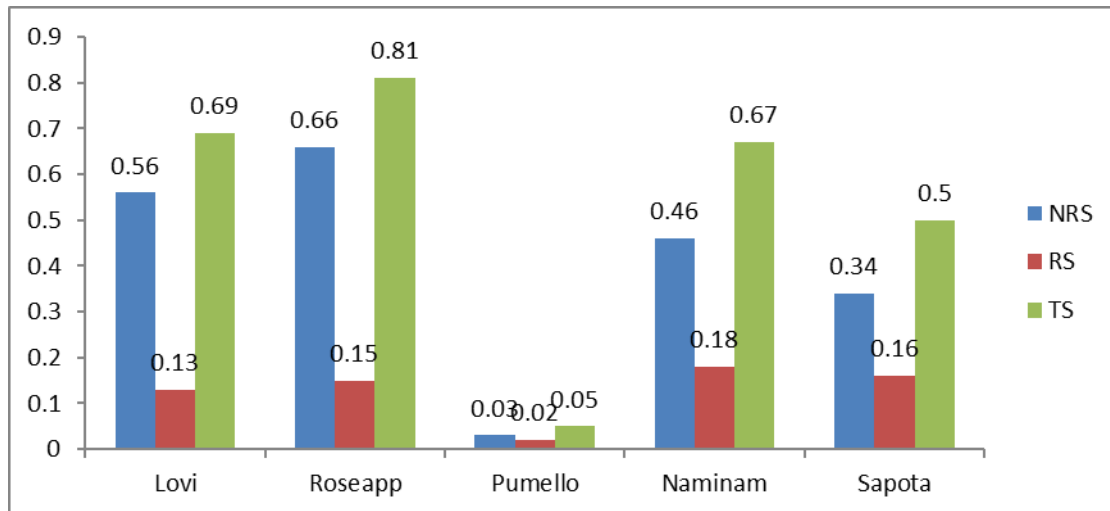


Figure 8: Reducing sugar, non-reducing sugar and total sugar content of selected underutilized fruits TS – Total sugar content RS- Reducing sugar content NRS- non-reducing sugar content

Ascorbic acid content

Pumello fruit yielded the highest ascorbic acid content (330 mg/ 100 g) among the selected fruits, while Lovi fruit had the lowest (58 mg/ 100 g) (Figure 9). These results indicate that Naminam, Pumello, and Sapota can provide more vitamin C than other selected fruits. Ascorbic acids, also known as vitamin C or L-ascorbic acid or antiscorbutic vitamin, are supplied by fruits and vegetables. More than 90% of the vitamin C in human diets is supplied by fruits and vegetables (Karunasena *et al.* 2018).

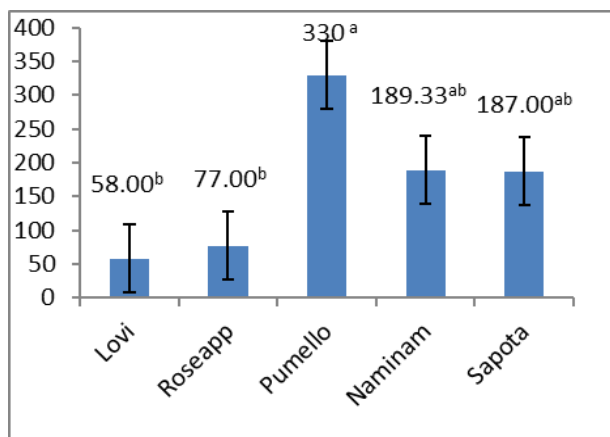


Figure 9: Ascorbic acid content (mg/100g) of selected underutilized fruit crops

Phenolic content

The total phenolic content of tested fruits ranges from 2.61 to 20.49. the highest value in the Sapota fruit and the lowest value in the Namnam fruit (Figure 10). Several factors could be added to be responsible for differences in the total phenol content of foodstuffs of the same or similar origin. They include variations in cultivars, cultivation practices, harvest and post-harvest handling, storage conditions, processing techniques during analytical determination, location, growing season, and maturity stage of fruits (Kubola *et al.* 2011; Pearson *et al.* 1999).

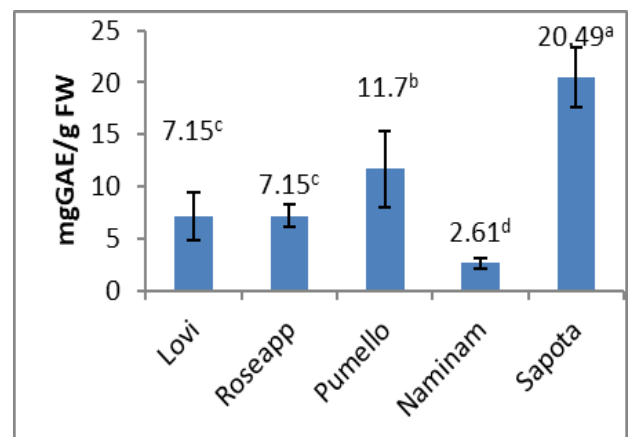


Figure 10: Phenolic content (mg GAE/G) of selected underutilized fruit crops

Radical scavenging capacity

Results are expressed as IC₅₀, which indicates the concentration of the sample required (g of fresh weight /ml) to scavenge 50 % of DPPH radicals in the reaction medium. Among the tested samples, the highest IC 50 value was observed in Sapota (0.069 g of Fw/ml), while the lowest value was recorded in Namnam fruit (0.031 g of Fw/ml), which indicates that a low concentration of Namnam fruit is required to scavenge 50% of free radicals (Figure 11).

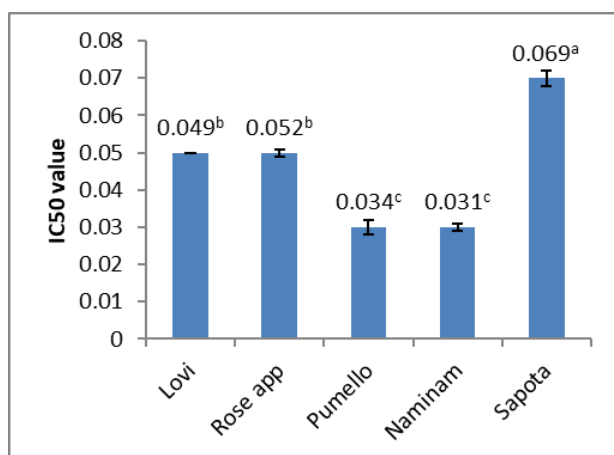


Figure 11: IC 50 value (g fw/ml) of selected underutilized fruit crops

The previous study (Mallawaarachchi *et al.* 2021) shows the IC 50 value of Namnam fruit on a dry weight basis was 0.47 g DW/ml. Dry weight basis accounts for the removal of water from the sample, providing a more concentrated measure of the active components; dry weight basis IC₅₀ is a measure of the concentration required to scavenge 50% of radicals in the absence of water, difference between two values may be a variation of the water content of the sample.

CONCLUSIONS

Namnam, Pumello, and Sapota fruits recorded the highest ascorbic acid content, the lowest IC 50 value (high in antioxidants), and the highest phenolic content, implying that they are rich in antioxidants. Rose apple fruit has high moisture content and low dry matter content, proving its perishable nature with a short shelf life. Lovi fruit has a high titratable

acidity content and low pH value. Pumello fruit has a high ash content, indicating a higher mineral content.

These underutilized fruits can be used to fulfill nutritional requirements and provide low-cost alternatives for resource-poor people.

ACKNOWLEDGEMENTS

NARP provided financial support for the research, the Director of Fruit Research and Development Institute provided facilities to carry out our experiments and all the others who supported the research.

AUTHOR CONTRIBUTION

BST designed the study, conducted experiments and data analysis and wrote the manuscript. RKKDS and SHS guided the study and carried out experiments and DSRN and WBLDL carried out experiments.

REFERENCES

- Adepoju TO 2009 Proximate composition and micronutrient potentials of three locally available wild fruits in Nigeria. African journal of agriculture research 4 (9): 887-892.
- Behall K, Resier 1986 Chemistry and function of pectin in ACS system symposium series 310.
- Connealy LE 2008 The importance of antioxidants in fruits and vegetables. Natural news, URL:<http://goo.gl/9UN60V>.
- Dhendevi PEM and Jeewon R 2015 Fruit and vegetable intake benefits and progress of nutrition education intervention-Narrative review article. Iranian journal of public health. 44(10): 1309-1321.
- Holloway DW 1983 Composition of fruit, vegetable and cereal dietary fiber. Journal of Science food agric. 34: 1236-1240.
- Ibrahim UK, Kamarrudin N, Suzihaque MUH and Hashib SA 2017 Local fruit waste as a potential source of natural antioxidants. An overview IOP conference series. Material science and engineering 206: 012940.

- Kumbhar SD, Kulwal PL, Patil JV, Sarawate CD, Gaikwad AP & Jadhav AS 2015 Genetic diversity and population structure in landraces and improved rice varieties from India. *Rice Science* 22(3): 99-107.
- Karunasena GADV, Chandrajith VGG, Nawaratne SB 2018 Physicochemical characteristics of peanut butter fruits (*Bunchosia armeniaca*). *International Journal of Food Science and Nutrition* 3(3): 46-51.
- Khan S, Arya R, Pooja Babli M, Pahive and Singh IA 2021 Review paper on fruit nutrition and health benefits, *The Pharma Innovation Journal*, 10(6): 119-123.
- Kubola J, Siriamornpun S and Meeso N, 2011 Phytochemicals, vitamin C and sugar content of Thai wild fruits. *Food Chemistry*, 126: 972-981.
- Luximon RA, Bahorun T and Crozier A, 2003 Antioxidant action and phenolic and vitamin C contents of common mauritian exotic fruits, *Journal of the Science of Food and Agriculture*. 83: 496-502.
- Madhu C, Krishma KL, Reddy KR, Lakshmi PJ 2017 Estimation of crude fiber content from natural food stuffs and its laxative activity induced in rats. *International Journal of Pharma Research and Health Sciences* 5 (3): 1703-06.
- Magaia T, Uamusse A, Sjöholm I, Shog K 2013 Proximate analysis of five wild fruits of Mazambique. *The scientific world journal* Article ID 601435, 6 pages. Research article hindawi.
- Mallawaarachchi MALN, Dissanayaka KSV and Madhujith T 2015 Antioxidant potential of selected underutilized fruit species grown in Sri Lanka. *proc 8th International Research Conference* 103-106, Kotalawala Defence University Sri Lanka.
- Mallawaarachchi MALN, Madhujith T, Suriyagoda LDB and Pushpakumara DKN 2021 Antioxidant efficacy of selected underutilized fruit species grown in Sri Lanka. *Tropical Agricultural Research* 32(1): 68-80.
- Othman OC 2009 Physical and chemical composition of storage ripened papaya (*Carica papaya* L.) fruit of Eastern Tanzania. *Tanzania Journal of Science*, 35: 47-56.
- Peorson DA, Tan CH, German B, Davis PA, Gerahwin ME 1999 Apple juice inhibits human low density lipoprotein oxidation. *Life Science* 64: 1913-1920.
- Ranawake AL 2021 The underutilized resources in the lowland wet zone forests of Sri Lanka and untapped indigenous knowledge of peripheral households. *Journal of University of Ruhuna* 9:47-71.
- Ranawake AL and Pathirana R 2024 Indigenous knowledge in the Kithul (*Caryota urens* L.) industry of Sri Lanka and its scientific basis. *Genetic Resources and Crop Evolution*, pp.1-20.
- Ranganna S 2007 Handbook of analysis and quality control fruit and vegetable products. (2nd ed) New Delhi, Tata McGraw Hill pub co. Ltd.
- Ranganna S 2010 Handbook of analysis and quality control for fruit and vegetable products. Tata Mc Graw, Hill Education.
- Ravimannan N and Nisansala A 2017 Study on antioxidant activity in fruits and vegetable, a review. *International Journal of Advanced Research in Biological Science*, 4(3): 93-101.
- Saka JDK and Msonthi JD 1994 Nutritional value of edible fruits of indigenous wild trees in Malawi. *Forest Ecology and Management*, (2-3): 245-248.
- Sharma HP, Patel H, Sharma S and Vaishali 2018 Study of physico-chemical changes during woodapple (*Feronia limonia*) maturation. *Journal of Food Research and Technology* 2 (4): 148-152.
- Siddhuraju P, Vijayakumari K, Janardhanan K 1996 Chemical composition and protein quality of the little known legume, velvet bean (*Mucuna pruriens* (a) DC). *Journal of Agriculture and Food Chemistry* 44: 2636- 2641.
- Su L, Yin JJ, Charles D, Zhon K, Moore J and Yu L 2007 Total phenolic contents

chelating capacities radical scavenging properties of black peppercorn, nutmeg, rosehip, cinnamon and oregano leaf. *Journal of Food Chemistry* 100: 990-997.

Sri AN, Subramanyam AKM, Kumari R. Sai D and Venkateswarla G 2022 Post-harvest handling to reduce loss of fruit and vegetables. *The Pharma Innovation Journal* sp 11(6): 1873-1879.

Tiwari U, Cummins E 2013 Factors influencing levels of phytochemicals in selected fruit and vegetables during pre and post-harvest food processing operations. *Food Research International* 50(2): 497-506.

Tressler DK, Arsdell VWB, Copley MJ, 1980 *The freezing preservation of foods*. 4th Edn.23 AVI Publishing, Co, Westport, Conn.

Untalan KMC, Perez IFR, Reyes GH, Escalona KMH, Guzman LD, Lumanglas RFL 2015 Proximate Analysis and Antioxidant Properties of Selected Fruits in Batangas. *Asia Pacific Journal of Multidisciplinary Research*, 3(4): 41-45.

Yu L, Perret J and Davy B, Wilson J and Melby CL 2002 Antioxidant properties of cereal products. *Journal of Food Science* 67 (7): 2600-2603.

Zaman W, Biswas SK, Helali MOH, Ibrahim M and Hassan P 2006 Physicochemical composition of four papaya varieties grown at Rajshahi. *Journal of Bioscience* 14: 83- 86.