

SHARED PATHWAYS TO PEPTIDES IN BIOCHEMISTRY AND PREBIOTIC CHEMISTRY THROUGH ESTER-AMIDE EXCHANGE CHEMISTRY

Yishi Ezerzer¹, Disha-Gajanan Hiregange², Anat Bashan^{2,*}, Ada Yonath^{2,*}, Moran Frenkel-Pinter^{1,3,4,*}

¹Institute of Chemistry, The Hebrew University of Jerusalem, Jerusalem 9190401, Israel

²Department of Chemical and Structural Biology, Weizmann Institute of Science, Rehovot 7610001, Israel

³The Center for Nanoscience and Nanotechnology, The Hebrew University of Jerusalem, Jerusalem 9190401, Israel

⁴Casali Center of Applied Chemistry, The Hebrew University of Jerusalem, Jerusalem 9190401, Israel

Abstract

The origin of life requires plausible pathways for the emergence of polymers under realistic early-Earth conditions. Increasing geological and geochemical evidence supports highly dynamic environments characterized by strong temperature gradients and fluctuating hydration states that repeatedly drive chemistry out of equilibrium. Wet-dry cycling in shallow settings can promote polymer formation by favoring condensation during dehydration and hydrolysis during rehydration, thereby imposing a rudimentary, persistence-based selection pressure. Here we highlight a conceptual chemical bridge between prebiotic polymer chemistry and extant biochemistry: ester-mediated amide formation via ester-amide exchange. In modern translation, peptide bond formation proceeds not by direct condensation of amino acids but by nucleophilic attack of the α -amine of the A-site aminoacyl-tRNA on the ester linkage of the P-site peptidyl-tRNA at the ribosomal peptidyl transferase center. Strikingly similar chemistry arises abiotically under wet-dry cycling in mixtures of hydroxy acids and amino acids, where dehydration readily generates esters that subsequently undergo ester-amide exchange to form depsipeptides, which are oligomers composed of mixed ester and amide linkages, without sophisticated catalysts. We argue that wet-dry cycling and ester-mediated exchange may outline a plausible continuum from non-coded depsipeptide formation to the refined ribosomal peptide synthesis, offering an experimentally tractable framework for interrogating the constraints that shaped early polymer evolution.

Keywords

origins of life • chemical evolution • wet-dry cycling • peptide evolution • protoribosome

Few scientific questions are as profound as the origin of life. One of the biggest mysteries in origin of life research concerns the emergence of today's biopolymers. Unraveling this question requires careful consideration of early Earth environments. Geological and geochemical evidence points toward highly dynamic settings, characterized by strong temperature gradients and fluctuating hydration states, which are continuously driving chemical systems out of equilibrium (1–8). Such non-equilibrium environments force molecular systems to explore broader chemical space, favoring reactions and pathways that would be hindered under constant, fully aqueous conditions (9–11).

Wet-dry cycling is one of the key non-equilibrium processes proposed to promote molecular complexation on early Earth (9, 12–20). In shallow aqueous environments, such as small ponds or surface pools, repeated evaporation and rehydration act as a simple yet powerful physical driver of chemical evolution (21). During drying phases, condensation-dehydration reactions are favored, whereas

during rehydration, more labile products are selectively hydrolyzed. This constant and repeated pressure introduces a rudimentary form of selection, favoring molecular structures that can persist and accumulate over prolonged timescales.

Peptide bond formation has been demonstrated via several mechanisms under plausible prebiotic conditions (22–24). Despite their central role in modern biology, amino acids do not readily form peptide bonds in aqueous environments; peptide bond formation is thermodynamically disfavored by competition with hydrolysis and kinetically limited by the high activation energy required for amide bond formation. These limitations have led to the proposal of numerous pathways invoking solutions such as activated monomers, catalysts, or constrained environmental reaction conditions.

A conceptually elegant solution to peptide bond formation emerges by considering peptide formation in environments undergoing wet-dry cycling, proceeding through an exchange reaction rather than a direct condensation between two amino

* Corresponding authors e-mail: moran.fp@mail.huji.ac.il; ada.yonath@weizmann.ac.il; anat.bashan@weizmann.ac.il

acid monomers. Modern biology itself relies on this strategy: ribosomal peptide synthesis proceeds through activated ester intermediates rather than direct amide bond formation (25). Examination of the ribosomal peptidyl transferase centre (PTC) reveals two functionally distinct sites: the P site and the A site (26–30). The P site holds the growing peptide chain esterified to a transfer RNA (tRNA), while the A site accommodates an incoming aminoacyl-tRNA poised for incorporation into the elongating peptide. Notably, peptide bond formation in this context does not proceed via a direct condensation-dehydration reaction. Instead, elongation occurs through an ester-amide exchange reaction, in which the amine of the amino acid in the A site nucleophilically attacks the ester linkage between the peptide and the tRNA in the P-site (31). These chemical parallels are consistent with the protoribosome hypothesis, which proposes that a primitive RNA-based catalytic scaffold was capable of promoting peptide bond formation before the advent of genetically encoded translation (26, 27, 31). Support for this view comes from structural evolutionary analyses of the ribosome (25, 32–34). These studies suggest that the ribosome evolved in an accretionary, ‘onion-like’ manner, in which new structural layers were added over time without substantial loss or rearrangement of the ancestral core. The peptidyl transferase center, located at the heart of the ribosome, appears to be among the most ancient components, with progressively younger structural and functional elements radiating outward. This layered architecture preserves a molecular record of evolutionary history, offering a plausible timeline by which a simple RNA-based catalytic scaffold could have gradually acquired increasing complexity while retaining its original chemical function.

Importantly, such a protoribosomal system would not require sequence specificity or templated information transfer, but only the spatial organization of activated substrates. Structural and mechanistic analyses of the modern ribosomal peptidyl transferase center suggest that it functions primarily as an entropic and geometric catalyst rather than a chemically active enzyme, relying on precise positioning of reactants to facilitate ester-amide exchange. This observation raises the possibility that a much simpler assembly, lacking coded specificity yet capable of binding activated substrates, could plausibly have supported early peptide elongation (and possibly elongation of other proto-polymers). In this view, the ribosome is not an abrupt evolutionary innovation but the refined descendant of a prebiotic chemical system that already exploited ester-mediated peptide formation.

A similar chemical trajectory to peptide bond formation in the ribosome can be recapitulated in reactions of mixtures of hydroxy acids and amino acids under wet-dry cycling conditions. During dehydration, hydroxy acids readily form

esters via condensation-dehydration reactions (13, 35–39). In the presence of amino acids, subsequent ester-amide exchange reactions generate depsipeptides, which are heterogeneous polymers containing both ester and amide linkages (16, 17, 40–48). Notably, this process requires no sophisticated catalysts (49) and emerges naturally from the environmental cycling of prebiotically available building blocks (hydroxy acids and amino acids) (50–54).

Furthermore, within these fluctuating wet-dry environments, primitive RNA/pre-RNA oligomers or RNA-peptide assemblies capable of transient substrate binding could have further biased ester-amide exchange reactions toward peptide formation. Rather than merely driving peptide bond formation, wet-dry cycling functions as a chemical filter that selects for certain molecular traits. Across repeated wet-dry cycles, a gradual enrichment of peptide bonds relative to ester bonds has been observed, consistent with the greater resistance of amide linkages to hydrolytic stress compared with ester bonds (17). In this regime, the environment itself biases the system toward peptide-rich polymers. Hence, mere chemical stability, rather than biological function, emerges as an early dominant selective pressure shaping polymer composition.

While plausible routes to peptide formation have been explored, several key questions remain unresolved. Most prominent among these are - why biology converged on a limited subset of 20 α -amino acids, how amino acid homochirality might have emerged, and how early polymers gradually acquired increased structural and functional complexity (55–62). In this regard, several studies have highlighted depsipeptides as experimentally tractable models for early chemical evolution of peptides. For example, Cronin group has demonstrated that recursive wet-dry cycling of hydroxy acids and amino acids, resulting in depsipeptide formation, can give rise to emergent function. In that work, esterase activity arose only after repeated wet-dry cycles, underscoring the importance of environmental recursion and non-equilibrium conditions for functional emergence, in contrast to static reaction conditions. Moreover, work by Williams, Leman, Hud, and colleagues has shown the selective incorporation of proteinogenic over non-proteinogenic cationic amino acids into depsipeptides under drying conditions, providing evidence that modern biochemical building blocks could have been enriched through purely chemical selection mechanisms (44). As another example, a recent work from our group revealed a selection mechanism favoring α -backbone over β -backbone architectures in depsipeptide libraries, driven by their propensity to assemble into stable droplets via liquid-liquid phase separation (63). This finding suggests that basic physicochemical constraints may have biased early polymer composition. In addition, Hud and colleagues explored depsipeptide nucleic acids as

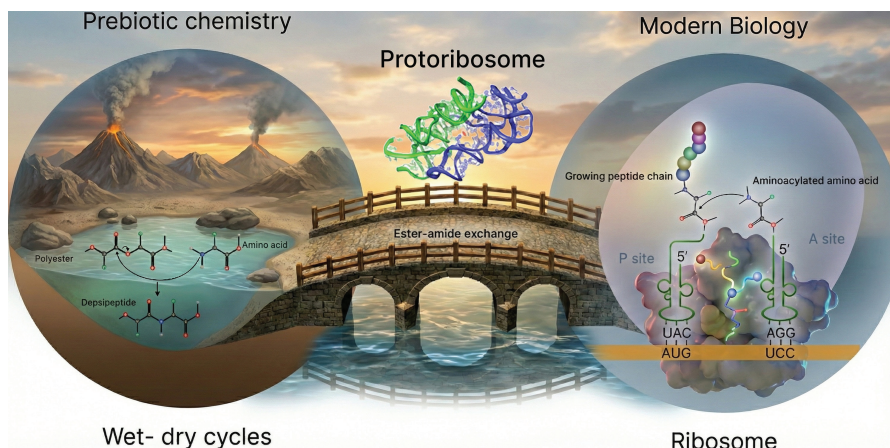


Figure 1. Conceptual link between prebiotic chemistry and modern biology. Ester–amide exchange reactions occurring in a small aqueous pond enable peptide formation, paralleled by an analogous ester–amide exchange reaction in the ribosome that elongates the peptide chain during protein synthesis – all connected by the chemical evolution of a protoribosome complex.

potential prebiotic alternatives to RNA, demonstrating that ester-amide-based backbones can support base pairing and molecular recognition, thereby expanding the landscape of plausible early informational polymers (64).

Overall, wet-dry cycling provides an experimentally tractable and conceptually coherent framework for exploring the prebiotic formation of polymers such as peptides and peptide analogs. This platform has also been utilized for the formation of other molecules, such as polyesters (36, 37), nucleic acids (64), amino acid-keto acid conjugates (65), and oligosaccharides (66). Altogether, integrating insights from modern biology with prebiotic chemistry allows us to probe not only the origins of life, but also the chemical and physical constraints required for life to emerge (67–69). From this perspective, the emergence of the ribosome can be understood not as the origin of peptide synthesis, but as its refinement. Environmental wet-dry cycling, ester-mediated exchange chemistry, and simple catalytic scaffolds together define a continuum in which early, non-coded peptide formation gradually transitioned toward the highly regulated translational machinery observed today (Figure 1). The protoribosome concept thus provides a conceptual link between prebiotic polymer chemistry and extant biology, reinforcing the idea that modern biochemical complexity arose through incremental stabilization and organization of pre-existing chemical processes.

Although the precise historical sequence by which life emerged on Earth 4 billion years ago will remain inaccessible to direct reconstruction, contemporary studies outline chemically and physically plausible pathways consistent with early terrestrial conditions. In this context, the systematic interrogation of extant biochemistry serves not merely as a

retrospective inquiry into our evolutionary origins, but as a means of identifying the underlying constraints, regularities, and organizing principles that render the emergence of living systems possible. Such work reframes the problem of life's origin from a singular historical event to a question about the fundamental laws and boundary conditions under which matter can transition into self-sustaining, evolving complexity.

References

1. Airapetian VS, Glöckner A, Gronoff G, Hébrard E, Danchi W. Prebiotic chemistry and atmospheric warming of early Earth by an active young Sun. *Nature Geoscience*. 2016;9(6): 452–455. doi: 10.1038/ngeo2719
2. Bada JL. New insights into prebiotic chemistry from Stanley Miller's spark discharge experiments. *Chemical Society Reviews*. 2013;42(5): 2186. doi: 10.1039/c3cs35433d
3. Cleaves H. Prebiotic chemistry: geochemical context and reaction screening. *Life*. 2013;3(2): 331–345. doi: 10.3390/life3020331
4. Islam S, Powner MW. Prebiotic systems chemistry: complexity overcoming clutter. *Chem*. 2017;2(4): 470–501. doi: 10.1016/j.chempr.2017.03.001
5. Azevedo HS, Perry SL, Korevaar PA, Das D. Complexity emerges from chemistry. *Nature Chemistry*. 2020;12(9): 793–794. doi: 10.1038/s41557-020-0537-x
6. Singh A, Parvin P, Saha B, Das D. Non-equilibrium self-assembly for living matter-like properties. *Nature Reviews Chemistry*. 2024;8(10): 723–740. doi: 10.1038/s41570-024-00640-z
7. Ianeselli A, Salditt A, Mast C, Ercolano B, Kufner CL, Scheu B, et al. Physical non-equilibria for prebiotic nucleic acid chemistry. *Nature Reviews Physics*. 2023;5(3): 185–195. doi: 10.1038/s42254-022-00550-3

8. Schwintek P, Eren E, Mast CB, Braun D. Prebiotic gas flow environment enables isothermal nucleic acid replication. *eLife*. 2025;13: RP100152. doi: 10.7554/eLife.100152
9. Damer B, Deamer D. The hot spring hypothesis for an origin of life. *Astrobiology*. 2020;20(4): 429–452. doi: 10.1089/ast.2019.2045
10. Danger G, Plasson R, Pascal R. Pathways for the formation and evolution of peptides in prebiotic environments. *Chemical Society Reviews*. 2012;41(16): 5416–5429. doi: 10.1039/c2cs35064e
11. Dondi D, Merli D, Zeffiro A. *Photochemistry of the prebiotic atmosphere*. Royal Society of Chemistry; 2013; p.342–359.
12. Ross D, Deamer D. Dry/Wet cycling and the thermodynamics and kinetics of prebiotic polymer synthesis. *Life*. 2016;6(3): 28. doi: 10.3390/life6030028
13. Mamajanov I, Macdonald PJ, Ying J, Duncanson DM, Dowdy GR, Walker CA, et al. Ester formation and hydrolysis during wet-dry cycles: generation of far-from-equilibrium polymers in a model prebiotic reaction. *Macromolecules*. 2014;47(4): 1334–1343. doi: 10.1021/ma402256d
14. Frenkel-Pinter M, Samanta M, Ashkenasy G, Leman LJ. Prebiotic peptides: molecular hubs in the origin of life. *Chemical Reviews*. 2020;120(11): 4707–4765. doi: 10.1021/acs.chemrev.9b00664
15. Edri R, Joshi MP, Frenkel-Pinter M, Hud NV, Keating CD, Leman LJ. From polymerization-enabled folding and assembly to chemical evolution: key processes for emergence of functional polymers in the origin of life. *Astrobiology*. 2025; 15311074251365943. doi: 10.1177/15311074251365943
16. Frenkel-Pinter M, Smith KH, Rivera-Santana VF, Sargon AB, Jacobson KC, et al. Water-based dynamic depsipeptide chemistry: building block recycling and oligomer distribution control using hydration–dehydration cycles. *JACS Au*. 2022;2(6): 1395–1404. doi: 10.1021/jacsau.2c00087
17. Forsythe JG, Yu SS, Mamajanov I, Grover MA, Krishnamurthy R, Fernández FM, et al. Ester-mediated amide bond formation driven by wet-dry cycles: a possible path to polypeptides on the prebiotic earth. *Angewandte Chemie*. 2015;127(34): 10009–10013. doi: 10.1002/ange.201503792
18. Joshi MP, Sawant AA, Rajamani S. Spontaneous emergence of membrane-forming protoamphiphiles from a lipid–amino acid mixture under wet–dry cycles. *Chemical Science*. 2021;12(8): 2970–2978. doi: 10.1039/D0SC05650B
19. Ruiz-Mirazo K, Briones C, De La Escosura A. Prebiotic systems chemistry: new perspectives for the origins of life. *Chemical Reviews*. 2014;114(1): 285–366. doi: 10.1021/cr2004844
20. Yadav M, Kumar R, Krishnamurthy R. Chemistry of abiotic nucleotide synthesis. *Chemical Reviews*. 2020;120(11): 4766–4805. doi: 10.1021/acs.chemrev.9b00546
21. Capera-Aragones P, Matange K, Rajaei V, Pinter Y, Petrov AS, Williams LD, et al. Stringent selection on kinetics of condensation reactions: early steps in chemical evolution. *Physical Chemistry Chemical Physics*. 2026; 28: 7292–7303. doi: 10.1039/D5CP03057A
22. Rode BM. Peptides and the origin of life. *Peptides*. 1999;20(6): 773–786. doi: 10.1016/S0196-9781(99)00062-5
23. Fox SW, Harada K. Thermal copolymerization of amino acids to a product resembling protein. *Science*. 1958;128(3333): 1214. doi: 10.1126/science.128.3333.1214
24. Whitaker D, Powner MW. On the aqueous origins of the condensation polymers of life. *Nature Reviews Chemistry*. 2024;8(11): 817–832. doi: 10.1038/s41570-024-00648-5
25. Bowman JC, Petrov AS, Frenkel-Pinter M, Penev PI, Williams LD. Root of the tree: the significance, evolution, and origins of the ribosome. *Chemical Reviews*. 2020;120(11): 4848–4878. doi: 10.1021/acs.chemrev.9b00742
26. Davidovich C, Belousoff M, Wekselman I, Shapira T, Krupkin M, Zimmerman E, et al. The proto-ribosome: an ancient nanomachine for peptide bond formation. *Israel Journal of Chemistry*. 2010;50(1): 29–35. doi: 10.1002/ijch.201000012
27. Bashan A, Agmon I, Zarivach R, Schlutzenzen F, Harms J, Berisio R, et al. Structural basis of the ribosomal machinery for peptide bond formation, translocation, and nascent chain progression. *Molecular Cell*. 2003;11(1): 91–102. doi: 10.1016/S1097-2765(03)00009-1
28. Bose T, Fridkin G, Davidovich C, Krupkin M, Dinger N, Falkovich AH, et al. Origin of life: protoribosome forms peptide bonds and links RNA and protein dominated worlds. *Nucleic Acids Research*. 2022;50(4): 1815–1828. doi: 10.1093/nar/gkac052
29. Agmon I, Bashan A, Zarivach R, Yonath A. Symmetry at the active site of the ribosome: structural and functional implications. *Biological Chemistry*. 2005;386(9): 833–844. doi: 10.1515/BC.2005.098
30. Schlutzenzen F, Tocilj A, Zarivach R, Harms J, Gluehmann M, Janell D, et al. Structure of functionally activated small ribosomal subunit at 3.3 Å resolution. *Cell*. 2000;102(5): 615–623. doi: 10.1016/S0092-8674(00)00084-2
31. Trobro S, Åqvist J. Mechanism of peptide bond synthesis on the ribosome. *Proceedings of the National Academy of Sciences*. 2005;102(35): 12395–12400. doi: 10.1073/pnas.0504043102
32. Petrov AS, Gulen B, Norris AM, Kovacs NA, Bernier CR, Lanier KA, et al. History of the ribosome and the origin of translation. *Proceedings of the National Academy of Sciences*. 2015;112(50): 15396–15401. doi: 10.1073/pnas.1509761112
33. Hsiao C, Mohan S, Kalahar BK, Williams LD. Peeling the onion: ribosomes are ancient molecular fossils. *Molecular Biology and Evolution*. 2009;26(11): 2415–2425. doi: 10.1093/molbev/msp163
34. Ditzler MA, Petrov AS, Rothschild-Mancinelli B, et al. Co-evolution of RNase P and the ribosome. *Proceedings of the National Academy of Sciences*. 2026.
35. Petrov, AS, Alvarez-Carreño C, Dean Williams L, Ditzler MA. Coevolution of RNase P and the ribosome. *Proceedings of the National Academy of Sciences*. (2026); 123(10): e2518495123; doi: 10.1073/pnas.2518495123

36. Chandru K, Mamajanov I, Cleaves HJ, Jia TZ. Polyesters as a model system for building primitive biologies from non-biological prebiotic chemistry. *Life*. 2020;10(1): 6. doi: 10.3390/life10010006
37. Jia TZ, Chandru K, Hongo Y, Afrin R, Usui T, Myojo K, et al. Membraneless polyester microdroplets as primordial compartments at the origins of life. *Proceedings of the National academy of Sciences of the United States of America*. 2019;116(32): 15830–15835. doi: 10.1073/pnas.1902336116
38. Mamajanov I, Callahan MP, Dworkin JP, Cody GD. Prebiotic alternatives to proteins: structure and function of hyperbranched polyesters. *Origins of Life and Evolution of Biospheres*. 2015;45(1–2): 123–137. doi: 10.1007/s11084-015-9430-9
39. Campbell TD, Febrian R, McCarthy JT, Kleinschmidt HE, Forsythe JG, Bracher PJ. Prebiotic condensation through wet–dry cycling regulated by deliquescence. *Nature Communications*. 2019;10(1): 4508. doi: 10.1038/s41467-019-11834-1
40. Doran D, Abul-Haija YM, Cronin L. Emergence of function and selection from recursively programmed polymerisation reactions in mineral environments. *Angewandte Chemie*. 2019;58(33): 11253–11256. doi: 10.1002/anie.201902287
41. Edri R, Fisher S, Menor-Salvan C, Williams LD, Frenkel-Pinter M. Assembly-driven protection from hydrolysis as key selective force during chemical evolution. *FEBS Letters*. 2023;597(23): 2879–2896. doi: 10.1002/1873-3468.14766
42. Ezerzer Y, Frenkel-Pinter M, Kolodny R, Ben-Tal N. A building blocks perspective on protein emergence and evolution. *Current Opinion in Structural Biology*. 2025;91: 102996. doi: 10.1016/j.sbi.2025.102996
43. Forsythe JG, Petrov AS, Millar WC, Yu SS, Krishnamurthy R, Grover MA, et al. Surveying the sequence diversity of model prebiotic peptides by mass spectrometry. *Proceedings of the National Academy of Sciences*. 2017;114(37): E7652–E9. doi: 10.1073/pnas.1711631114
44. Frenkel-Pinter M, Haynes JW, C M, Petrov AS, Burcar BT, Krishnamurthy R, et al. Selective incorporation of proteinaceous over nonproteinaceous cationic amino acids in model prebiotic oligomerization reactions. *Proceedings of the National academy of Sciences of the United States of America*. 2019;116(33): 16338–16346. doi: 10.1073/pnas.1904849116
45. Frenkel-Pinter M, Jacobson KC, Eskew-Martin J, Forsythe JG, Grover MA, Williams LD, et al. Differential oligomerization of alpha versus beta amino acids and hydroxy acids in abiotic proto-peptide synthesis reactions. *Life*. 2022;12(2): 265. doi: 10.3390/life12020265
46. Frenkel-Pinter M, Sargon AB, Glass JB, Hud NV, Williams LD. Transition metals enhance prebiotic depsipeptide oligomerization reactions involving histidine. *RSC Advances*. 2021;11(6): 3534–3538. doi: 10.1039/D0RA07965K
47. Yu S-S, Krishnamurthy R, Fernández FM, Hud NV, Schork FJ, Grover MA. Kinetics of prebiotic depsipeptide formation from the ester–amide exchange reaction. *Physical Chemistry Chemical Physics*. 2016;18(41): 28441–28450. doi: 10.1039/C6CP05527C
48. Zhang L, Ying J. Amino acid analogues provide multiple plausible pathways to prebiotic peptides. *Journal of the Royal Society Interface: Royal Society Publishing*. 2024;21(214): 20240014. doi: 10.1098/rsif.2024.0014
49. Edri R, Williams LD, Frenkel-Pinter M. From catalysis of evolution to evolution of catalysis. *Accounts of Chemical Research*. 2024;57(21): 3081–3092. doi: 10.1021/acs.accounts.4c00196
50. John R. Cronin, Carleton B. Moore. Amino Acid Analyses of the Murchison, Murray, and Allende Carbonaceous Chondrites. *Science*. 1971;172: 1327–1329. doi: 10.1126/science.172.3990.1327
51. Aponte JC, Elsil JE, Hein JE, Dworkin JP, Glavin DP, McLain HL, et al. Analysis of amino acids, hydroxy acids, and amines in CR chondrites. *Meteoritics and Planetary Science*. 2020;55(11): 2422–2439. doi: 10.1111/maps.13586
52. Chyba C, Sagan C. Endogenous production, exogenous delivery and impact-shock synthesis of organic molecules: an inventory for the origins of life. *Nature*. 1992;355(6356): 125–132. doi: 10.1038/355125a0
53. Zaia DAM, Zaia CTBV, De Santana H. Which amino acids should be used in prebiotic chemistry studies? *Origins of Life and Evolution of Biospheres*. 2008;38(6): 469–488. doi: 10.1007/s11084-008-9150-5
54. Muchowska KB, Varma SJ, Moran J. Nonenzymatic metabolic reactions and life's origins. *Chemical Reviews*. (2020);120(15): 7708–7744. doi: 10.1021/acs.chemrev.0c00191
55. Cleaves HJ II. The origin of the biologically coded amino acids. *Journal of Theoretical Biology*. 2010;263(4): 490–498. doi: 10.1016/j.jtbi.2009.12.014
56. Newton MS, Morrone DJ, Lee KH, Seelig B. Genetic code evolution investigated through the synthesis and characterisation of proteins from reduced-alphabet libraries. *Chembiochem: A European Journal of Chemical Biology*. 2019;20(6): 846–856. doi: 10.1002/cbic.201800668
57. Makarov M, Sanchez Rocha AC, Krystufek R, Cherepashuk I, Dzmitruk V, Charnavets T, et al. Early selection of the amino acid alphabet was adaptively shaped by biophysical constraints of foldability. *Journal of the American Chemical Society*. 2023;145(9): 5320–5329. doi: 10.1021/jacs.2c12987
58. Trifonov EN. Consensus temporal order of amino acids and evolution of the triplet code. *Gene*. 2000;261(1): 139–151. doi: 10.1016/S0378-1119(00)00476-5
59. Ilardo M, Meringer M, Freeland S, Rasulev B, Cleaves HJ II. Extraordinarily adaptive properties of the genetically encoded amino acids. *Scientific Reports*. 2015;5(1): 9414. doi: 10.1038/srep09414
60. Hartman H. Speculations on the evolution of the genetic code. *Origins of Life*. 1975;6(3): 423–427. doi: 10.1007/BF01130344
61. Harrison SA, Palmeira RN, Halpern A, Lane N. A biophysical basis for the emergence of the genetic code in protocells.

- Biochimica et Biophysica Acta - Bioenergetics*. 2022;1863(8): 148597. doi: 10.1016/j.bbabo.2022.148597
62. Halpern A, Bartsch LR, Ibrahim K, Harrison SA, Ahn M, Christodoulou J, et al. Biophysical interactions underpin the emergence of information in the genetic code. *Life*. 2023;13(5): 1129. doi: 10.3390/life13051129
 63. Fisher S, Ezerzer Y, Edri R, Akulenko D, Marland E, Frenkel-Pinter M. Protopeptide backbone affects assembly in aqueous solutions. *Proceedings of the National Academy of Sciences*. 2025;122(40): e2500503122. doi: 10.1073/pnas.2500503122
 64. Fialho DM, Karunakaran SC, Greeson KW, Martínez I, Schuster GB, Krishnamurthy R, et al. Depsipeptide nucleic acids: prebiotic formation, oligomerization, and self-assembly of a new proto-nucleic acid candidate. *Journal of the American Chemical Society*. 2021;143(34): 13525–13537. doi: 10.1021/jacs.1c02287
 65. Marland E, Lipson G, Edri R, Cohen O, Guzman-Martinez A, Shalev O, et al. Robust synthesis of prebiotic precursors in drying reactions of amino acids and keto acids. *ChemSystemsChem*. 2025; 8(1): e00018. doi: 10.1002/syst.202500018.
 66. Li Z, Li L, McKenna KR, Schmidt M, Pollet P, Gelbaum L, et al. The oligomerization of glucose under plausible prebiotic conditions. *Origins of Life and Evolution of Biospheres*. 2019;49(4): 225–240. doi: 10.1007/s11084-019-09588-3
 67. Brunk CF, Marshall CR. 'Whole organism', systems biology, and top-down criteria for evaluating scenarios for the origin of life. *Life*. 2021;11(7): 690. doi: 10.3390/life11070690.
 68. Kocher CD, Dill KA. The prebiotic emergence of biological evolution. *Royal Society Open Science*. 2024;11(7): 240431. doi: 10.1098/rsos.240431
 69. Kocher C, Dill KA. Origins of life: first came evolutionary dynamics. *QRB Discovery*. 2023;4: e4. doi: 10.1017/qrd.2023.2