



Epigenetic Regulation of Angiogenesis in Peripheral Artery Disease

REVIEW

NASEEB KAUR MALHI, BMEDSC, PHD

KEVIN W. SOUTHERLAND, MD

LI LAI, PHD

ZHEN BOUMAN CHEN, MB, PHD

*Author affiliations can be found in the back matter of this article

HOUSTON
Methodist
DEBAKEY HEART &
VASCULAR CENTER

ABSTRACT

Peripheral arterial disease (PAD) represents a global health concern with a rising prevalence attributed to factors such as obesity, diabetes, aging, and smoking. Among patients with PAD, chronic limb-threatening ischemia (CLTI) is the most severe manifestation, associated with substantial morbidity and mortality. While revascularization remains the primary therapy for CLTI, not all patients are candidates for such interventions, highlighting the need for alternative approaches. Impaired angiogenesis, the growth of new blood vessels, is a central feature of PAD, and despite decades of research, effective clinical treatments remain elusive. Epigenetics, the study of heritable changes in gene expression, has gained prominence in understanding PAD pathogenesis. Here, we explore the role of epigenetic regulation in angiogenesis within the context of PAD, with a focus on long non-coding RNAs and fibroblast-endothelial cell transdifferentiation. Additionally, we discuss the interplay between metabolic control and epigenetic regulation, providing insights into potential novel therapeutic avenues for improving PAD treatments. This review aims to offer a concise update on the application of epigenetics in angiogenesis and PAD research, inspiring further investigations in this promising field.

CORRESPONDING AUTHORS:

Zhen Bouman Chen, MB, PhD

Beckman Research Institute,
City of Hope, Duarte, California,
US

zhenchen@coh.org

Li Lai, PhD

Houston Methodist Research
Institute, Houston, Texas, US

llai@houstonmethodist.org

KEYWORDS:

epigenetics; peripheral arterial disease (PAD); hindlimb ischemia (HLI); angiogenesis; long non-coding RNAs (lncRNAs); fibroblast-endothelial transdifferentiation

TO CITE THIS ARTICLE:

Malhi NK, Southerland KW, Lai L, Chen ZB. Epigenetic Regulation of Angiogenesis in Peripheral Artery Disease. Methodist DeBakey Cardiovasc J. 2023;19(5):47-57. doi: 10.14797/mdcvj.1294

INTRODUCTION

Peripheral arterial disease (PAD), which is defined as arterial occlusive lesions in the lower extremity, is a major cause of morbidity and mortality. Currently, 200 million individuals worldwide are affected by PAD,¹ with the number predicted to rise as a result of increased detrimental environmental factors such as obesity, diabetes, aging, and smoking.² Approximately 10% of PAD patients present with the most severe clinical manifestation, termed chronic limb-threatening ischemia (CLTI).³ CLTI, which presents as rest pain, nonhealing wounds, or gangrene is a devastating pathology with a 1-year limb loss rate of 25% and a 5-year mortality of 50%.^{4,5} The primary therapy for CLTI patients is surgical or endovascular revascularization.⁶ Despite vast improvements in revascularization strategies, amputation rates remain high. In fact, up to 20% of CLTI patients are not candidates for revascularization attempts based on anatomic constraints.⁷ Hence, there is a great need for molecular therapies aimed at improving perfusion to the ischemic limb.

Among the many damaging functional aspects contributing to PAD, impaired angiogenesis is one of the most prominent and intensively investigated. Angiogenesis is defined as the growth and proliferation of blood vessels from existing vasculature. In the context of PAD, the hypoxic and ischemic environment triggers the release of growth factors that act upon the endothelial cells (ECs), causing them to proliferate, migrate, and elongate to expand the microvasculature. This angiogenic process can be therapeutically enhanced by growth factors such as vascular endothelial growth factor (VEGF). Therapeutic angiogenesis has been investigated for over two decades for its potential clinical use to treat PAD.^{8,9} However, none of these therapies have translated to clinical success yet.

Epigenetics investigates the heritable changes in gene expression that occur without alterations to the underlying DNA sequence. The aforementioned environmental factors currently plaguing the western world, such as obesity and diabetes, are implicated in promoting epigenetic changes. Some of the key molecular processes under investigation by epigenetics include DNA methylation (DNAm), histone modifications, long non-coding RNAs (lncRNAs), and chromatin remodeling. During the induction of angiogenesis, ECs can undergo epigenetic changes, in which the quiescent ECs become activated and mobilized. In the context of PAD, epigenetic modifications have been shown to play a pivotal role in the development and progression of this vascular disorder through influencing the expression of genes involved in not only angiogenesis but also other contributing factors such as inflammation, oxidative stress, and vascular remodeling. Understanding

how epigenetic changes contribute to the onset and progression of PAD not only deepens our comprehension of its underlying mechanisms but also offers potential avenues for therapeutic interventions and personalized treatment strategies.

In this review, we will focus on epigenetic regulation, angiogenesis, and PAD. We will provide an overview of epigenetic regulation and its role in angiogenesis in the context of PAD. Specifically, we will emphasize the role of lncRNAs, which are showing potential as novel targets to achieve therapeutic angiogenesis. We also will discuss fibroblast-EC transdifferentiation, an alternative source of angiogenesis and an epigenetic cell-reprogramming process that may be leveraged to develop therapeutics for PAD. Through discussion of fibroblast-EC transdifferentiation, we will touch upon the connection between metabolic control and epigenetic regulation, two key aspects of cell biology. This review is not meant to be exhaustive nor comprehensive. Instead, we intend to provide readers with a brief update on the application of epigenetics in angiogenesis and PAD research and spur interest in future efforts in leveraging epigenetics to improve PAD treatments.

OVERVIEW OF EPIGENETIC REGULATION

In a broad sense, epigenetics bridges between genotype and phenotype.⁹ For DNAm and histone modifications, there are numerous writers, readers, and erasers—for example, enzymes that mediate the addition, processing, and removal of the modifications. Interested readers are referred to extensive reviews on these topics.^{10–13}

DNA METHYLATION

The first identified epigenetic regulation, DNAm, involves the addition of methyl groups to specific cytosine bases in DNA, typically at CpG dinucleotides, which are enriched in the promoter regions. In most cases, DNAm can repress gene transcription and is often associated with gene silencing. DNAm has been extensively studied in cancer, with the promoter regions of many tumor suppressor genes hypermethylated. Homocysteine, a circulating marker of PAD, is known to mediate DNAm.¹⁴ One prominent drug used to target DNAm is 5-azacytidine, often referred to as 5-aza, a demethylating agent applied to some cancers, including acute myeloid leukemia and myelodysplastic syndrome. Importantly, 5-aza has been shown to ameliorate atherosclerosis through suppressing macrophage inflammation in mice¹⁵; however, the importance of DNAm directly in PAD has been little studied.

HISTONE MODIFICATIONS

Histone modifications encompass over a hundred chemical alterations made to histone proteins, such as acetylation, methylation, phosphorylation, and ubiquitination, to name a few well-studied ones. These histone modifications play a crucial role in regulating gene expression by either loosening or tightening the chromatin structure, thereby controlling the accessibility of genes to transcriptional machinery. Different combinations of these modifications create a “histone code” that governs various cellular processes, and the dysregulation of histone modifications has been implicated in numerous diseases.

Like DNAm, our knowledge of histone modifications has come primarily from cancer research. One of the best-known examples of histone modification and its relevance to disease is the role of histone acetylation in cancer. In many cancer types, there is an aberrant increase in histone deacetylase (HDAC) activity, leading to the removal of acetyl groups from histone proteins. This deacetylation

results in a more condensed chromatin structure, which can silence tumor suppressor genes and promote oncogene expression. HDAC inhibitors have been developed to treat lymphoma and have shown promise in treating other cancer types. A histone deacetylase inhibitor, MCT-1, has shown some promise in rat models of another type of PAD, abdominal aortic aneurysm.¹⁶ However, similar to DNAm, histone modifications in PAD also have been rarely studied.

LONG NON-CODING RNAs

Long non-coding RNAs (lncRNAs) are generally defined as RNA transcripts over 200 nucleotide in length that do not code for proteins but instead function as regulatory molecules. Depending on their subcellular localization, lncRNAs can be nuclear, chromatin-bound or cytoplasmic (Figure 1). Those involved in epigenetic regulation are typically nuclear localized and chromatin-associated, which we refer to as chromatin-associated lncRNAs (ca-lncRNAs). Some of these ca-lncRNAs can interact

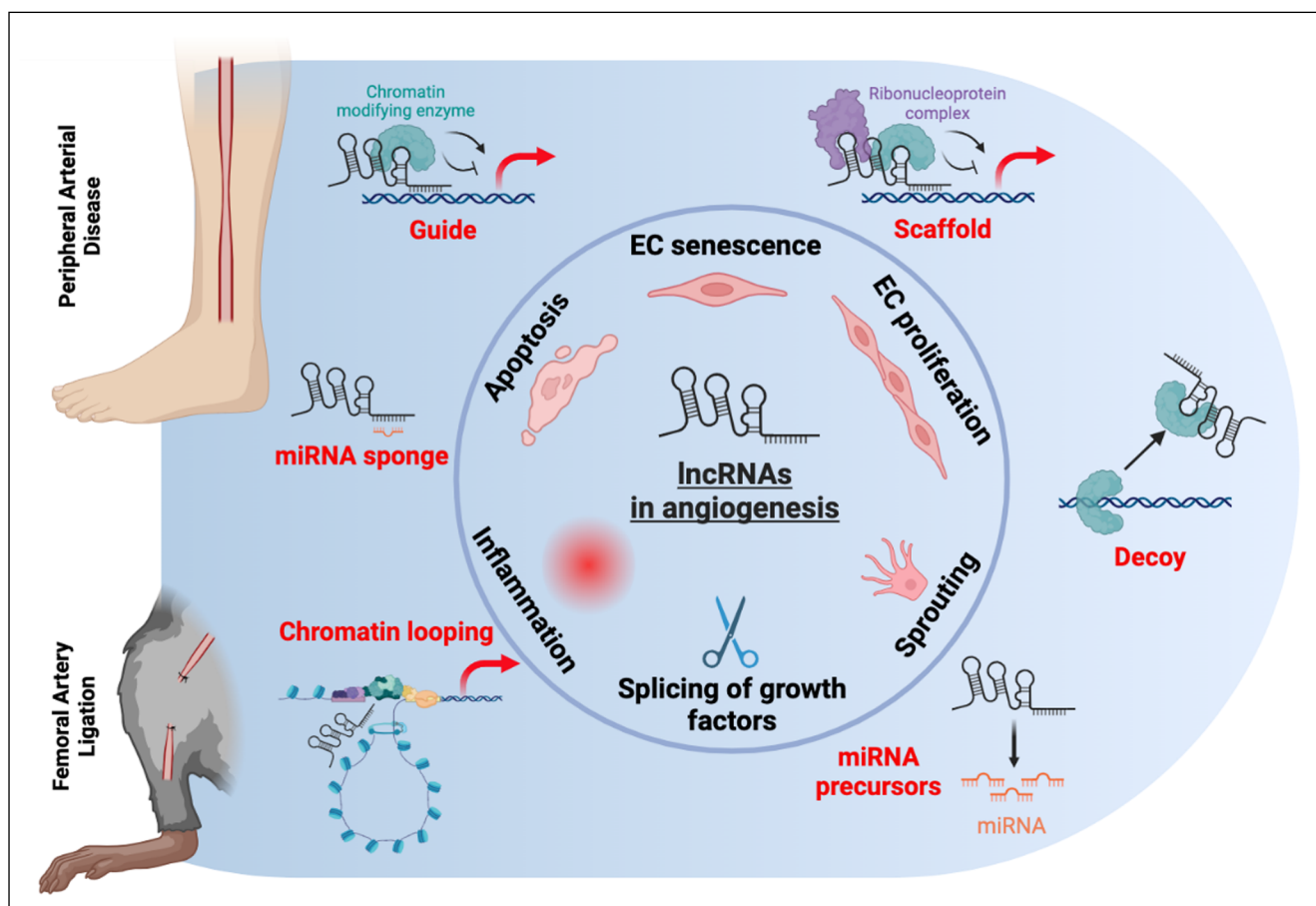


Figure 1 Potential modes of action of lncRNA in angiogenesis in peripheral arterial disease. lncRNAs are regulators of angiogenic pathways in endothelial cells, including their senescence and activation, proliferation, sprouting, and apoptosis. They also may modulate the expression of growth factors via splicing. In addition, they are involved in inflammation, a key driver of angiogenesis. The potential mechanisms through which they deploy their action are guiding, scaffolding, or decoying chromatin modifying enzymes. They also may act as miRNA sponges or precursors and are involved in chromatin structure via looping. lncRNAs: long non-coding RNAs; miRNA: microRNA

with chromatin-modifying complexes, such as histone methyltransferases or chromatin remodelers, to influence the structure and accessibility of chromatin. Other lncRNAs can act as decoys, scaffolds, or guides for transcription factors, RNA polymerases, and other regulatory proteins, helping to fine-tune gene expression. These can either enhance or inhibit transcription depending on the specific lncRNA and its interactions. lncRNAs can also mediate genomic imprinting, a process where specific genes are expressed based on their parental origin. For example, the lncRNA XIST is essential for X-chromosome inactivation in females.^{17,18} Additionally, lncRNAs can help maintain epigenetic memory across cell divisions by recruiting chromatin-modifying enzymes to specific genomic loci, ensuring the stable transmission of epigenetic marks.

A number of excellent reviews summarizing the mode-of-action of lncRNAs and their implications in various diseases are available for interested readers.^{19–22} Pertaining to PAD, a recent review has summarized the role of ncRNAs, including microRNAs and lncRNAs, as functional mediators or biomarkers in the pathophysiology of CLTI.²³ In the next section, we will focus on lncRNAs as epigenetic regulations in angiogenesis in the context of PAD, and we will provide several examples how these lncRNAs can be involved in the pathogenesis of PAD. A better understanding of these interesting regulators may aid in the development of improved therapeutics to treat PAD.

ROLE OF lncRNAs IN ANGIOGENESIS IN PAD

To date, dozens of lncRNAs have been reported to play a regulatory role in angiogenesis (Table 1), and the list is still increasing. Highlighted here are several that have been implicated in PAD due to its regulation of angiogenesis.

MALAT1

Metastasis-Associated Lung Adenocarcinoma Transcript 1 (MALAT1) is one of the most highly studied and conserved lncRNA. It has been associated with numerous disorders, including vascular diseases, and is one of the most abundant lncRNA transcripts in all cells including ECs. Importantly, it was found to promote EC proliferation but reduce EC migration and sprouting. Inhibition of MALAT1 in mice reduced blood flow recovery in femoral artery ligation-induced hindlimb ischemia (HLI), a mouse model of PAD.^{24,25} A global knockout of MALAT1 induced similar results, hypothesized to be mediated through vascular endothelial growth factor receptor 2 binding.²⁶ Moreover, reduced MALAT1 levels were found in human atherosclerotic lesions of symptomatic patients, associated with a

poor prognosis. In macrophages, MALAT1 was found to enhance the biological functions of high glucose-impaired macrophages, leading to improved phagocytosis, a shift towards a pro-healing phenotype, reduced apoptosis, and thus ultimately promoting the healing of diabetic wounds.²⁷ However, another study concluded MALAT1 overexpression increased inflammation and EC dysfunction in diabetes.²⁸ Thus, while it is clear that MALAT1 is involved in EC function and vascular repair, the effect of its modulation may be more context-dependent.

MEG3

Maternally expressed gene 3 (MEG3), transcribed from a maternally imprinted gene, is another highly expressed lncRNA in ECs. Increased in senescent ECs, its inhibition led to improved EC sprouting and migration in vitro and restoration of blood perfusion in mice with HLI.²⁹ This effect is thought to be mediated in part by the Notch signaling pathway and by inducing microRNA-21,³⁰ a positive regulator of angiogenesis.³¹ In the brains of MEG3-knockout mice, there was significantly higher cortical microvessel density compared with controls, coupled with an increased expression of angiogenesis-related genes such as VEGF and Notch1.³² Of note, the DNAm in 14q32 locus, where MEG3 is transcribed from, has been associated with type 2 diabetes, atherosclerosis, and PAD.^{33–35}

ANRIL

Antisense non-coding RNA in the INK4 Locus (ANRIL) is transcribed from human chromosome 9p21,³⁶ a susceptibility locus of coronary artery disease now considered the most robust genetic marker of atherosclerotic cardiovascular disease, including PAD.³⁷ Intriguingly, linear isoforms of ANRIL are associated with increased atherosclerosis susceptibility and an elevated risk of atherosclerotic plaques. In contrast, circular isoforms of ANRIL offer protection against atherosclerotic plaques and mitigate the risk of atherosclerosis.³⁸ The different functions of distinct isoforms of ANRIL may be related to seemingly inconsistent findings regarding the role of ANRIL in EC function. For instance, in the context of chronic kidney disease, ANRIL expression was found to increase with uremia toxin, which also increased expression of inflammatory cytokines in ECs. Inhibition of ANRIL was shown to alleviate endothelial dysfunction in mouse models of chronic kidney disease.³⁹

Consistently, another study showed that knockdown of ANRIL reduced apoptosis and inflammation and increased proliferation and tube formation of human umbilical vein endothelial cells (HUVECs), which was suggested to be mediated by the TGF- β 1/Smad pathway.⁴⁰ A more recent study found that ANRIL regulates the inflammatory

response of HUVECs through regulating alternative splicing.⁴¹ However, inhibition of ANRIL has also been shown to reduce endothelial nitric oxide synthase (eNOS) protein levels and nitric oxide (NO) release from HUVECs by other studies. In fact, ANRIL overexpression was shown to increase eNOS expression and promoted post-ischemic angiogenesis while improving cardiac function in mice following myocardial ischemia.⁴² ANRIL is not conserved in mice, but deficiency of *CDKN2B*, a gene found in the 9p21 locus, showed increased tissue necrosis and reduced blood flow in mice undergoing HLI.⁴³

SNHG12

The evolutionary conserved small nucleolar host gene 12 (SNHG12) was found to be decreased in aortic ECs in atherosclerotic lesions.⁴⁴ The same group then investigated its role in angiogenesis and PAD and found that SNHG12 was decreased in ischemic ECs and increased with perfusion recovery. In mice with SNHG12 inhibition, there was an impaired angiogenic response, which was more pronounced in diabetic (*db/db*) mice, suggesting a positive effect of SNHG12 in angiogenesis and neovascularization. Of note, despite relatively similar knockdown of SNHG12 in both the EC and non-EC fractions, RNA-seq analysis showed different enriched pathways encompassing inflammation and angiogenesis between the two fractions. This points to a need to evaluate the tissue/cell type-specific role of lncRNAs in ischemic response.⁴⁵

LEENE

lncRNA that enhances eNOS expression (LEENE) is a novel regulator in EC biology, angiogenesis, and ischemic response. It is encoded by *LINC00520*, an enhancer region in human ECs. It was found that both the chromatin accessibility and the transcriptional activity of *LINC00520* was enhanced by flow, particularly by shear stress.^{46,47} LEENE expression was also increased by hypoxia in ECs but exhibited a decline under diabetic conditions, evident in cultured ECs, mouse hindlimb muscles, and human arteries. Inhibition of LEENE within human microvascular ECs led to a reduction in their angiogenic capacity, accompanied by disruptions in the normal angiogenic gene program. In a murine model of diabetes, mice lacking the mouse homolog of human LEENE displayed compromised angiogenesis and impaired perfusion after HLI, an experimental model of PAD. Notably, the introduction of human LEENE overexpression successfully rectified the impaired ischemic response in *leene*-knockout mice, both at the level of tissue function and in single-cell transcriptomic analyses. The potential augmentation of LEENE functioned to reinstate angiogenesis, thereby aiding tissue repair and regeneration, may be a promising strategy in addressing PAD.^{48,49}

MESENCHYMAL-EC TRANSDIFFERENTIATION AND METABOLIC REGULATION OF EPIGENETICS

While promoting the angiogenic capacity of existing ECs is a direct approach to enhance angiogenesis and improve blood flow perfusion in ischemic tissues, another promising approach is to enhance angiogenic transdifferentiation from tissue fibroblasts into ECs.⁶⁴ The contribution of such transdifferentiation to vascular regeneration has been demonstrated in the murine HLI model using the fibroblast lineage tracing approach and single cell fate mapping.^{65,66} Through the transdifferentiation process, fibroblasts acquire the gene expression profile and functional characteristics of microvascular ECs.⁶⁴ During this event, thousands of genes are up- or down-regulated, accompanied by a remarkable epigenetic reprogramming.⁶⁷⁻⁶⁹ As a result, the metabolic activity of the transdifferentiated cells becomes drastically different from the original state, signified by a glycolytic switch.

While such metabolic alterations may seem to be passive adaptations as the mesenchymal cells get converted to endothelial lineage, recent studies have pointed to an active role of metabolic modulation on the epigenetic reprogramming of the fibroblasts.⁶⁷ A glycolytic switch was found to occur before the expression of EC markers in transdifferentiating fibroblasts. Inhibiting glycolysis impaired the generation of transdifferentiated ECs, whereas promoting glycolysis enhanced the transdifferentiation. Furthermore, during this process, nuclear ATP-citrate lyase (ACL/ACLY) was increased, along with citrate, which is converted to acetyl-CoA by ACL. Knockdown of ACL/ACLY attenuated both glycolytic switch and the consequent transdifferentiation. These findings suggest a novel role of ACL in angiogenic transdifferentiation and also highlight the link between metabolism and epigenetic modulation in transdifferentiation.

Acetyl-CoA is the substrate for histone acetylation, which increases the accessibility of chromatin to transcriptional activation.⁷⁰⁻⁷³ Thus, the increase in the nuclear ACL can increase the nuclear pool of acetyl-CoA, increasing histone acetylation and promoting the chromatin accessibility necessary for the transdifferentiation. Aside from acetyl-CoA,^{74,75} many other metabolites, such as S-adenosylmethionine, succinate,⁷⁶ lactate,⁷⁷ α -ketoglutarate, and uridine diphosphate N-acetylglucosamine, are also essential building blocks, cofactors, and substrates for epigenetic processes.⁷⁸ Many of these metabolites can couple with epigenetic modifiers to influence chromatin states to maintain or alter cell fate (Figure 2). Therefore, better understanding of the metabolic-epigenetic crosstalk

LNCRNA	CELL SPECIFICITY	RELATIONSHIP TO PAD	PROPOSED MECHANISM
MALAT1 ²⁴⁻²⁸	Multiple cell types including ECs	Reduced blood flow recovery in KO model of HLI, involved in EC proliferation	VEGFR2
MEG3 ^{29,30,32,33}	Multiple cell types including ECs and fibroblasts	Improved blood flow recovery in KO model of HLI, expressed in senescent ECs and reduced during EC sprouting	Notch signaling, miR-21
ANRIL ^{36,38-42}	Multiple cell types including ECs, myocytes and fibroblasts	cdkn2b-deficient mouse under HLI showed reduced blood flow recovery and higher digital amputation	TGF- β R1/Smad pathway, eNOS
SNHG12 ^{44,45}	Multiple cell types including ECs, immune cells and fibroblasts	In KO mice, impaired angiogenic response to HLI, more pronounced in diabetes	Wnt, Notch, and angiotensin signaling pathways
LEENE ⁴⁶⁻⁴⁹	EC enriched	Impaired perfusion recovery in HLI of KO mice, restored with LEENE expression	KDR, eNOS
STEEL ⁵⁰⁻⁵²	Endothelial enriched	Stimulates formation and maturation of vascular flow networks; decreased expression in disturbed flow	KD reduces expression of shear stress related genes KLF2, eNOS; feedback loop with KLF2
H19 ⁵²⁻⁵⁴	Multiple cell types including ECs, myoblasts and PBMCs	Increased in PAD mouse model after 14 days of ischemia, involved in myogenesis, reduced capillary density EC specific-KO model of HLI	STAT3 signaling; IL-6; VEGF
MIR22HG ^{52,54}	Multiple cell types including ECs	Elevated in hypoxic ECs and HLI model	Unknown
LINC00607 ⁵⁵⁻⁵⁷	ECs and VSMCs	Associated SNP found predisposing patients to PAD	c-Myc
HIF1A-AS1 ^{58,59}	VSMCs and ECs	Hypoxia regulated element	BRG1
SENCR ^{62,60}	Highest in ECs but also expressed in VSMCs	Expression is reduced in human critical limb ischemia patients	CX1CL3, CCL5, CEACAM1
FENDRR ⁶¹	ECs and VSMCs	Decreased in hypoxia	DRP1 DNA methylation, p53
HIF1A-AS2 ^{62,63}	VSMCs and ECs	Hypoxia regulated element	USF1 to elevate ATF2

Table 1 Reported long non-coding RNAs with functional relevance in peripheral arterial disease.^{24-30,32,33,36,38-42,44-63} ECs: endothelial cells; KO: knockout; HLI: hindlimb ischemia; miR: microRNA; eNOS: endothelial nitric oxide synthase; KDR: kinase insert domain receptor; PBMCs: peripheral blood mononuclear cells; PAD: peripheral artery disease; VEGF: vascular endothelial growth factor; SNP: single nucleotide polymorphisms; VSMCs: vascular smooth muscle cells; BRG1: Brahma-related gene 1

can provide valuable insights and therapeutic strategies to promote angiogenic transdifferentiation and tissue regeneration.

CONCLUSION AND PERSPECTIVE

The emerging field of epigenetic regulation in angiogenesis holds immense promise in shedding light on the intricate mechanisms underlying PAD. The dynamic interplay between epigenetic modifications and the angiogenic processes within the vascular system has unveiled new avenues for understanding the pathogenesis of PAD and exploring innovative therapeutic strategies. For example, diabetes is a prominent risk factor for PAD, and the hyperglycemic condition is well known to induce heritable epigenetic changes. Elucidating how diabetic conditions may modulate epigenome supporting the angiogenic function of ECs may facilitate the design of more

effective therapies to intervene in diabetic PAD. Similarly, understanding how epigenetic variations contribute to PAD heterogeneity may enable ways to stratify patients into distinct subgroups, each with tailored treatment approaches. In addition, understanding the mechanisms underlying the metabolic-epigenetic control of fibroblast-EC transdifferentiation may make it possible to augment the cell plasticity and maximize the transdifferentiation potential, which is desirable in regenerative medicine for PAD. While many drugs targeting the epigenome are being tested for treatment of cancer and other cardiovascular diseases, future research is warranted to determine the promise of using these epigenetic drugs in the treatment of PAD.

As we deepen our comprehension of the epigenetic regulatory mechanisms, we are poised to develop targeted interventions that may mitigate the progression of PAD and restore vascular function to improve the quality of life for those affected by this debilitating condition. This

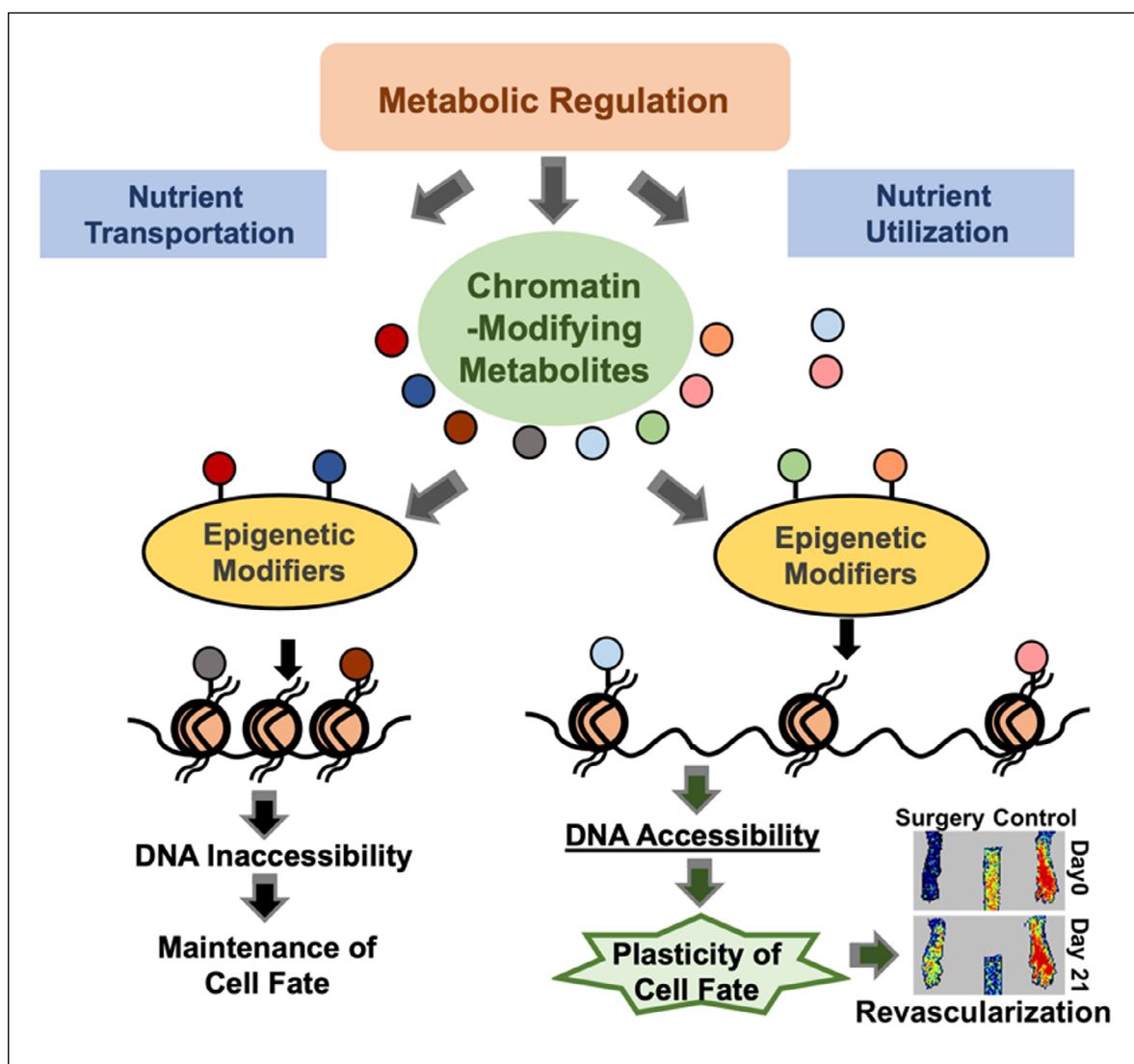


Figure 2 Metabolic regulation on epigenetics. The chromatin-modifying metabolites, regulated by their rate-limiting enzymes and the availability of their precursors, can bind to histone tails or epigenetic modifiers to affect chromatin accessibility, which further modulates the cell fate transition.

intersection of epigenetics and angiogenesis in PAD research represents a promising frontier in the quest for more effective treatments and enhanced patient outcomes.

KEY POINTS

- Epigenetic regulatory mechanisms are important modulators of angiogenesis that may be harnessed to treat peripheral arterial disease.
- Histone modifications and DNA methylation regulate the epigenome to effect endothelial angiogenic capacity but require further study in peripheral arterial disease.
- Long non-coding RNAs are involved in most, if not all, steps of endothelial angiogenesis, from expression of growth factors to sprouting. Targeting these

transcripts has recently shown therapeutic potential.

- Chromatin binding metabolites can alter the epigenome, to either maintain or alter cell fate, such as in fibroblast-EC transdifferentiation.

AUTHOR CONTRIBUTIONS

Li Lai and Zhen Bouman Chen are co-corresponding authors.

FUNDING INFORMATION

This work is funded in part by R01HL145170 from the NIH, Ella Fitzgerald Foundation, and a City of Hope-HBCU pilot project (to ZBC) and R56HL169204-01 (to LL).

COMPETING INTERESTS

The authors have no competing interests to declare.

AUTHOR AFFILIATIONS

Naseeb Kaur Malhi, BMedSc, PhD  orcid.org/0000-0002-8981-2974

Beckman Research Institute City of Hope, Duarte, California, US

Kevin W. Southerland, MD  orcid.org/0000-0002-0306-2130
Duke University Medical Center, Durham, North Carolina, US

Li Lai, PhD  orcid.org/0000-0002-5731-2705

Houston Methodist Research Institute, Houston, Texas, US

Zhen Bouman Chen, MB, PhD  orcid.org/0000-0002-3291-1090

Beckman Research Institute City of Hope, Duarte, California, US

REFERENCES

- Fowkes FG, Rudan D, Rudan I**, et al. Comparison of global estimates of prevalence and risk factors for peripheral artery disease in 2000 and 2010: a systematic review and analysis. *Lancet*. 2013 Oct 19;382(9901):1329-40. doi: [10.1016/S0140-6736\(13\)61249-0](https://doi.org/10.1016/S0140-6736(13)61249-0)
- Marso SP, Hiatt WR**. Peripheral arterial disease in patients with diabetes. *J Am Coll Cardiol*. 2006 Mar 7;47(5):921-9. doi: [10.1016/j.jacc.2005.09.065](https://doi.org/10.1016/j.jacc.2005.09.065)
- Nehler MR, Duval S, Diao L**, et al. Epidemiology of peripheral arterial disease and critical limb ischemia in an insured national population. *J Vasc Surg*. 2014 Sep;60(3):686-95 e2. doi: [10.1016/j.jvs.2014.03.290](https://doi.org/10.1016/j.jvs.2014.03.290)
- Duff S, Mafilios MS, Bhounsule P, Hasegawa JT**. The burden of critical limb ischemia: a review of recent literature. *Vasc Health Risk Manag*. 2019 Jul 1;15:187-208. doi: [10.2147/VHRM.S209241](https://doi.org/10.2147/VHRM.S209241)
- Farber A**. Chronic Limb-Threatening Ischemia. *N Engl J Med*. 2018 Jul 12;379(2):171-80. doi: [10.1056/NEJMc1709326](https://doi.org/10.1056/NEJMc1709326)
- Farber A, Menard MT, Conte MS**, et al. Surgery or Endovascular Therapy for Chronic Limb-Threatening Ischemia. *N Engl J Med*. 2022 ec 22;387(25):2305-16. doi: [10.1056/NEJMoa2207899](https://doi.org/10.1056/NEJMoa2207899)
- Shishehbor MH, Powell RJ, Montero-Baker MF**, et al. Transcatheter Arterialization of Deep Veins in Chronic Limb-Threatening Ischemia. *N Engl J Med*. 2023 Mar 30;388(13):1171-80. doi: [10.1056/NEJMoa2212754](https://doi.org/10.1056/NEJMoa2212754)
- Iyer SR, Annex BH**. Therapeutic Angiogenesis for Peripheral Artery Disease: Lessons Learned in Translational Science. *JACC Basic Transl Sci*. 2017 Oct;2(5):503-12. doi: [10.1016/j.jacbts.2017.07.012](https://doi.org/10.1016/j.jacbts.2017.07.012)
- Cooke JP, Losordo DW**. Modulating the vascular response to limb ischemia: angiogenic and cell therapies. *Circ Res*. 2015 Apr 24;116(9):1561-78. doi: [10.1161/CIRCRESAHA.115.303565](https://doi.org/10.1161/CIRCRESAHA.115.303565)
- Chen Z, Li S, Subramaniam S, Shyy JY, Chien S**. Epigenetic Regulation: A New Frontier for Biomedical Engineers. *Annu Rev Biomed Eng*. 2017 Jun 21;19:195-219. doi: [10.1146/annurev-bioeng-071516-044720](https://doi.org/10.1146/annurev-bioeng-071516-044720)
- Morselli M, Dieci G**. Epigenetic regulation of human non-coding RNA gene transcription. *Biochem Soc Trans*. 2022 Apr 29;50(2):723-36. doi: [10.1042/BST20210860](https://doi.org/10.1042/BST20210860)
- Allis CD, Jenuwein T**. The molecular hallmarks of epigenetic control. *Nat Rev Genet*. 2016 Aug;17(8):487-500. doi: [10.1038/nrg.2016.59](https://doi.org/10.1038/nrg.2016.59)
- Fitz-James MH, Cavalli G**. Molecular mechanisms of transgenerational epigenetic inheritance. *Nat Rev Genet*. 2022 Jun;23(6):325-41. doi: [10.1038/s41576-021-00438-5](https://doi.org/10.1038/s41576-021-00438-5)
- Krishna SM, Dear A, Craig JM, Norman PE, Golledge J**. The potential role of homocysteine mediated DNA methylation and associated epigenetic changes in abdominal aortic aneurysm formation. *Atherosclerosis*. 2013 Jun;228(2):295-305. doi: [10.1016/j.atherosclerosis.2013.02.019](https://doi.org/10.1016/j.atherosclerosis.2013.02.019)
- Cao Q, Wang X, Jia L**, et al. Inhibiting DNA Methylation by 5-Aza-2'-deoxycytidine ameliorates atherosclerosis through suppressing macrophage inflammation. *Endocrinology*. 2014 Dec;155(12):4925-38. doi: [10.1210/en.2014-1595](https://doi.org/10.1210/en.2014-1595)
- Vinh A, Gaspari TA, Liu HB, Dousha LF, Widdop RE, Dear AE**. A novel histone deacetylase inhibitor reduces abdominal aortic aneurysm formation in angiotensin II-infused apolipoprotein E-deficient mice. *J Vasc Res*. 2008;45(2):143-52. doi: [10.1159/000110041](https://doi.org/10.1159/000110041)
- Marahrens Y, Panning B, Dausman J, Strauss W, Jaenisch R**. Xist-deficient mice are defective in dosage compensation but not spermatogenesis. *Genes Dev*. 1997 Jan 15;11(2):156-66. doi: [10.1101/gad.11.2.156](https://doi.org/10.1101/gad.11.2.156)
- Penny GD, Kay GF, Sheardown SA, Rastan S, Brockdorff N**. Requirement for Xist in X chromosome inactivation. *Nature*. 1996 Jan 11;379(6561):131-7. doi: [10.1038/379131a0](https://doi.org/10.1038/379131a0)
- Statello L, Guo CJ, Chen LL, Huarte M**. Author Correction: Gene regulation by long non-coding RNAs and its biological functions. *Nat Rev Mol Cell Biol*. 2021 Feb;22(2):159. doi: [10.1038/s41580-021-00330-4](https://doi.org/10.1038/s41580-021-00330-4)
- Kopp F, Mendell JT**. Functional Classification and Experimental Dissection of Long Noncoding RNAs. *Cell*. 2018 Jan 25;172(3):393-407. doi: [10.1016/j.cell.2018.01.011](https://doi.org/10.1016/j.cell.2018.01.011)
- Rinn JL, Chang HY**. Long Noncoding RNAs: Molecular Modalities to Organismal Functions. *Annu Rev Biochem*. 2020 Jun 20;89:283-308. doi: [10.1146/annurev-biochem-062917-012708](https://doi.org/10.1146/annurev-biochem-062917-012708)
- Mattick JS, Amaral PP, Carninci P**, et al. Long non-coding RNAs: definitions, functions, challenges and recommendations. *Nat Rev Mol Cell Biol*. 2023 Jun;24(6):430-47. doi: [10.1038/s41580-022-00566-8](https://doi.org/10.1038/s41580-022-00566-8)
- Perez-Cremades D, Cheng HS, Feinberg MW**. Noncoding RNAs in Critical Limb Ischemia. *Arterioscler Thromb Vasc Biol*. 2020 Mar;40(3):523-33. doi: [10.1161/ATVBAHA.119.312860](https://doi.org/10.1161/ATVBAHA.119.312860)
- Michalik KM, You X, Manavski Y**, et al. Long noncoding RNA MALAT1 regulates endothelial cell function and vessel

- growth. *Circ Res*. 2014 Apr 25;114(9):1389-97. doi: [10.1161/CIRCRESAHA.114.303265](https://doi.org/10.1161/CIRCRESAHA.114.303265)
25. **Zhang X, Tang X, Hamblin MH, Yin KJ.** Long Non-Coding RNA Malat1 Regulates Angiogenesis in Hindlimb Ischemia. *Int J Mol Sci*. 2018 Jun 11;19(6). doi: [10.3390/ijms19061723](https://doi.org/10.3390/ijms19061723)
 26. **Cremer S, Michalik KM, Fischer A,** et al. Hematopoietic Deficiency of the Long Noncoding RNA MALAT1 Promotes Atherosclerosis and Plaque Inflammation. *Circulation*. 2019 Mar 5;139(10):1320-34. doi: [10.1161/CIRCULATIONAHA.117.029015](https://doi.org/10.1161/CIRCULATIONAHA.117.029015)
 27. **Kuang L, Zhang C, Li B, Deng H, Chen R, Li G.** Human Keratinocyte-Derived Exosomal MALAT1 Promotes Diabetic Wound Healing by Upregulating MFG8 via microRNA-1914-3p. *Int J Nanomedicine*. 2023 Feb 21;18:949-70. doi: [10.2147/IJN.S399785](https://doi.org/10.2147/IJN.S399785). eCollection 2023
 28. **Liu JY, Yao J, Li XM,** et al. Pathogenic role of lncRNA-MALAT1 in endothelial cell dysfunction in diabetes mellitus. *Cell Death Dis*. 2014 Oct 30;5(10):e1506. doi: [10.1038/cddis.2014.466](https://doi.org/10.1038/cddis.2014.466)
 29. **Boon RA, Hofmann P, Michalik KM,** et al. Long Noncoding RNA Meg3 Controls Endothelial Cell Aging and Function: Implications for Regenerative Angiogenesis. *J Am Coll Cardiol*. 2016 Dec 13;68(23):2589-91. doi: [10.1016/j.jacc.2016.09.949](https://doi.org/10.1016/j.jacc.2016.09.949)
 30. **Wu Z, He Y, Li D,** et al. Long noncoding RNA MEG3 suppressed endothelial cell proliferation and migration through regulating miR-21. *Am J Transl Res*. 2017 Jul 15;9(7):3326-35. PMID: 28804550
 31. **Liu LZ, Li C, Chen Q,** et al. MiR-21 induced angiogenesis through AKT and ERK activation and HIF-1 α expression. *PLoS One*. 2011 Apr 22;6(4):e19139. doi: [10.1371/journal.pone.0019139](https://doi.org/10.1371/journal.pone.0019139)
 32. **Gordon FE, Nutt CL, Cheunsuchon P,** et al. Increased expression of angiogenic genes in the brains of mouse meg3-null embryos. *Endocrinology*. 2010 Jun;151(6):2443-52. doi: [10.1210/en.2009-1151](https://doi.org/10.1210/en.2009-1151)
 33. **Kameswaran V, Bramswig NC, McKenna LB,** et al. Epigenetic regulation of the DLK1-MEG3 microRNA cluster in human type 2 diabetic islets. *Cell Metab*. 2014 Jan 17;19(1):135-45. doi: [10.1016/j.cmet.2013.11.016](https://doi.org/10.1016/j.cmet.2013.11.016)
 34. **Aavik E, Lumivuori H, Leppanen O,** et al. Global DNA methylation analysis of human atherosclerotic plaques reveals extensive genomic hypomethylation and reactivation at imprinted locus 14q32 involving induction of a miRNA cluster. *Eur Heart J*. 2015 Apr 21;36(16):993-1000. doi: [10.1093/eurheartj/ehu437](https://doi.org/10.1093/eurheartj/ehu437)
 35. **Heuslein JL, Gorick CM, Price RJ.** Epigenetic regulators of the revascularization response to chronic arterial occlusion. *Cardiovasc Res*. 2019 Mar 15;115(4):701-12. doi: [10.1093/cvr/cvz001](https://doi.org/10.1093/cvr/cvz001)
 36. **Broadbent HM, Peden JF, Lorkowski S,** et al. Susceptibility to coronary artery disease and diabetes is encoded by distinct, tightly linked SNPs in the ANRIL locus on chromosome 9p. *Hum Mol Genet*. 2008 Mar 15;17(6):806-14. doi: [10.1093/hmg/ddm352](https://doi.org/10.1093/hmg/ddm352)
 37. **Ye S, Willeit J, Kronenberg F, Xu Q, Kiechl S.** Association of genetic variation on chromosome 9p21 with susceptibility and progression of atherosclerosis: a population-based, prospective study. *J Am Coll Cardiol*. 2008 Jul 29;52(5):378-84. doi: [10.1016/j.jacc.2007.11.087](https://doi.org/10.1016/j.jacc.2007.11.087)
 38. **Razeghian-Jahromi I, Karimi Akhormeh A, Zibaenezhad MJ.** The Role of ANRIL in Atherosclerosis. *Dis Markers*. 2022 Feb 9;2022:8859677. doi: [10.1155/2022/8859677](https://doi.org/10.1155/2022/8859677)
 39. **Su H, Liu B, Chen H,** et al. LncRNA ANRIL mediates endothelial dysfunction through BDNF downregulation in chronic kidney disease. *Cell Death Dis*. 2022 Jul 29;13(7):661. doi: [10.1038/s41419-022-05068-1](https://doi.org/10.1038/s41419-022-05068-1)
 40. **Liu X, Li S, Yang Y,** et al. The lncRNA ANRIL regulates endothelial dysfunction by targeting the let-7b/TGF- β R1 signalling pathway. *J Cell Physiol*. 2021 Mar;236(3):2058-69. doi: [10.1002/jcp.29993](https://doi.org/10.1002/jcp.29993)
 41. **Wufuer A, Luohemanjiang X, Du L,** et al. ANRIL overexpression globally induces expression and alternative splicing of genes involved in inflammation in HUVECs. *Mol Med Rep*. 2023 Feb;27(2). doi: [10.3892/mmr.2022.12915](https://doi.org/10.3892/mmr.2022.12915)
 42. **Huang Q, Pan M, Zhou JP, Yin F.** Overexpression of long non-coding RNA ANRIL promotes post-ischaemic angiogenesis and improves cardiac functions by targeting Akt. *J Cell Mol Med*. 2020 Jun;24(12):6860-8. doi: [10.1111/jcmm.15343](https://doi.org/10.1111/jcmm.15343)
 43. **Nanda V, Downing KP, Ye J,** et al. CDKN2B Regulates TGF β Signaling and Smooth Muscle Cell Investment of Hypoxic Neovessels. *Circ Res*. 2016 Jan 22;118(2):230-40. doi: [10.1161/CIRCRESAHA.115.307906](https://doi.org/10.1161/CIRCRESAHA.115.307906)
 44. **Haemmig S, Yang D, Sun X,** et al. Long noncoding RNA SNHG12 integrates a DNA-PK-mediated DNA damage response and vascular senescence. *Sci Transl Med*. 2020 Feb 19;12(531). doi: [10.1126/scitranslmed.aaw1868](https://doi.org/10.1126/scitranslmed.aaw1868)
 45. **Gross DA, Cheng HS, Zhuang R,** et al. Deficiency of lncRNA SNHG12 impairs ischemic limb neovascularization by altering an endothelial cell cycle pathway. *JCI Insight*. 2022 Jan 11;7(1). doi: [10.1172/jci.insight.150761](https://doi.org/10.1172/jci.insight.150761)
 46. **Miao Y, Ajami NE, Huang TS,** et al. Enhancer-associated long non-coding RNA LEENE regulates endothelial nitric oxide synthase and endothelial function. *Nat Commun*. 2018 Jan 18;9(1):292. doi: [10.1038/s41467-017-02113-y](https://doi.org/10.1038/s41467-017-02113-y)
 47. **Stanicek L, Lozano-Vidal N, Bink DI,** et al. Long non-coding RNA LASSIE regulates shear stress sensing and endothelial barrier function. *Commun Biol*. 2020 May 26;3(1):265. doi: [10.1038/s42003-020-0987-0](https://doi.org/10.1038/s42003-020-0987-0)
 48. **Tang X, Luo Y, Yuan D,** et al. Long noncoding RNA LEENE promotes angiogenesis and ischemic recovery in diabetes models. *J Clin Invest*. 2023 Feb 1;133(3). doi: [10.1172/JCI161759](https://doi.org/10.1172/JCI161759)

49. **Kallapur A, Sallam T.** Endothelial cells LEENE on noncoding RNAs in diabetic vasculopathy. *J Clin Invest.* 2023 Feb 1;133(3). doi: [10.1172/JCI167047](https://doi.org/10.1172/JCI167047)
50. **Man HSJ, Sukumar AN, Lam GC,** et al. Angiogenic patterning by STEEL, an endothelial-enriched long noncoding RNA. *Proc Natl Acad Sci U S A.* 2018 Mar 6;115(10):2401-6. doi: [10.1073/pnas.1715182115](https://doi.org/10.1073/pnas.1715182115)
51. **Zhang SZ, Lu ZF, Xu YJ, Shi HF, Liu YZ, Rui YJ.** STEEL participates in fracture healing through upregulating angiogenesis-related genes by recruiting PARP 1. *Eur Rev Med Pharmacol Sci.* 2018 Jun;22(12):3669-75. doi: [10.26355/eurev_201806_15245](https://doi.org/10.26355/eurev_201806_15245)
52. **Yu B, Wang S.** Angio-LncRs: LncRNAs that regulate angiogenesis and vascular disease. *Theranostics.* 2018 Jun 8;8(13):3654-75. doi: [10.7150/thno.26024](https://doi.org/10.7150/thno.26024)
53. **Hofmann P, Sommer J, Theodorou K,** et al. Long non-coding RNA H19 regulates endothelial cell aging via inhibition of STAT3 signalling. *Cardiovasc Res.* 2019 Jan 1;115(1):230-42. doi: [10.1093/cvr/cvy206](https://doi.org/10.1093/cvr/cvy206)
54. **Voellenkle C, Garcia-Manteiga JM, Pedrotti S,** et al. Implication of Long noncoding RNAs in the endothelial cell response to hypoxia revealed by RNA-sequencing. *Sci Rep.* 2016 Apr 11;6:24141. doi: [10.1038/srep24141](https://doi.org/10.1038/srep24141)
55. **Boos F, Oo JA, Warwick T,** et al. The endothelial-enriched lncRNA LINC00607 mediates angiogenic function. *Basic Res Cardiol.* 2023 Jan 26;118(1):5. doi: [10.1007/s00395-023-00978-3](https://doi.org/10.1007/s00395-023-00978-3)
56. **Calandrelli R, Xu L, Luo Y,** et al. Stress-induced RNA-chromatin interactions promote endothelial dysfunction. *Nat Commun.* 2020 Oct 15;11(1):5211. doi: [10.1038/s41467-020-18957-w](https://doi.org/10.1038/s41467-020-18957-w)
57. **Cullina S, Wojcik GL, Shemirani R,** et al. Admixture Mapping of Peripheral Artery Disease in a Dominican Population Reveals a Novel Risk Locus on 2q35. *Front Genet.* 2023 Aug 1;14:1181167. doi: [10.3389/fgene.2023.1181167](https://doi.org/10.3389/fgene.2023.1181167)
58. **He Q, Tan J, Yu B, Shi W, Liang K.** Long noncoding RNA HIF1A-AS1A reduces apoptosis of vascular smooth muscle cells: implications for the pathogenesis of thoracoabdominal aorta aneurysm. *Pharmazie.* 2015 May;70(5):310-5. PMID: 26062299
59. **Wang S, Zhang X, Yuan Y,** et al. BRG1 expression is increased in thoracic aortic aneurysms and regulates proliferation and apoptosis of vascular smooth muscle cells through the long non-coding RNA HIF1A-AS1 in vitro. *Eur J Cardiothorac Surg.* 2015 Mar;47(3):439-46. doi: [10.1093/ejcts/ezu215](https://doi.org/10.1093/ejcts/ezu215)
60. **Boulberdaa M, Scott E, Ballantyne M,** et al. A Role for the Long Noncoding RNA SENCN in Commitment and Function of Endothelial Cells. *Mol Ther.* 2016 May;24(5):978-90. doi: [10.1038/mt.2016.41](https://doi.org/10.1038/mt.2016.41)
61. **Wang X, Li Q, He S,** et al. LncRNA FENDRR with m6A RNA methylation regulates hypoxia-induced pulmonary artery endothelial cell pyroptosis by mediating DRP1 DNA methylation. *Mol Med.* 2022 Oct 25;28(1):126. doi: [10.1186/s10020-022-00551-z](https://doi.org/10.1186/s10020-022-00551-z)
62. **Li P, Xing J, Zhang J,** et al. Inhibition of long noncoding RNA HIF1A-AS2 confers protection against atherosclerosis via ATF2 downregulation. *J Adv Res.* 2020 Jul 31;26:123-35. doi: [10.1016/j.jare.2020.07.015](https://doi.org/10.1016/j.jare.2020.07.015)
63. **Shu L, Wang C, Ding Z,** et al. A novel regulated network mediated by downregulation HIF1A-AS2 lncRNA impairs placental angiogenesis by promoting ANGPTL4 expression in preeclampsia. *Front Cell Dev Biol.* 2022 Aug 9;10:837000. doi: [10.3389/fcell.2022.837000](https://doi.org/10.3389/fcell.2022.837000)
64. **Sayed N, Wong WT, Ospino F,** et al. Transdifferentiation of human fibroblasts to endothelial cells: role of innate immunity. *Circulation.* 2015 Jan 20;131(3):300-9. doi: [10.1161/CIRCULATIONAHA.113.007394](https://doi.org/10.1161/CIRCULATIONAHA.113.007394)
65. **Meng S, Lv J, Chanda PK, Owusu I, Chen K, Cooke JP.** Reservoir of Fibroblasts Promotes Recovery From Limb Ischemia. *Circulation.* 2020 Oct 27;142(17):1647-62. doi: [10.1161/CIRCULATIONAHA.120.046872](https://doi.org/10.1161/CIRCULATIONAHA.120.046872)
66. **Cooke JP, Lai L.** Role of angiogenic transdifferentiation in vascular recovery. *Front Cardiovasc Med.* 2023 May 2;10:1155835. doi: [10.3389/fcvm.2023.1155835](https://doi.org/10.3389/fcvm.2023.1155835)
67. **Lai L, Reineke E, Hamilton DJ, Cooke JP.** Glycolytic Switch Is Required for Transdifferentiation to Endothelial Lineage. *Circulation.* 2019 Jan 2;139(1):119-33. doi: [10.1161/CIRCULATIONAHA.118.035741](https://doi.org/10.1161/CIRCULATIONAHA.118.035741)
68. **Meng S, Zhou G, Gu Q, Chanda PK, Ospino F, Cooke JP.** Transdifferentiation Requires iNOS Activation: Role of RING1A S-Nitrosylation. *Circ Res.* 2016 Oct 14;119(9):e129-e38. doi: [10.1161/CIRCRESAHA.116.308263](https://doi.org/10.1161/CIRCRESAHA.116.308263)
69. **Liu C, Ruan H, Himmati F,** et al. HIF1α Regulates Early Metabolic Changes due to Activation of Innate Immunity in Nuclear Reprogramming. *Stem Cell Reports.* 2020 Feb 11;14(2):192-200. doi: [10.1016/j.stemcr.2020.01.006](https://doi.org/10.1016/j.stemcr.2020.01.006)
70. **Feron O.** The many metabolic sources of acetyl-CoA to support histone acetylation and influence cancer progression. *Ann Transl Med.* 2019 Dec;7(Suppl 8):S277. doi: [10.21037/atm.2019.11.140](https://doi.org/10.21037/atm.2019.11.140)
71. **Bannister AJ, Kouzarides T.** Regulation of chromatin by histone modifications. *Cell Res.* 2011 Mar;21(3):381-95. doi: [10.1038/cr.2011.22](https://doi.org/10.1038/cr.2011.22)
72. **Eberharter A, Becker PB.** Histone acetylation: a switch between repressive and permissive chromatin. Second in review series on chromatin dynamics. *EMBO Rep.* 2002 Mar;3(3):224-9. doi: [10.1093/embo-reports/kvf053](https://doi.org/10.1093/embo-reports/kvf053)
73. **Sterner DE, Berger SL.** Acetylation of histones and transcription-related factors. *Microbiol Mol Biol Rev.* 2000 Jun;64(2):435-59. doi: [10.1128/MMBR.64.2.435-459.2000](https://doi.org/10.1128/MMBR.64.2.435-459.2000)
74. **Lei I, Tian S, Gao W,** et al. Acetyl-CoA production by specific metabolites promotes cardiac repair after myocardial

- infarction via histone acetylation. *Elife*. 2021 Dec 23;10:e60311. doi: [10.7554/eLife.60311](https://doi.org/10.7554/eLife.60311)
75. **Campbell SL, Wellen KE.** Metabolic Signaling to the Nucleus in Cancer. *Mol Cell*. 2018 Aug 2;71(3):398-408. doi: [10.1016/j.molcel.2018.07.015](https://doi.org/10.1016/j.molcel.2018.07.015)
76. **Jing Y, Ding D, Tian G,** et al. Semisynthesis of site-specifically succinylated histone reveals that succinylation regulates nucleosome unwrapping rate and DNA accessibility. *Nucleic Acids Res*. 2020 Sep 25;48(17):9538-49. doi: [10.1093/nar/gkaa663](https://doi.org/10.1093/nar/gkaa663)
77. **Zhang D, Tang Z, Huang H,** et al. Metabolic regulation of gene expression by histone lactylation. *Nature*. 2019 Oct;574(7779):575-80. doi: [10.1038/s41586-019-1678-1](https://doi.org/10.1038/s41586-019-1678-1)
78. **Sakabe K, Wang Z, Hart GW.** Beta-N-acetylglucosamine (O-GlcNAc) is part of the histone code. *Proc Natl Acad Sci U S A*. 2010 Nov 16;107(46):19915-20. doi: [10.1073/pnas.1009023107](https://doi.org/10.1073/pnas.1009023107)

TO CITE THIS ARTICLE:

Malhi NK, Southerland KW, Lai L, Chen ZB. Epigenetic Regulation of Angiogenesis in Peripheral Artery Disease. *Methodist DeBakey Cardiovasc J*. 2023;19(5):47-57. doi: [10.14797/mdcvj.1294](https://doi.org/10.14797/mdcvj.1294)

Submitted: 18 September 2023 **Accepted:** 19 September 2023 **Published:** 16 November 2023

COPYRIGHT:

© 2023 The Author(s). This is an open-access article distributed under the terms of the Attribution-NonCommercial 4.0 International (CC BY-NC 4.0), which permits unrestricted use, distribution, and reproduction in any noncommercial medium, provided the original author and source are credited. See <https://creativecommons.org/licenses/by-nc/4.0/>.

Methodist DeBakey Cardiovascular Journal is a peer-reviewed open access journal published by Houston Methodist DeBakey Heart & Vascular Center.