

APPLICATION DEVELOPMENT ON *BRASSICA* SPECIES IN AQUACULTURE – A REVIEW

Strirengaraj Vijayaram¹, Hary Razafindralambo^{2,3}, Yun-Zhang Sun^{1*}, Hamed Ghafarifarsani^{4*}, Seyed Hossein Hoseinifar⁵,
Mahdieh Raeeszadeh⁶, Hien Van Doan⁷

¹Fisheries College, Jimei University, Xiamen 361021, China

²ProBioLab, Campus Universitaire de la Faculté de Gembloux Agro-Bio Tech/Université de Liège, B-5030 Gembloux, Belgium

³BioEcoAgro Joint Research Unit, TERRA Teaching and Research Centre, Microbial Processes and Interactions,
Gembloux AgroBio Tech/Université de Liège, Gembloux, Belgium

⁴Department of Animal Science, Chaharmahal and Bakhtiari Agricultural and Natural Resources Research and Education Center, AREEO,
Shahrekord, Iran

⁵Department of Fisheries, Faculty of Fisheries and Environmental Sciences, Gorgan University of Agricultural Sciences and Natural Resources,
Gorgan, Iran

⁶Department of Basic Sciences, Sanandaj Branch, Islamic Azad University, Sanandaj, Iran

⁷Department of Animal and Aquatic Sciences, Faculty of Agriculture, Chiang Mai University, Chiang Mai, Thailand

*Corresponding author: Hamed Ghafarifarsani: hamed_ghafari@alumni.ut.ac.ir, Yun-Zhang Sun: jmusunyunzhang@163.com

Abstract

Brassicaceae family are traditional, medicinal and nutritional value plants. *Brassicaceae* family produces valuable components to provide different types of beneficial effects for humans and animals such as anti-oxidant, antimicrobial, anticancer, anti-scurvy, anti-inflammatory, anti-hyperglycemic, anti-tumor, anti-diabetes, anti-stomach diseases, and anti-cardiac. This review mainly focuses to observe *Brassicaceae* family response in aquatic animals and provides some information collected from literature analysis regarding the development of aquaculture applications like *Brassicaceae* family plants used as a supplement to enhance nutritional value, replacement of fish meal, antimicrobial activity against pathogens, immunomodulatory effects, antioxidant, and growth activity of aquatic animals. This literature review analysis is helpful to enlighten for future research and development on the application in the aquaculture sector.

Key words: *Brassica* sp., antimicrobial, antioxidant, immune system, growth, aquaculture

Brassicaceae family can be present approximately on the entire land surface of the globe, but the family is not present in the Antarctica region, and also not present in several areas in the tropics i.e. the Congo basin, north-east Brazil, tropical Australasia, and maritime Southeast Asia. *Brassicaceae* family species are important agricultural crops; some plants are considered indigenous. The economic importance of *Brassicaceae* family plants like cauliflower, broccoli, kale, cabbage, rape, brown mustard, turnip, and rutabaga is high (Schmidt and Bancroft, 2011). Iran's annual vegetable production reaches 30 m tons. Of this amount, almost 800 tons are allocated to broccoli. The United States is the 3rd largest producer of broccoli in the world, with California leading in US production (90%) (Van Hayek, 1911; Schulz, 1936; Azarashkan et al., 2022). *Brassicaceae* (*Cruciferae*) is one of the biggest angiosperm families belonging to the order Brassicales. It includes 12–15 groups with 338–360 genera and 3709 species spread all around the world in the entire continents excluding Antarctica (Al-Shehbaz, 1973, 1984; Appel and Al-Shehbaz, 2003; Al-Shehbaz, 2006). *Brassica* spp. is generally categorized into three different varieties: coles, mustards, and rapeseed. The coles are leafy

and stalk vegetable category incorporated in cauliflower (*Brassica oleracea*), broccoli (*B. oleracea*), cabbage (*B. oleracea*), troncchuda (*B. oleracea*), Brussels sprouts (*B. oleracea*) and kale (*B. oleracea*) and reported to be a most vital source of folates and vitamin C (Dias, 2019). The *Brassica* species are classified into different types such as Asian mustard (*B. juncea*), black mustard (*B. nigra*), white mustard (*B. alba*), Ethiopian mustard (*B. carinata*), rapeseed (*B. rapa*), oilseed rape (*B. napus*), and canola (*B. napus*), which are crucial, economically vital oilseed crops (Rahman et al., 2018). *Brassica napus* is the oil seed crops such as rapeseed or canola, including turnip, Chinese cabbage, and pak choi. *Brassica napus* is one of the significant oil crops after maize and oil palm (Beszterda and Nogala-Kalucka, 2019). Numerous researchers and taxonomists classified the *Brassicaceae* family into various tribes; the *Brassicaceae* family contains annuals, biennials, and also herbaceous perennials. *Brassicaceae* family plants and herbs have the least percentage (5%) of characteristically woody such as *Cramboxylon* spp., *Parolinia* spp., *Zilla spinosa*, *Heliophila scandens*, *Vella* spp., *Faresetia somalensis*, etc. (Franzke et al., 2011). In addition, canola and *Brassica napus* are this

family's major significant oil crops (Alagoz and Toorchi, 2018). Different wild *Brassica* species are a vital source of food in the Mediterranean diet, some studies suggested the nutritional and medicinal effects of the edible wild plants concerning cultivated crops (Malfa et al., 2020; Raeeszadeh et al., 2021 a, b). *Brassica* vegetables have played potent various biological responses such as antibacterial, antiviral, and anticancer activity for the non-specific immune system (Chauhan et al., 2016). Anti-nutritional effects cause different activities such as growth retardation, poor egg production, goiter, and liver damage in animals that have been studied for glucosinolates (GLSs) (Griffiths et al., 1998; Cartea and Velasco, 2008).

Wild *Brassica* species contain numerous beneficial bioactive compounds, and in adaptation to challenging environmental conditions plants directly synthesize secondary metabolites as a chemical defense mechanism (Sánchez-Mata et al., 2012; Vijayaram et al., 2022). *Brassica* species contain a high quantity of flavonols with quercetin, and isorhamnetin as the major aglycones, they are frequently acylated with various hydroxycinnamic acids (Nielsen et al., 1993). *Brassica* vegetable sources contain a high quantity of vitamins like vitamin C, folic acid, tocopherols, and carotenes (Ghafarifarsani et al., 2022 a, b; Kurilich et al., 2002). *Brassica* species is the potential source to produce the rich level of minerals such as calcium (Ca), iron (Fe), phosphorus (P), chlorine (Cl), sulfur (S), potassium (K), and strontium (Sr) (Kmiciek et al., 2007). In addition, *Brassica* species have potential hyperaccumulators in soil-polluted heavy metals such as nickel, zinc, and cadmium as well as *Brassica* species capable of aggregating and tolerating the heavy metals (Devi, 2017). Generally, *B. nigra* species known as black mustard are universally produced and consumed plant species; these species are utilized in different ways like fodder crop, oilseed crop, industrial oil, salad crop, green manure, and green vegetables (Thomas et al., 2012). Black mustard contains different kinds of components such as amino acids, fatty acids, minerals, oligosaccharides, glucosinolates, anti-nutritional factors, and several phenolic compounds as well as the plant species have numerous positive effects like immunomodulatory, antioxidant, antithrombotic, anti-inflammatory, antifungal, antibacterial, anticonvulsant and antidiabetic effects (De Zoysa and Waisundara, 2021; Ghafarifarsani et al., 2021; Ajdari et al., 2022; Ghafarifarsani et al., 2022 c, d, e, 2023).

Lakwani et al. (2022) pointed out that a *Brassica* sp. supplement of 1–2% is supportive to increase the intestinal immunity in rainbow trout for 30 days trial period. Numerous studies described that the incorporation of rapeseed oil supportive to partially replaced fish oil is used to enhance the growth performance of diverse fish species namely red sea bream (*Pagrus major*), yellowtail kingfish (*Seriola lalandi*), chinook salmon (*Oncorhynchus tshawytscha*), rainbow trout (*Oncorhynchus mykiss*), Atlantic salmon (*Salmo salar*) and common carp (*Cyprinus carpio*) (Drew et al., 2007; Huang et al., 2007, 2008; Bowyer et al., 2012; Ghafarifarsani et al., 2022 c, d,

2023). *Brassica* species produced glucosinolates present in rapeseed (*Brassica* spp.) meal and mustard oil cake, and potential significant protein sources in fish feed. *Brassica* species are the alternative development of fish nutrition supplements and they have noticeably improved aquaculture development (Naylor et al., 2009). Various types of plant protein sources were examined to replace fish meal in turbot nutrition (Fournier et al., 2004; Bonaldo et al., 2011; Nagel et al., 2012 a; Yousefi et al., 2022 a, b, c). Kasiga (2018) reported examining the composition and digestibility of carinata (*B. carinata*) meal to hybrid striped bass (*Morone chrysops* × *M. saxatilis*). Carinata meal contains 48% crude protein (which is 98.9% digestible) and different types of components similar to leucine but more arginine, histidine, methionine, threonine, tryptophan, and cysteine. Amino acid level is high compared to solvent-extracted (SE) soybean meal (SE-SBM) with hulls (National Research Council, 2011). Mzengereza et al. (2021 a) reported that dietary supplementation of carinata meal at 20.5 kg/kg (60 days) noticeably enhanced immunity, growth, digestive enzymes, nutrient digestibility, feed utilization capability, and stress resistance activity of red sea bream. Additionally, numerous studies reported the potential importance of camelina oil as a lipid source in aquaculture and livestock sectors (Hixson and Parrish, 2014; Betancor et al., 2018; Toyas-Vargas et al., 2020; Raeeszadeh et al., 2021 a). Dietary incorporation of camelina produces vital components such as sinapine, phytic acid glucosinolates (GSL), and condensed tannins are providing a beneficial response in rainbow trout (Schuster and Friedt, 1998; Matthäus and Zubr, 2000; Russo and Reggiani, 2012). Canola meal and its products have provided beneficial responses like growth and feed utilization of salmonids (Mwachireya et al., 1999; Thiessen et al., 2003, 2004) and also canola meal concentration level increases to improve the performance of fish species (Collins et al., 2012, 2013). Canola meal protein is the most successful protein supplement in aquatic animals such as rainbow trout (*Oncorhynchus mykiss* Walbaum, 1792), high-value crustaceans such as mitten crab, and shrimp species (Cruz-Suarez et al., 2001; Suárez et al., 2009; Luo et al., 2011; Nagel et al., 2012 b; Slawski et al., 2012; Adem et al., 2014; Turchini et al., 2019). Canola meal (19.02%) can replace less than 30% of soybean meal without compromising on growth and feed conversion in juvenile hybrid tilapia (*Oreochromis niloticus* × *Oreochromis aureus*) (Zhou and Yue, 2010). Red cabbage (*B. oleracea* L. var. *Capitata* f. *rubra* DC.) contains more vitamins, minerals, and antioxidant levels compared to other cabbage. Red cabbage supplement is more helpful to enhance the growth, and resistance power of tilapia species (Taufik, 2012). This review provides some information collected from literature analysis regarding the development of aquaculture applications like *Brassica* species used as a supplement to enhance nutritional value, replacement of fish meal, antimicrobial activity against pathogens, immunomodulatory effects, antioxidant, and growth activity of aquatic animals.

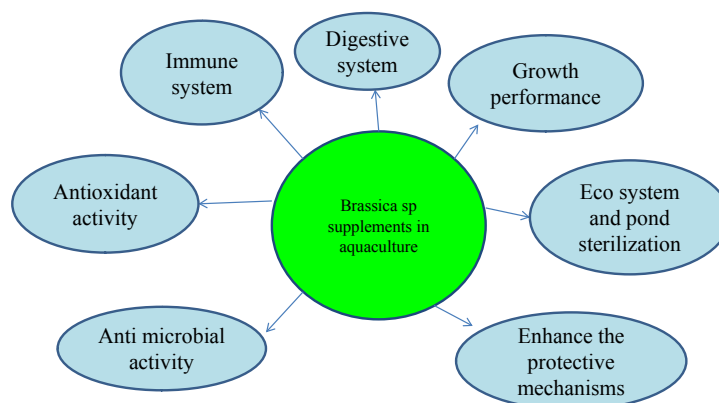
Applications of *Brassica* speciesApplication of *Brassica* sp supplements in aquacultureFigure 1. Role of *Brassica* sp. in aquatic animals

Table 1. Effect of broccoli on growth in aquaculture

<i>Brassica</i> sp. scientific name	Local name	Phytochemical components of <i>Brassica</i> sp.	Aquatic animals Local/scientific name	Response	Reference
1	2	3	4	5	6
<i>Brassica rapa</i> L. and <i>Brassica oleracea</i>	Pak choi and kailan	Polyphenols, phenolic acids, flavonoids, carotenoids (zeaxanthin, lutein, β -carotene), alkaloids, tannins, saponins, anthocyanins, phytosterols, chlorophyll, glucosinolates, phytosteroids, terpenoids, glycosides, vitamin	Tilapia (<i>Oreochromis</i> sp.)	↑ FCR and SGR value	Ariadi et al. (2020)
<i>Brassica oleracea</i>	Cabbage		Nile tilapia (<i>Oreochromis niloticus</i>)	↑ Body weight, final weight gain, growth rate, feed conversion ratio	Oke et al. (2017)
<i>Brassica nigra</i>	Black mustard		Rainbow trout (<i>Oncorhynchus mykiss</i>)	↑ Intestinal immunity	Lakwani et al. (2021)
<i>Brassica carinata</i>	Ethiopian mustard		Hybrid striped bass (<i>Morone saxatilis</i>)	↑ Concentration of essential amino acids	Kasiga et al. (2021)
<i>Camelina</i>	Siberian oilseed		Red seabream (<i>Pagrus major</i>)	Maintain growth, digestive enzymes, immunity, stress resistance and feed utilization	Mzengereza et al. (2021 a, b)
<i>Camelina</i> high-oil residual camelina meal	Siberian oilseed		Rainbow trout (<i>Oncorhynchus mykiss</i>)	↑ Glucosinolate level ↓ Feed intake	Buller et al. (2016)
			Rainbow trout (<i>Oncorhynchus mykiss</i>)	No adverse effects on growth performance	Pan et al. (2011)
<i>Brassica napus</i>	Rapeseed		Rainbow trout (<i>Oncorhynchus mykiss</i>)	↑ Overall growth ↓ Feed intake	Kaiser et al. (2021)
<i>Camelina</i>	Siberian oilseed		Rainbow trout (<i>Oncorhynchus mykiss</i>)	↑ Overall growth	Koskela et al. (2021)
<i>Brassica oleracea</i>	Red cabbage		Nile tilapia (<i>Oreochromis niloticus</i>)	↑ Growth and survival range	Nasution (2021)
<i>Brassica napus</i>	Rapeseed		GIFT (juvenile genetically improved farmed tilapia)	23.2% is suitable level to enhance growth, feed utilization and physiological function	Wu et al. (2021)
Canola oil	Turnip		Pacific white shrimp (<i>Litopenaeus vannamei</i>)	100% replacement of fish meal	Gia Vo et al. (2021)

Table 1 – contd.

1	2	3	4	5	6
<i>Brassica rapa</i> L.	Pak choi		Nile tilapia (<i>Oreochromis niloticus</i>)	Maintain the level of ammonia, nitrate and nitrite in fish ponds	Febriani et al. (2018)
<i>Brassica napus</i> , <i>Camelina sativa</i> and <i>Brassica carinata</i>	Canola, camelina and Ethiopian mustard		Rainbow trout fingerlings (<i>Oncorhynchus mykiss</i>)	↑ Growth performance	Bullerwell et al. (2016); Anderson et al. (2018)
<i>Brassica juncea</i> L.	Samhong mustard		Nile tilapia (<i>Oreochromis niloticus</i>)	↑ Growth performance and removal of nutrients from fish farm water	Deswati et al. (2020)
<i>Brassica oleracea</i> , <i>B. napus</i> and <i>B. juncea</i>	Broccoli, cole and potherb mustard		Shrimp <i>Limpenaeus vannamei</i>	Suitable model to enhance shrimp quality and ecological development of shrimp pond	Ni et al. (2020)

Table 2. Broccoli and its systemic effects in aquatic animals

Scientific name	Local name	Aquatic animals Local/scientific name	Response	Reference
<i>Brassica oleracea</i>	Red cabbage	Nile tilapia (<i>Oreochromis niloticus</i>)	↑ Detoxification metabolism of fish	Villa-Cruz et al. (2009)
<i>Brassica oleracea</i>	Broccoli	Nile tilapia (<i>Oreochromis niloticus</i>)	Novel protective agents against water pollutants	Davila et al. (2010)
<i>Brassica oleracea</i>	Red cabbage	Common carp (<i>Cyprinus carpio</i>)	↑ Nutrient efficiency, growth performance, and hematological parameters	Kadhim et al. (2022)
<i>Brassica oleracea</i>	Red cabbage	Nile tilapia (<i>Oreochromis niloticus</i>)	↑ Cellular and humoral immunity	Pisuttharachai et al. (2020)

Antimicrobial activity

Brassica species contain volatile sulfur compounds originating from S-alk(en)yl-L-cysteine sulfoxide, as well as glucosinolates. Sulfur compounds are commonly present in two families: the *Brassicaceae* and the *Liliaceae* (Kyung and Lee, 2001). *Brassica* species cabbage (*B. tolerance*) extract showed potential antimicrobial response (Sherman and Hodge, 1936). *B. rapa* seeds showed potential antimicrobial response against *Salmonella paratyphi*, *Pseudomonas aeruginosa*, *Staphylococcus aureus*, *Escherichia coli*, and *Klebsiella pneumonia* (Muluye et al., 2015; Danlami et al., 2016). Researchers pointed out that *Brassica* species like radish root, kale leaves, and mustard seeds showed antimicrobial action against *Bacillus subtilis*, *Staphylococcus aureus*, *Salmonella typhimurium*, *S. aureus*, *Enterobacter faecalis*, *Moraxella catarrhalis*, *Listeria monocytogenes* and *Escherichia coli* (Ayaz et al., 2008; Beevi et al., 2009; Engels et al., 2012).

Immune system response

Brassica species and their components like tocopherol, ascorbic acid, canolol, carotenoids (lutein and carotene), phenolic acids (gallic acid, caffeic acid, sinapic acid, ferulic acid, and 3,4-di-hydroxybenzoic acid), and flavonoids (rutin, quercetin, and kaempferol) showed antioxidant activity to protect the immune system by neutralizing free radicals (Dua et al., 2014; Salehi et

al., 2021; Grygier, 2022). Dietary incorporation of *B. oleracea* powder (0.5 g/100 g) noticeably enhanced feed utilization and nonspecific immune diseases in Nile tilapia (Oke et al. 2017; Raeeszadeh et al., 2022 a). Black mustard (*B. nigra*) supplement at 1–2 g/kg dosage for 30 days provides beneficial effects to improve immune response, pro- and anti-inflammatory responses in the intestine, and disease resistance against *Aeromonas hydrophila* in rainbow trout (Lakwani et al., 2022). Dietary incorporation of carinata 30% is improving palatability and overall nutrient value in sunshine bass; carinata supplements contain high iron contents and are useful to enhance the hemoglobin content of red blood cells (Kasiga and Brown, 2019; Raeeszadeh et al., 2022 b).

Antioxidant activity

Brassica species contain high antioxidant activity; numerous studies pointed out that different species of *Brassica* such as *B. rapa*, *B. oleracea*, *B. napus*, and *B. juncea* extract showed antioxidant activity (Raeeszadeh et al., 2022 a, b). The antioxidant activity was measured by *in vitro* methods based on the use of free radicals scavenging (Costa et al., 2005; Gawlik-Dziki, 2008; Miglio et al., 2008; Lemoine et al., 2010; Cogo et al., 2011; Liang et al., 2019; Ninh Le et al., 2019). Arora and Vig (2015) reported that *B. rapa* dichloromethane extract showed antioxidant activity by the inhibition of hydrox-

yl radicals responsible for oxidative damage to DNA at 1000 g/ml concentration by phenylethylbrassinin and 3-phenylpropionitrile by *in vivo* methods. *B. fruticulosa* subsp. *fruticulosa* leaf hydroalcoholic extract has phytochemical and antioxidant properties. The plant extract is non-toxic and potentially safe for shrimp larvae and is also a potential health-promoting factor in aquatic animals (Cavò et al., 2022). Aqueous and ethanolic extracts of broccoli showed the potential antioxidant activity of different reactive oxygen species/free radicals by *in vitro* conditions. Broccoli is a source of natural antioxidants or nutraceuticals with the possible function to reduce oxidative stress with consequent health benefits (Bidchol et al., 2011; Raeeszadeh et al., 2022 a). Aqueous and ethanolic extracts of cauliflower have effective antioxidant activity by various *in vitro* assays such as DPPH scavenging, ABTS⁺ scavenging, DMPD⁺ scavenging, Cu²⁺-Cu⁺ reducing power assays, ferric thiocyanate method, Fe³⁺-Fe²⁺ and superoxide anion radical scavenging, hydrogen peroxide scavenging, and metal chelating activity. The antioxidant results compared standard antioxidant compounds α -tocopherol (a natural antioxidant) and Trolox (the water-soluble analog of tocopherol) (Qaderi Forough et al., 2017). Cauliflower contains natural antioxidants and potential use for food additives or pharmaceutical applications (Köksal and Gülçin, 2008).

Growth response

Dietary incorporation of rapeseed oil was applied to examine the production and growth parameters of rainbow trout and the study results indicate no changes in final weight, weight gain, hepatosomatic index, or viscerosomatic index compared to fish meal and fish oil-based diet (Caballero et al., 2002; Drew et al., 2007; Turchini et al., 2013; Kaiser et al., 2021). The dietary additive of pak choy (*B. rapa* subsp. *chinensis*) supplement is a noticeable improvement of significant growth ratio of 3.15% and a feed conversion ratio at 1.21 of tilapia compared to the kalia plant in the aquaponic system (Ariadi et al., 2020; Raeeszadeh et al., 2022 b). Dietary additives of camelina oil are utilized to replace fish oil. Camelina oil contains limited fatty acids, eicosapentaenoic acid (EPA), and docosahexaenoic acids (DHA) which are helpful to maintain growth and sustain health performance in red seabream (*Pagrus major*). A camelina diet supplement is a successful effective method for the growth development of aquatic animals (Mzengereza et al., 2021 b). Dietary supplementation of red cabbage liquid organic supplement had a noticeable effect to improve growth performance at 2.84–3.90% per day and survival range from 88 to 92% in tilapia. Red cabbage liquid supplement was supportive to enhance growth performance and decreased mortality rate in tilapia (*Oreochromis niloticus*) (Nasution, 2021). Dietary supplementation of rapeseed meal at 23.2% noticeably improved physiological function, feed utilization, flesh quality, and growth performance of improved farmed tilapia (GIFT);

the higher concentration of rapeseed diet (46.4%) can reduce growth action, damaged liver function, and suppress the immune function of tilapia. Appropriate quantity of the supplement helps to improve growth performance in tilapia (Wu et al., 2021). Dietary supplementation was of canola oil helpful to replace menhaden fish oil up to 100% in fishmeal-based diets and up to 75% in poultry meals without affecting the growth performance of *L. vannamei*. Canola oil supplements are not only developed for particular species, they are potentially important multi-species feed ingredients in the aquaculture sector (Gia Vo et al., 2021). Dietary supplementation of canola (*B. napus*) meal with GLS content of 2.18 μ moles/g significantly did not affect feed intake or growth performance of red seabream (Glencross et al., 2004 a, b). Dietary supplementation of glucosinolates and sinapic acid concentrations has no adverse effects on the growth and physiological function of turbot, and plant proteins are helpful to replace a fish meal or fish oil, and most significant supplement for fish nutrition (von Danwitz and Schulz, 2020).

Conclusion

Brassica species is the most important traditional, medicinal, and food-value of plants. *Brassica* species and their components have more positive effects in human and livestock such as anti-oxidant, antimicrobial, anticancer, anti-scurvy, anti-inflammatory, anti-hyperglycemic, anti-tumor, diabetes, stomach diseases, and cardioprotective, but so far very limited applications are available in aquaculture. This review provides some information about *Brassica* species, and their components that are utilized to enhance antimicrobial activity, antioxidant activity, immune system against infections, and growth performance in aquatic animals. This review is helpful to develop the aquaculture application in future research. In the future *Brassica* species and their components may be utilized to enhance aquatic animals' health, increase healthy aquatic production and aquatic feed quality in the feed industry, and also to understand clear metabolic pathway study.

Data availability

The data that support the findings of this study are available upon reasonable request to the corresponding authors.

Compliance with ethical standards

All applicable international, national, and/or institutional guidelines for the care and use of animals were followed.

Conflicts of interest

The authors declare no conflict of interest.

Acknowledgment

This research work was supported by Xiamen Key Laboratory for Feed Quality Testing and Safety Evalu-

ation, Fisheries College, Jimei University, Xiamen 361021, and additionally supported by China Chiang Mai University, Chiang Mai, Thailand.

References

- Adem H.N., Tressel R.P., Pudal F., Slawski H., Schulz C. (2014). Rapeseed use in aquaculture. *OCL*, 21: D105.
- Ajdari A., Ghafarifarsani H., Hoseinifar S.H., Javahery S., Narimanizad F., Gatphayak K., Van Doan H. (2022). Effects of dietary supplementation of primaLac inulin and biomin imbo on growth performance antioxidant and innate immune responses of common carp (*Cyprinus carpio*). *Aquacult. Nutr.*, 2022: 1–13.
- Alagoz S.M., Toorchi M. (2018). An investigation of some key morpho-physiological attributes and leaf proteome profile in canola (*Brassica napus* L.) under salinity stress. *Pak. J. Bot.*, 50: 847–852.
- Al-Shehbaz I.A. (1973). The biosystematics of the genus *Thelypodium* (*Cruciferae*). Contributions from the Gray Herbarium of Harvard University, 204: 3–148.
- Al-Shehbaz I.A. (1984). The tribes of *Cruciferae* (*Brassicaceae*) in the southeastern United States. *J. Arnold Arboretum.*, 65: 343–373.
- Al-Shehbaz I.A., Beilstein M.A., Kellogg E.A. (2006). Systematics and phylogeny of the *Brassicaceae* (*Cruciferae*): an overview. *Plant Sys. Evol.*, 259: 89–120.
- Anderson D.M., MacPherson M.J., Collins S.A., MacIsaac P.F. (2018). Yellow- and brown-seeded canola (*Brassica napus*), camelina (*Camelina sativa*) and Ethiopian mustard (*Brassica carinata*) in practical diets for rainbow trout fingerlings. *J. Appl. Aquacult.*, 30: 187–195.
- Appel O., Al-Shehbaz I.A. (2003). *Cruciferae*. In: Flowering plants. Dicotyledons, Kubitzki K., Bayer C. (eds). Springer Berlin Heidelberg, pp. 75–174.
- Ariadi H., Pandaingan I.A., Soeprijanto A., Maemunah Y., Wafi A. (2020). Effectiveness of using pakcoy (*Brassica rapa* L.) and kailan (*Brassica oleracea*) plants as vegetable media for aquaponic culture of tilapia (*Oreochromis* sp.). *J. Aquacult. Dev. Env.*, 3: 156–162.
- Arora S., Vig A.P. (2015). Inhibition of DNA oxidative damage and antimutagenic activity by dichloromethane extract of *Brassica rapa* var. *rapa* L. seeds. *Ind. Crops. Prod.*, 74: 585–591.
- Ayaz F.A., Hayirlioglu-Ayaz S., Alpay-Karaoglu S., Grúz J., Valentová K., Ulrichová J., Strnad M. (2008). Phenolic acid contents of kale (*Brassica oleracea* L. var. *acephala* DC.) extracts and their antioxidant and antibacterial activities. *Food Chem.*, 107: 19–25.
- Azarashkan Z., Motamedzadegan A., Ghorbani-HasanSaraei A., Biparva P., Rahaiee S. (2022). Investigation of the physicochemical antioxidant rheological and sensory properties of ricotta cheese enriched with free and nano-encapsulated broccoli sprout extract. *Food Sci. Nutr.*, 10: 4059–4072.
- Beevi S.S., Mangamoori L.N., Dhand V., Ramakrishna D.S. (2009). Isothiocyanate profile and selective antibacterial activity of root stem and leaf extracts derived from *Raphanus sativus* L. *Foodborne Path. Dis.*, 6: 129–136.
- Beszterda M., Nogala-Kalucka M. (2019). Current research developments on the processing and improvement of the nutritional quality of rapeseed (*Brassica napus* L.). *European J. Lipid Sci. Technol.*, 121: 1–18.
- Betancor M.B., Li K., Bucerzan V.S., Sprague M., Sayanova O., Usher S., Olsen R.E. (2018). Oil from transgenic *Camelina sativa* containing over 25% n-3 long-chain PUFA as the major lipid source in feed for Atlantic salmon (*Salmo salar*). *Brit. J. Nutr.*, 119: 1378–1392.
- Bidchol A.M., Wilfred A., Abhijna P., Harish R. (2011). Free radical scavenging activity of aqueous and ethanolic extract of *Brassica oleracea* L. var. *italica*. *Food Bioproc. Technol.*, 4: 1137–1143.
- Bonaldo A., Parma L., Mandrioli L., Sirri R., Fontanillas R., Badiani A., Gatta P.P. (2011). Increasing dietary plant proteins affects growth performance and ammonia excretion but not digestibility and gut histology in turbot (*Psetta maxima*) juveniles. *Aquaculture*, 318: 101–108.
- Bowyer J.N., Qin J.G., Smullen R.P., Stone D.A.J. (2012). Replacement of fish oil by poultry oil and canola oil in yellowtail kingfish (*Seriola lalandi*) at optimal and suboptimal temperatures. *Aquaculture*, 356: 211–222.
- Bullerwell C.N., Collins S.A., Lall S.P., Anderson D.M. (2016). Growth performance proximate and histological analysis of rainbow trout fed diets containing *Camelina sativa* seeds meal (high-oil and solvent-extracted) and oil. *Aquaculture*, 452: 342–350.
- Caballero M.J., Obach A., Rosenlund G., Montero D., Gisvold M., Izquierdo M.S. (2002). Impact of different dietary lipid sources on growth lipid digestibility tissue fatty acid composition and histology of rainbow trout *Oncorhynchus mykiss*. *Aquaculture*, 214: 253–271.
- Cartea M.E., Velasco P. (2008). Glucosinolates in *Brassica* foods: bioavailability in food and significance for human health. *Phytochem. Rev.*, 7: 213–229.
- Cavò E., Taviano M.F., Davi F., Cacciola F., Oulad El Majdoub Y., Mondello L., Miceli N. (2022). Phenolic and volatile composition and antioxidant properties of the leaf extract of *Brassica fruticulosa* subsp. *fruticulosa* (*Brassicaceae*) growing wild in Sicily (Italy). *Molecules*, 27: 2768.
- Chauhan E.S., Tiwari A., Singh A. (2016). Phytochemical screening of red cabbage (*Brassica oleracea*) powder and juice – A comparative study. *J. Med. Plants Stud.*, 4: 196–199.
- Cogo S.L.P., Chaves F.C., Schirmer M.A., Zambiasi R.C., Nora L., Silva J.A., Rombaldi C.V. (2011). Low soil water content during growth contributes to preservation of green colour and bioactive compounds of cold-stored broccoli (*Brassica oleracea* L.) florets. *Postharvest Biol. Technol.*, 60: 158–163.
- Collins S.A., Desai A.R., Mansfield G.S., Hill J.E., Van Kessel A.G., Drew M.D. (2012). The effect of increasing inclusion rates of soybean pea and canola meals and their protein concentrates on the growth of rainbow trout: concepts in diet formulation and experimental design for ingredient evaluation. *Aquaculture*, 344: 90–99.
- Collins S.A., Øverland M., Skrede A., Drew M.D. (2013). Effect of plant protein sources on growth rate in salmonids: Meta-analysis of dietary inclusion of soybean pea and canola/rapeseed meals and protein concentrates. *Aquaculture*, 400: 85–100.
- Costa M.L., Civello P.M., Chaves A.R., Martínez G.A. (2005). Effect of hot air treatments on senescence and quality parameters of harvested broccoli (*Brassica oleracea* L. var. *italica*) heads. *J. Sci. Food. Agric.*, 85: 1154–1160.
- Cruz-Suarez L.E., Ricque-Marie D., Tapia-Salazar M., McCallum I.M., Hickling D. (2001). Assessment of differently processed feed pea (*Pisum sativum*) meals and canola meal (*Brassica* sp.) in diets for blue shrimp (*Litopenaeus stylirostris*). *Aquaculture*, 196: 87–104.
- Danlami U., Orishadipe Abayomi T., Lawal D.R. (2016). Phytochemical nutritional and antimicrobial evaluations of the aqueous extract of *Brassica nigra* (*Brassicaceae*) seeds. *Am. J. Appl. Chem.*, 4: 161.
- Davila J., Marcial-Martinez L.M., Viana M.T., Vazquez-Duhalt R. (2010). The effect of broccoli in diet on the cytochrome P450 activities of tilapia fish (*Oreochromis niloticus*) during phenol exposure. *Aquaculture*, 304: 58–65.
- De Zoysa H., Waisundara V.Y. (2021). Mustard (*Brassica nigra*) seed. In: Oilseeds: Health attributes food applications, Tanwar B., Goyal A. (eds). Springer, pp. 191–210.
- Deswati D., Yani E., Safni S., Norita Tetra O., Pardi H. (2020). Development methods in aquaponics systems using biofloc to improve water quality (ammonia nitrite nitrate) and growth of tilapia and samhong mustard. *Int. J. Env. Anal. Chem.*, 102: 7824–7834.
- Devi M. (2017). Biological properties of soil and nutrient uptake in cauliflower (*Brassica oleracea* var. *botrytis* L.) as influenced by integrated nutrient management. *J. Pharmacogn. Phytochem.*, 6: 325–328.
- Dias J.S. (2019). Nutritional quality and effect on disease prevention of vegetables. In: Nutrition in health and disease-our challenges now and forthcoming time, Mozsik G., Figler M. (eds). IntechOpen.

- Drew M.D., Ogunkoya A.E., Janz D.M., Van Kessel A.G. (2007). Dietary influence of replacing fish meal and oil with canola protein concentrate and vegetable oils on growth performance fatty acid composition and organochlorine residues in rainbow trout (*Oncorhynchus mykiss*). *Aquaculture*, 267: 260–268.
- Dua A., Chander S., Agrawal S., Mahajan R. (2014). Antioxidants from defatted Indian mustard (*Brassica juncea*) protect biomolecules against *in vitro* oxidation. *Physiol. Molecul. Biol. Plants*, 20: 539–543.
- Engels C., Schieber A., Gänzle M.G. (2012). Sinapic acid derivatives in defatted Oriental mustard (*Brassica juncea* L.) seed meal extracts using UHPLC-DAD-ESI-MSⁿ and identification of compounds with antibacterial activity. *European Food. Res. Technol.*, 234: 535–542.
- Febriani N., Pardi H., Yusuf Y., Suyani H. (2018). Applications of aquaponics on pakcoy (*Brassica rapa* L.) and nila fish (*Oreochromis niloticus*) to the concentration of ammonia nitrite and nitrate. *Orient. J. Chem.*, 34: 2447.
- Fournier V., Huelvan C., Desbruyeres E. (2004). Incorporation of a mixture of plant feedstuffs as substitute for fish meal in diets of juvenile turbot (*Psetta maxima*). *Aquaculture*, 236: 451–465.
- Franzke A., Lysak M.A., Al-Shehbaz I.A., Koch M.A., Mummenhoff K. (2011). Cabbage family affairs: the evolutionary history of *Brassicaceae*. *Trends Plant Sci.*, 16: 108–116.
- Gawlik-Dziki U. (2008). Effect of hydrothermal treatment on the antioxidant properties of broccoli (*Brassica oleracea* var. *botrytis Italica*) florets. *Food Chem.*, 109: 393–401.
- Ghafariarsani H., Rashidian G., Sheikhlari A., Naderi Farsani M., Hoseinifar S.H., Van Doan H. (2021). The use of dietary oak acorn extract to improve haematological parameters mucosal and serum immunity skin mucus bactericidal activity and disease resistance in rainbow trout (*Oncorhynchus mykiss*). *Aquacult. Res.*, 52: 2518–2527.
- Ghafariarsani H., Hoseinifar S.H., Javahery S., Van Doan H. (2022 a). Effects of dietary vitamin C thyme essential oil and quercetin on the immunological and antioxidant status of common carp (*Cyprinus carpio*). *Aquaculture*, 553: 738053.
- Ghafariarsani H., Hoseinifar S.H., Javahery S., Yazici M., Van Doan H. (2022 b). Growth performance, biochemical parameters, and digestive enzymes in common carp (*Cyprinus carpio*) fed experimental diets supplemented with vitamin C, thyme essential oil, and quercetin. *Italian. J. Anim. Sci.*, 21: 291–302.
- Ghafariarsani H., Hoseinifar S.H., Sheikhlari A., Raissy M., Chaharmahali F.H., Maneepitaksanti W., Van Doan H. (2022 c). The effects of dietary thyme oil (*Thymus vulgaris*) essential oils for common carp (*Cyprinus carpio*): growth performance digestive enzyme activity antioxidant defense tissue and mucus immune parameters and resistance against *Aeromonas hydrophila*. *Aquacult. Nutr.*, 13: 7942506.
- Ghafariarsani H., Hoseinifar S.H., Aftabgard M., Van Doan H. (2022 d). The improving role of savory (*Satureja hortensis*) essential oil for Caspian roach (*Rutilus rutilus*) fry: Growth haematological immunological and antioxidant parameters and resistance to salinity stress. *Aquaculture*, 548: 737653.
- Ghafariarsani H., Yousefi M., Hoseinifar S.H., Paolucci M., Lumsangkul C., Jaturasitha S., Van Doan H. (2022 e). Beneficial effects of Persian shallot (*Allium hirtifolium*) extract on growth performance biochemical immunological and antioxidant responses of rainbow trout *Oncorhynchus mykiss* fingerlings. *Aquaculture*, 555: 738162.
- Ghafariarsani H., Aftabgard M., Hoseinifar S.H., Raeeszadeh M., Van Doan H. (2023). Comparative effects of savory (*Satureja hortensis*), dill (*Anethum graveolens*) and mooseer (*Allium hirtifolium*) essential oils on growth digestive immunoantioxidant parameters and resistance to *Aeromonas hydrophila* in juvenile common carp (*Cyprinus carpio*). *Aquaculture*, 739541.
- Gia Vo L.L., Galkanda-Arachchige H.S., Iassonova D.R., Davis D.A. (2021). Efficacy of modified canola oil to replace fish oil in practical diets of Pacific white shrimp *Litopenaeus vannamei*. *Aquacult. Res.*, 52: 2446–2459.
- Glencross B., Hawkins W., Curnow J. (2004 a). Nutritional assessment of Australian canola meal. Part I. Evaluation of canola oil extracted methods and meal processing conditions on the digestible value of canola meal fed to the red seabream (*Pagrus auratus*). *Aquacult. Res.*, 35: 15–24.
- Glencross B., Hawkins W., Curnow J. (2004 b). Nutritional assessment of Australian canola meal. Part II. Evaluation of canola oil extracted methods on the protein value of canola meal fed to the red seabream (*Pagrus auratus*). *Aquacult. Res.*, 35: 25–34.
- Griffiths D.W., Birch A.N.E., Hillman J.R. (1998). Antinutritional compounds in the *Brasi* analysis biosynthesis chemistry and dietary effects. *J. Hort. Sci. Biotechnol.*, 73: 1–18.
- Grygier A. (2022). Mustard seeds as a bioactive component of food. *Food Rev. Int.*, 1–14.
- Hixson S.M., Parrish C.C. (2014). Substitution of fish oil with camelina oil and inclusion of camelina meal in diets fed to Atlantic cod (*Gadus morhua*) and their effects on growth tissue lipid classes and fatty acids. *J. Anim. Sci.*, 92: 1055–1067.
- Huang S.S.Y., Oo A.N., Higgs D.A., Brauner C.J., Satoh S. (2007). Effect of dietary canola oil level on the growth performance and fatty acid composition of juvenile red sea bream *Pagrus major*. *Aquaculture*, 271: 420–431.
- Huang S.S.Y., Fu C.H.L., Higgs D.A., Balfry S.K., Schulte P.M., Brauner C.J. (2008). Effects of dietary canola oil level on growth performance fatty acid composition and ionoregulatory development of spring chinook salmon parr *Oncorhynchus tshawytscha*. *Aquaculture*, 274: 109–117.
- Kadhim A.H., Yesser A.T., Al-Niaem K.S. (2022). Effect of broccoli *Brassica oleracea* extract on growth performance and some blood parameters of common carp *Cyprinus carpio*. *Int. J. Aquatic Biol.*, 10: 398–405.
- Kaiser F., Harloff H.J., Tressel R.P., Steffens D., Seibel H., Jung C., Schulz C. (2021). Effects of supplemented anti-nutritive substances from rapeseed on growth and health performance of rainbow trout (*Oncorhynchus mykiss*). *Aquaculture*, 536: 736422.
- Kasiga T. (2018). Nutritional value of modified novel oilseed meals in teleost diets. South Dakota State University.
- Kasiga T., Brown M.L. (2019). Nutritional utilization of modified carinata *Brassica carinata* meals in sunshine bass diets. *North American J. Aquacult.*, 81: 372–384.
- Kasiga T., White B.M., Gibbons W.R., Brown M.L. (2021). Amino acid availability of processed *Brassica carinata* meals in hybrid striped bass *Morone chrysops* ♀ × *M. saxatilis* ♂. *Aquacult. Rep.*, 19: 100597.
- Kmiecik W., Lisiewska Z., Korus A. (2007). Retention of mineral constituents in frozen brassicas depending on the method of preliminary processing of the raw material and preparation of frozen products for consumption. *European Food Res. Technol.*, 224: 573–579.
- Köksal E., Gülçin İ. (2008). Antioxidant activity of cauliflower (*Brassica oleracea* L.). *Turkish J. Agricult. Forestry*, 32: 65–78.
- Koskela J., Leskinen H., Mattila P., Airaksinen S., Rinne M., Pihlavan J.M., Pihlavan A. (2021). The effect of gradual addition of camelina seeds in the diet of rainbow trout (*Oncorhynchus mykiss*) on growth feed efficiency and meat quality. *Aquacult. Res.*, 52: 4681–4692.
- Kurilich A.C., Jeffery E.H., Juvik J.A., Wallig M.A., Klein B.P. (2002). Antioxidant capacity of different broccoli (*Brassica oleracea*) genotypes using the oxygen radical absorbance capacity (ORAC) assay. *J. Agricult. Food Chem.*, 50: 5053–5057.
- Kyung K.H., Lee Y.C. (2001). Antimicrobial activities of sulfur compounds derived from S-alk (en) yl-L-cysteine sulfoxides in *Allium* and *Brassica*. *Food Rev. Int.*, 17: 183–198.
- Lakwani M.A., Kenanoğlu O.N., Taştan Y., Bilen S. (2022). Effects of black mustard (*Brassica nigra*) seed oil on growth performance digestive enzyme activities and immune responses in rainbow trout (*Oncorhynchus mykiss*). *Aquacult. Res.*, 53: 300–313.
- Lemoine M.L., Chaves A.R., Martínez G.A. (2010). Influence of combined hot air and UV-C treatment on the antioxidant system of minimally processed broccoli (*Brassica oleracea* L. var. *Italica*). *LWT – Food Sci. Technol.*, 43: 1313–1319.
- Liang J.L., Yeow C.C., Teo K.C., Gnanaaraj C., Chang Y.P. (2019). Valorizing cabbage (*Brassica oleracea* L. var. *capitata*) and cap-

- sicum (*Capsicum annuum* L.) wastes: In vitro health-promoting activities. *J. Food Sci. Technol.*, 56: 4696–4704.
- Luo Z., Li X.D., Wang W.M., Tan X.Y., Liu X. (2011). Partial replacement of fish meal by a mixture of soybean meal and rapeseed meal in practical diets for juvenile Chinese mitten crab *Eriocheir sinensis*: effects on growth performance and *in vivo* digestibility. *Aquacult. Res.*, 42: 1615–1622.
- Malfa G.A., Acquaviva R., Bucchini E., Ragusa S., Raimondo F.M., Spadaro V. (2020). The Sicilian wild cabbages as biological resources: taxonomic update and a review on chemical constituents and biological activities. *Fl. Medit.*, 30: 245–260.
- Matthäus B., Zubr J. (2000). Variability of specific components in *Camelina sativa* oilseed cakes. *Indu Crops Products.*, 12: 9–18.
- Miglio C., Chiavaro E., Visconti A., Fogliano V., Pellegrini N. (2008). Effects of different cooking methods on nutritional and physico-chemical characteristics of selected vegetables. *J. Agric. Food Chem.*, 56: 139–147.
- Muluye A.B., Melese E., Adinew G.M. (2015). Antimalarial activity of 80% methanolic extract of *Brassica nigra* (L.) Koch. (*Brassicaceae*) seeds against *Plasmodium berghei* infection in mice. *BMC Compl. Alter. Med.*, 15: 1–8.
- Mwachireya S.A., Beames R.M., Higgs D.A., Dosanjh B.S. (1999). Digestibility of canola protein products derived from the physical enzymatic and chemical processing of commercial canola meal in rainbow trout *Oncorhynchus mykiss* (Walbaum) held in fresh water. *Aquacult. Nutr.*, DOI:10.1046/J.1365-2095.1999.00089.X
- Mzengereza K., Ishikawa M., Koshio S., Yokoyama S., Yukun Z., Shadrack R.S., Dawood M. A. (2021 a). Growth performance growth-related genes digestibility digestive enzyme activity immune and stress responses of *de novo* camelina meal in diets of red seabream (*Pagrus major*). *Animals*, 11: 3118.
- Mzengereza K., Ishikawa M., Koshio S., Yokoyama S., Yukun Z., Shadrack R.S., Dawood M.A. (2021 b). Effect of substituting fish oil with camelina oil on growth performance fatty acid profile digestibility liver histology and antioxidative status of red seabream (*Pagrus major*). *Animals*, 11: 1990.
- Nagel F., von Danwitz A., Tusche K., Kroeckel S., van Bussel C.G., Schlachter M., Schulz C. (2012 a). Nutritional evaluation of rapeseed protein isolate as fish meal substitute for juvenile turbot (*Psetta maxima* L.) – impact on growth performance body composition nutrient digestibility and blood physiology. *Aquaculture*, 356: 357–364.
- Nagel F., Slawski H., Adem H., Tressel R.P., Wysujack K., Schulz C. (2012 b). Albumin and globulin rapeseed protein fractions as fish meal alternative in diets fed to rainbow trout (*Oncorhynchus mykiss* W.). *Aquaculture*, 354: 121–127.
- Nasution N. (2021). The effect of liquid organic plant supplements on aquaponic culture of red cabbage (*Brassica oleracea* L. var. *Capitata* f. *rubra* DC.) and tilapia (*Oreochromis niloticus* L.). *Proc. IOP Conference Series: Earth and Environmental Science*, 912: 012102.
- National Research Council (2011). Nutrient requirements of fish and shrimp. National Academies Press.
- Naylor R.L., Hardy R.W., Bureau D.P., Chiu A., Elliott M., Farrell A.P., Nichols P.D. (2009). Feeding aquaculture in an era of finite resources. *Proc. Nat. Acad. Sci.*, 106: 15103–15110.
- Ni M., Yuan J., Hua J., Lian Q., Guo A., Liu M., Gu Z. (2020). Shrimp – vegetable rotational farming system: an innovation of shrimp aquaculture in the tidal flat ponds of Hangzhou Bay China. *Aquaculture*, 518: 734864.
- Nielsen J.K., Olsen C.E., Petersen M.K. (1993). Acylated flavonol glycosides from cabbage leaves. *Phytochemistry*, 34: 539–544.
- Ninh Le T., Luong H.Q., Li H.P., Chiu C.H., Hsieh P.C. (2019). Broccoli (*Brassica oleracea* L. var. *Italica*) sprouts as the potential food source for bioactive properties: A comprehensive study on *in vitro* disease models. *Food*, 8: 532.
- Oke I.O., Dada A.A., Adebayo A.O. (2017). Dietary effects of *Brassica oleracea* on growth performance and haematological parameters of *Oreochromis niloticus* fingerlings. *J. Food Agric. Environ.*, 13: 133–136.
- Pan X., Xie W., Caldwell C.D., Anderson D.M. (2011). Growth performance and carcass composition of rainbow trout (*Oncorhynchus mykiss*) fed practical diets containing graded levels of high fat residue Camelina meal. *Canadian J. Anim. Sci.*, 91: 484–484.
- Pisuttharachai D., Nalinanon W., Areechon N. (2020). Effects of dietary supplementation with broccoli sprouts (*Brassica oleracea*) on the hematology and immunological responses of Nile tilapia (*Oreochromis niloticus*). *J. Fisheries Environ.*, 44: 65–75.
- Qaderi Forough M.M., Raeeszadeh A., Amiri A. (2017). Dose-response changes of *Brassica oleracea* var. *italica* hydroalcoholic extract in the control of oxidative stress by induction of diazinon on the cells of testicular tissue in male adult rat. *J. Rafsanjan. Univ. Med. Sci.*, 16: 593–604.
- Raeeszadeh M., Akbari A. (2021 a). The effects of broccoli and caraway extracts on serum oxidative markers testicular structure and function and sperm quality before and after sperm cryopreservation. *Cryobiology*, 99: 11–19.
- Raeeszadeh M., Moradi M., Ayar P., Akbari A. (2021 b). The antioxidant effect of *Medicago sativa* L. (alfalfa) ethanolic extract against mercury chloride (HgCl₂) toxicity in rat liver and kidney: an *in vitro* and *in vivo* study. *Evidence-Based Compl. Alt. Med.*, 8388002.
- Raeeszadeh M., Karimi P., Khademi N., Mortazavi P. (2022 a). The effect of broccoli extract in arsenic-induced experimental poisoning on the hematological biochemical and electrophoretic parameters of the liver and kidney of rats. *Evidence-Based Compl. Alternat. Med.*, 3509706.
- Raeeszadeh M., Shokrollahi B., Akbari A. (2022 b). Superior effect of broccoli methanolic extract on control of oxidative damage of sperm cryopreservation and reproductive performance in rats: A comparison with vitamin C and E antioxidant. *Theriogenology*, 181: 50–58.
- Rahman M., Khatun A., Liu L., Barkla B.J. (2018). *Brassicaceae* mustards: Traditional and agronomic uses in Australia and New Zealand. *Molecules*, 23: 1–18.
- Russo R., Reggiani R. (2012). Antinutritive compounds in twelve *Camelina sativa* genotypes. *American J. Plant Sci.*, 3: 1408–1412.
- Salehi B., Quispe C., Butnariu M., Sarac I., Marmouzi I., Kamle M., Martorell M. (2021). Phytotherapy and food applications from *Brassica* genus. *Phytotherapy Res.*, 35: 3590–3609.
- Sánchez-Mata M.C., Cabrera Loera R.D., Morales P., Fernández-Ruiz V., Cámara M., Díez Marqués C., Tardío J. (2012). Wild vegetables of the Mediterranean area as valuable sources of bioactive compounds. *Genetic Res. Crop Evol.*, 59: 431–443.
- Schmidt R., Bancroft I. (2011). Editors. Genetics and genomics of the Brassicaceae. New York: Springer, pp. 585–596.
- Schulz O.E. (1936). Cruciferae. In: Die natürlichen Pflanzenfamilien Band, Engler A., Harms H. (eds). Wilhelm Engelmann Leipzig, 17: 227–658.
- Schuster A., Friedt W. (1998). Glucosinolate content and composition as parameters of quality of camelina seed. *Ind. Crops Prod.*, 7: 297–302.
- Sherman J.M., Hodge H.M. (1936). The bactericidal properties of certain plant juices. *J. Bacteriol.*, 31: 96.
- Slawski H., Adem H., Tressel R.P., Wysujack K., Koops U., Kotzamanis Y., Schulz C. (2012). Total fish meal replacement with rapeseed protein concentrate in diets fed to rainbow trout (*Oncorhynchus mykiss* Walbaum). *Aquacult. Int.*, 20: 443–453.
- Suárez J.A., Gaxiola G., Mendoza R., Cadavid S., García G., Alanís G., Cuzon G. (2009). Substitution of fish meal with plant protein sources and energy budget for white shrimp *Litopenaeus vannamei* (Boone 1931). *Aquaculture*, 289: 118–123.
- Taufik I. (2012). Pendederan ikan nila (*Oreochromis niloticus*) dengan sistem akuaponik pada berbagai lokasi yang berbeda. *Prosiding Indoaqua – Forum Inovasi Teknologi Akuakultur*, p. 10.
- Thiessen D.L., Campbell G.L., Adelizi P.D. (2003). Digestibility and growth performance of juvenile rainbow trout (*Oncorhynchus mykiss*) fed with pea and canola products. *Aquacult. Nutr.*, 9: 67–75.
- Thiessen D.L., Maenz D.D., Newkirk R.W., Classen H.L., Drew M.D. (2004). Replacement of fishmeal by canola protein concentrate in diets fed to rainbow trout (*Oncorhynchus mykiss*). *Aquacult. Nutr.*, 10: 379–388.
- Thomas J., Kuruvilla K.M., Hrideek T.K. (2012). Mustard. In: *Handbook of Herbs and Spices*, Peter K.V. (ed.). Woodhead, pp. 388–398.

- Toyes-Vargas E.A., Parrish C.C., Viana M.T., Carreón-Palau L., Magallón-Servín P., Magallón-Barajas F.J. (2020). Replacement of fish oil with camelina (*Camelina sativa*) oil in diets for juvenile tilapia (var. GIFT *Oreochromis niloticus*) and its effect on growth feed utilization and muscle lipid composition. *Aquaculture*, 523: 735177.
- Turchini G.M., Moretti V.M., Hermon K., Caprino F., Busetto M. L., Bellagamba F., Francis D.S. (2013). Monola oil versus canola oil as a fish oil replacer in rainbow trout feeds: effects on growth fatty acid metabolism and final eating quality. *Food Chem.*, 141: 1335–1344.
- Turchini G.M., Trushenski J.T., Glencross B.D. (2019). Thoughts for the future of aquaculture nutrition: realigning perspectives to reflect contemporary issues related to judicious use of marine resources in aquafeeds. *N. Am. J. Aquacult.*, 81: 13–39.
- Vijayaram S., Sun Y.Z., Zuurro A., Ghafarifarsani H., Van Doan H., Hoseinifar S.H. (2022). Bioactive immunostimulants as health-promoting feed additives in aquaculture: a review. *Fish. Shellfish Immunol.*, 13: 294–308.
- Villa-Cruz V., Davila J., Viana M.T., Vazquez-Duhalt R. (2009). Effect of broccoli (*Brassica oleracea*) and its phytochemical sulforaphane in balanced diets on the detoxification enzymes levels of tilapia (*Oreochromis niloticus*) exposed to a carcinogenic and mutagenic pollutant. *Chemosphere*, 74: 1145–1151.
- Von Danwitz A., Schulz C. (2020). Effects of dietary rapeseed glucosinolates sinapic acid and phytic acid on feed intake growth performance and fish health in turbot (*Psetta maxima* L.). *Aquaculture*, 516: 734624.
- Von Hayek A. (1911). Entwurf eines Cruciferen-systems auf phylogenetischer Grundlage. *Botanisches Centralblatt*, 27: 127–335.
- Wu F., Tian J., Yu L., Wen H., Jiang M., Lu X. (2021). Effects of dietary rapeseed meal levels on growth performance biochemical indices and flesh quality of juvenile genetically improved farmed tilapia. *Aquacult. Rep.*, 20: 100679.
- Yousefi M., Ghafarifarsani H., Hoseini S.M., Hoseinifar S.H., Abtahi B., Vatnikov Y.A., Van Doan H. (2022 a). Effects of dietary thyme essential oil and prebiotic administration on rainbow trout (*Oncorhynchus mykiss*) welfare and performance. *Fish Shellfish Immunol.*, 120: 737–744.
- Yousefi M., Hoseini S.M., Abtahi B., Vatnikov Y.A., Kulikov E.V., Yurievna R.N. (2022 b). Effects of dietary methanolic extract of hyssop, *Hyssopus officinalis*, on growth performance, hepatic antioxidant, humoral and intestinal immunity, and intestinal bacteria of rainbow trout, *Oncorhynchus mykiss*. *Front. Mar. Sci.*, 1962.
- Yousefi M., Hoseini S.M., Kulikov E.V., Seleznev S.B., Petrov A.K., Babichev N.V., Davies S.J. (2022 c). Effects of dietary hyssop, *Hyssopus officinalis*, extract on physiological and antioxidant responses of rainbow trout, *Oncorhynchus mykiss*, juveniles to thermal stress. *Front. Vet. Sci.*, 9.
- Zhou Q.C., Yue Y.R. (2010). Effect of replacing soybean meal with canola meal on growth feed utilization and haematological indices of juvenile hybrid tilapia *Oreochromis niloticus* × *Oreochromis aureus*. *Aquacult. Res.*, 41: 982–990.

Received: 19 XII 2022

Accepted: 4 V 2023