

Fluxes and production of N₂O, CO₂ and CH₄ in boreal agricultural soil during winter as affected by snow cover

By M. MALJANEN^{1*}, A.-R. KOHONEN¹, P. VIRKAJÄRVI² and P. J. MARTIKAINEN¹

¹*University of Kuopio, Department of Environmental Science, P.O. Box 1627, FI-70211 Kuopio, Finland;* ²*MTT, Agrifood Research Finland, Animal Production Research, Halolantie 31 A, FI-71750 Maaninka, Finland*

(Manuscript received 31 October 2006; in final form 21 June 2007)

ABSTRACT

Agricultural soils are important source of atmospheric nitrous oxide (N₂O) and a considerable part of annual N₂O release occurs during the cold season in the boreal region. According to recent studies N₂O can be produced in soils at low temperatures, below 0 °C. We studied if removal of the snowcover lowers soil temperatures and whether this would affect flux rates of N₂O, carbon dioxide (CO₂) and methane (CH₄) from an agricultural soil in eastern Finland. Gas flux rates and concentrations in soil were measured from study plots with undisturbed snow cover and from plots with snow removed. This experiment simulates changes in the soil thermal conditions with less snowfall. Plots without snow had even 15 °C lower temperature at the depth of 5 cm and they had higher N₂O emissions during soil freezing and thawing. However, there were only minor changes in CH₄ or CO₂ flux rates after removal of snow over the cold season. N₂O and CO₂ accumulated in the soil during winter and were then released rapidly during thawing in spring. CH₄ concentrations in the soil remained lower than the atmospheric levels during winter and subsequently increased to the ambient levels after thawing. Future climate scenarios suggest possible decline in snowfall in northern Europe resulting in lower soil temperatures. This could lead to higher N₂O emissions from boreal agricultural soils.

1. Introduction

Greenhouse gases, nitrous oxide (N₂O), carbon dioxide (CO₂) and methane (CH₄) are produced and consumed in soils in microbial processes. These microbial processes are greatly regulated by soil chemical and physical characteristics. Production of CO₂ (respiration) and CH₄ is regulated, for example, by temperature and soil moisture as are denitrification and nitrification, the latter two being the key processes for N₂O production in soil. Agricultural soils are the most important global sources of N₂O and the concentration of N₂O in the atmosphere is increasing annually at the rate of 0.2–0.3% (Houghton et al., 2001). Also the concentration of CH₄ in the atmosphere is increasing (Houghton et al., 2001). Soils, for example, forest soils and drained agricultural soils, may act as sinks for atmospheric CH₄ whereas waterlogged soils (e.g. wetlands and rice fields) emit CH₄.

There are studies suggesting that large amount of N₂O can be produced and emitted from agricultural and forest soils even at temperatures below 0 °C (Holtan-Hartwig et al., 2002; Öquist

et al., 2004; Groffman et al., 2006; Koponen et al., 2006a). Emissions of N₂O during winter are important because they may account for more than half of the annual emissions in boreal and temperate areas (e.g. Röver et al., 1998; Teepe et al., 2000; van Bochove et al., 2000; Schürmann et al., 2002; Maljanen et al., 2003a; Regina et al., 2004).

The main mechanism for N₂O production in frozen soils is presumably denitrification (Mørkved et al., 2006; Öquist et al., 2007). N₂O can be produced in unfrozen water films in an anaerobic soil matrix in frozen soil (Teepe et al., 2001), and the production is limited by NO₃[−] availability (Mørkved et al., 2006; Öquist et al., 2007). High N₂O emissions during winter have been explained by sudden release of accumulated N₂O (Goodroad and Keeney, 1984; van Bochove et al., 2001) or by increasing denitrification activity due to high availability of organic C and N during freezing-thawing events (e.g. Christensen and Tiedje, 1990). It is also possible that at low temperature N₂O reductase is suppressed increasing the production of N₂O over that of N₂ (Holtan-Hartwig et al., 2002). However, the reasons for high N₂O emissions during winter and the production of N₂O in frozen soils are not well understood.

Snow is a good insulator regarding soil temperature during winter and therefore removal of snow can enhance soil freezing

*Corresponding author.
e-mail: marja.maljanen@uku.fi
DOI: 10.1111/j.1600-0889.2007.00304.x

and amplify the depth of frost (Schürmann et al., 2002). According to the recent climate scenarios the snow cover in the boreal region may decrease (Houghton et al., 2001) resulting in lower soil temperature.

Our hypothesis in this study was that without the insulating snow cover (lower soil temperature) N_2O emissions would increase and CO_2 production (respiration) would decrease as compared to the soil with a natural snow cover. The snow cover limits diffusion of CH_4 from the atmosphere into the soil and therefore thinner snow cover may enhance soil CH_4 uptake (Borken et al., 2006). We examined how snow cover affects the flux rates of N_2O , CO_2 and CH_4 by keeping half of the study plots free from snow during one winter period.

2. Methods

2.1. Study site and experimental design

The study site is located in eastern Finland (63°09'N, 27°20'E). The soil type according to FAO classification is Dystric Regosol, medium textured. The soil profile (0–25 cm) consisted mainly of fine sand (71%), silt (15%), and clay (6%). Organic matter content is 5.65% and soil pH (H_2O) is 5.8 in the 0–10 cm of the soil profile. Mean annual temperature (1971–2000) in the region is 2.8 °C and mean annual precipitation is 609 mm, of which approximately 50% falls as snow (Drebs et al., 2002). The long-term (1971–2000) average maximum snow depth has been 50 cm in the middle of March (Drebs et al., 2002). The study was conducted on timothy—meadow fescue lawn (*Phleum pratense* and *Festuca pratensis*, Huds.) which was fertilized in May 2004 with 90 kg N ha⁻¹, 15 kg P ha⁻¹, 25 kg K ha⁻¹ and in July 2004 with 96 N kg ha⁻¹.

There were four experimental plots in the study area of which two were kept free from snow (referred to as bare soil). The area of one plot was 3 × 6 m. Snow was removed from the bare plots with a spade on each day the snow height exceeded about 5 cm. The remaining two plots had an undisturbed snow cover. There were three gas sampling points on each plot.

2.2. Environmental variables

Soil frost was measured using frost-depth gauge filled with methylene blue (Gandahl, 1957) and soil temperatures (depth 5 cm) were recorded with iButton® temperature loggers (Dallas Semiconductor Corp., USA). Air temperature was recorded at a weather station, located 100 m from the study site.

Soil samples for the analysis of NO_3^- and NH_4^+ were collected at 0–10 cm depth six times (28 October, 12 November and 20 December 2004, 2 February, 18 April and 2 June 2005). NO_3^- was extracted with distilled water and NH_4^+ with 1 M KCl solution. NO_3^- was analysed by an ion chromatograph and NH_4^+ with a spectrophotometer (Fawcett and Scott, 1960).

2.3. Gas measurements

Gas flux measurements started in the snow-free period in October 2004 and continued weekly to biweekly until early June 2005. N_2O , CO_2 and CH_4 flux rates were measured with a static chamber method using aluminium chambers, equipped with a fan, (60 × 60 cm, h 30 cm) and aluminium collars (60 × 60 cm, h 15 cm) pre-installed in the soil (Maljanen et al., 2003a). After closing the chamber a gas sample was taken with a polypropylene syringes at 5, 10, 15 and 25 min intervals from the headspace of the chamber. Gas concentrations were analysed within 24 h after sampling with a gas chromatograph (Shimadzu GC-14B, Shimadzu Corp. Japan) equipped with electron capture (EC), thermal conductivity (TC) and flame ionization (FI) detectors (see Maljanen et al., 2003a). Compressed air containing 1.98 $\mu\text{l l}^{-1}$ of CH_4 , 396 $\mu\text{l l}^{-1}$ of CO_2 and 0.389 $\mu\text{l l}^{-1}$ of N_2O was used for daily calibration. Gas fluxes were calculated from the linear change in gas concentrations in the headspace of the chamber. The regression coefficient r^2 was computed for the sample sets from each chamber. If the r^2 was low (<0.8) or if there were any indications of failures in the gas sampling or gas analysis the results were discharged. However, if fluxes were negligible (especially during the midwinter) the r^2 was always low and it was not used as a criterion for omission whenever such conditions occurred.

Gas flux rates from the snow-covered soil were determined by measuring gas concentration gradients and by calculating associated diffusion rates in the snow (Sommerfeld et al., 1993; Maljanen et al., 2003b). Gas samples (40 ml) from the snow pack were drawn with a stainless steel probe (Ø 3 mm, length 50 cm). For the calculation of the diffusive fluxes, an ambient gas sample was taken above the snow pack and another sample inside the snow pack 2 cm above the soil surface. The linearity of gas gradient in the snow pack was checked during each sampling time by sampling gas at 10 cm intervals from the top of snow pack to the soil surface. Simultaneously, snow samples were collected with a PVC tube (Ø 10.2 cm, h 55 cm) for porosity measurements. The intact samples were weighed for calculation of the average porosity of snow using the density of pure ice (0.9168 g cm⁻³).

Samples (30 ml) for the soil gas concentration measurement were taken with syringes from silicon tubes (Ø 1.0 cm, wall thickness 0.3 cm, length 110 cm, $V = 86 \text{ cm}^3$) inserted horizontally at the depth of 5 cm in the soil (Kamman et al., 2001) close to the collars for the gas flux measurements. Gas samples were analysed as described above.

2.4. Statistical methods

The differences in the gas fluxes between the treatments were tested with Mann–Whitney *U*-test (SPSS statistical package 11.5). Correlations between gas concentrations and

environmental parameters were tested with nonparametric Spearman rank correlation test.

3. Results

3.1. Weather conditions and soil properties

Snow started to accumulate in late November in 2004, reached a maximum height of 43 cm in early February 2005 and it thawed totally by mid-April (Fig. 1). Mean daily air temperature reached a minimum of -19.2°C in February (Fig. 1). The early winter from November to January was mild; monthly mean temperatures remained $3.2\text{--}5.5^\circ\text{C}$ higher than the long term average (Drebs et al., 2002). The first time topsoil (1–2 cm) frost was detected in early November. The topsoil remained continuously frozen from late November 2004 until April 2005 (Fig. 1). The soil frost was more severe in areas without snow, reaching a maximum depth of 104 cm in the middle of April. In contrast, under undisturbed snow cover the maximum depth of frost was 20 cm in late March (Fig. 1). Soil temperature (depth 5 cm) in the area with snow cover was almost constant (from -0.8 to 0.8) during the winter, whereas in bare plots it varied from -15.0 to

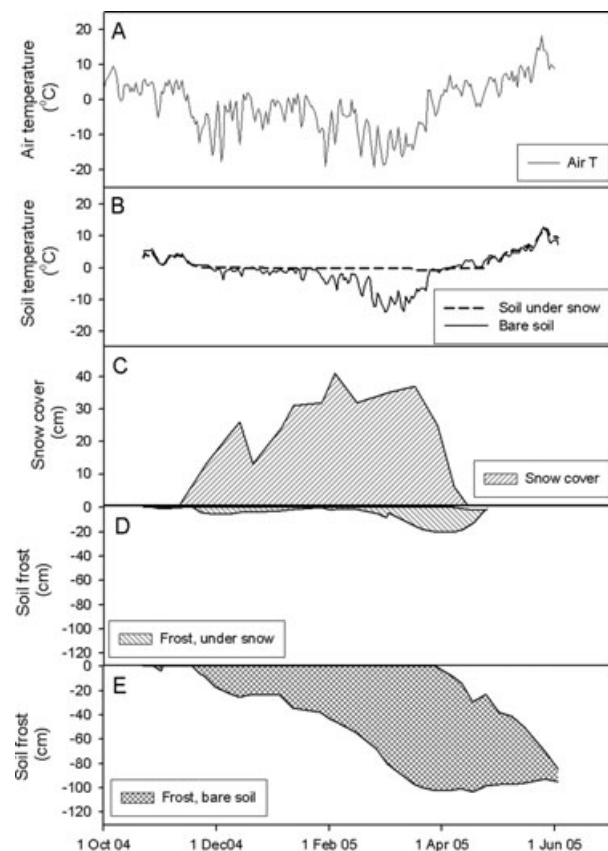


Fig. 1. Mean daily air temperature (A), soil temperature at depth of 5 cm (B), depth of snow cover (C), depth of soil frost under snow cover (D) and depth of soil frost on bare soil without snow (E).

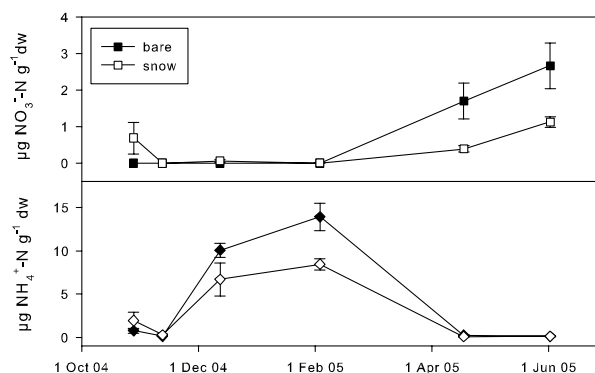


Fig. 2. Concentrations of NO_3^- -N (squares) and NH_4^+ -N (diamonds) in soil sampled from plots under snow (open symbols) or from plots without snow (black symbols).

0.2°C during the same period (Fig. 1). Removal of the snow cover affected also the survival of plants on the study plots, most perennial plants growing on bare plots died during the winter.

Soil NH_4^+ concentrations were low in the unfrozen soil ($<2\text{ mg NH}_4\text{-N g}^{-1}$), increased after freezing up to $14.0\text{ mg NH}_4\text{-N g}^{-1}$ in bare soil and up to $8.4\text{ mg NH}_4\text{-N g}^{-1}$ under snow during the coldest months and decreased again to $<0.10\text{ mg NH}_4\text{-N g}^{-1}$ after thawing (Fig. 2). NO_3^- concentrations were low, $<0.01\text{ mg NO}_3\text{-N g}^{-1}$, during the winter but increased after thawing to $2.6\text{ mg NO}_3\text{-N g}^{-1}$ in bare soil and to $1.1\text{ mg NO}_3\text{-N g}^{-1}$ in soil under snow (Fig. 2).

3.2. Gas fluxes

The bare plots emitted more N_2O than those with an undisturbed snow cover during soil freezing (a period of 3 weeks) ($p < 0.001$) and during thawing (a period of 8 weeks) ($p < 0.001$) (Fig. 3a). High emissions during freezing lasted about 3 weeks in November and December. Following high fluxes, N_2O emissions from all plots remained low (no statistical differences between the treatments). During thawing in April, N_2O emissions increased more from the bare plots, and remained at higher level until the end of the study period (Fig. 3a). The mean N_2O emissions from the bare plots during freezing ($23.5\text{ }\mu\text{g m}^{-2}\text{h}^{-1}$) and thawing ($44.6\text{ }\mu\text{g m}^{-2}\text{h}^{-1}$) were 35 and 4 times those from the plots under snow cover.

In contrast to N_2O emissions, CO_2 emissions were low during soil freezing (Fig. 3c). Also, CO_2 emissions from soils under snow and from the bare plots were low in mid-winter (Fig. 3c). When the soil started to thaw in April, CO_2 emissions increased first from the bare plots and a week later from the snow-covered plots (Fig. 3c). In late May and early June the soil with snow cover emitted more CO_2 than the bare plots (Fig. 3c).

The average CH_4 flux rate from all plots was negative during the study period implying that the soil was a small CH_4 sink (Fig. 3e). During the coldest part of the winter and spring thaw

there were no statistical differences in the CH₄ fluxes between the treatments.

3.3. Gas concentrations in soil

The N₂O concentrations in soil probes started to increase from an ambient concentration of 0.3 $\mu\text{l l}^{-1}$ after the topsoil was frozen (Fig. 3b) and reached a maximum of 30 $\mu\text{l l}^{-1}$ in the bare plots and 5 $\mu\text{l l}^{-1}$ in plots with snow. After thawing, they decreased rapidly close to the ambient levels. N₂O concentration in soil with snow cover correlated positively with the development of soil frost ($r = 0.722$, $p = 0.002$) whereas in bare plots it did not. N₂O concentration under snow cover correlated also with the depth of snow ($r = 0.786$, $p < 0.001$).

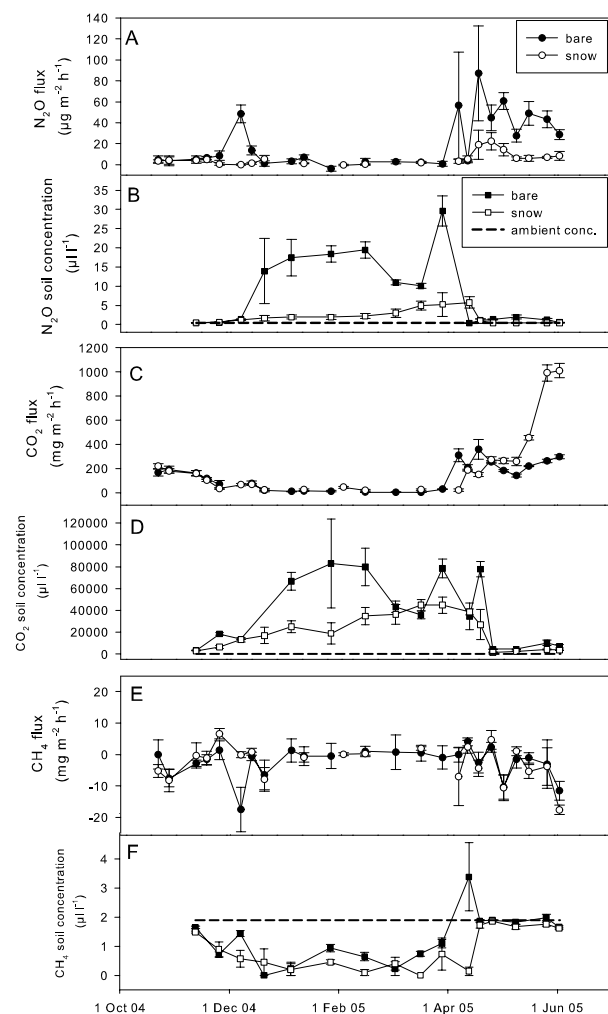


Fig. 3. Flux rates of N₂O (A), CO₂ (C) and CH₄ (E). Values are mean with standard error of six replicates. Concentration of N₂O (B), CO₂ (D) and CH₄ (F) in soil at depth of 5 cm. Values are mean with standard error of three replicates. Dashed lines indicate the ambient concentrations of gases.

Table 1. Mean concentrations ($\mu\text{l l}^{-1}$) of N₂O and CO₂ in unfrozen soil in autumn and in frozen soils with snow or without snow (bare) from January to February. Ratio U/F is the relative increase in the gas concentrations (concentration in unfrozen soil/concentration in frozen soil)

	N ₂ O		CO ₂	
	Concentration	Ratio U/F	Concentration	Ratio U/F
Unfrozen	0.455		3023	
Frozen (snow)	2.27	3.98	28 700	8.51
Frozen (bare)	16.1	35.4	65 100	21.5

As with N₂O, CO₂ also accumulated in the frozen soil (Fig. 3d). In bare soil the relative concentration of N₂O (the increase from the concentration before freezing) increased after soil freezing more than the relative concentration of CO₂. However, in the soil with snow the reverse was true (Table 1). The maximum concentration in bare plots was 80 and 45 ml l⁻¹ CO₂ under the snow cover. After thawing, CO₂ concentrations in soil decreased rapidly to a level of 2.0 ml l⁻¹ (with snow) and 6.5 ml l⁻¹ (bare soil). The concentration of CO₂ under snow-covered soil correlated positively with the depth of soil frost ($r = 0.771$, $p < 0.001$) and with the depth of snow cover ($r = 0.690$, $p = 0.003$), but it did not correlate with the depth of frost in bare plots.

CH₄ concentration began to decrease after the topsoil was frozen and it remained at a level lower than the ambient concentration during the winter months. After thawing CH₄ concentration in soil increased close to the ambient level of 1.8 $\mu\text{l l}^{-1}$. During thawing there was a slight increase in CH₄ concentration in the bare soil (Fig. 3e).

4. Discussion

4.1. Soil properties

Removal of snow resulted in more severe soil frost and lower soil temperatures than those under natural snow cover. Under the snow pack the top soil temperature remained relatively constant during winter months supporting the observations reported in previous studies (e.g. Schürmann et al., 2002). The death of vegetation on bare plots illustrates the importance of insulating function of snow cover for perennial plant species. It is likely that mechanical shovelling itself was not the reason for the death of *Phleum pratense* and *Festuca pratensis* plants because the overwintering parts of these plants are located under the soil surface (Virkajärvi and Järvenranta, 2001).

NH₄⁺ concentration increased during the winter indicating N mineralization also in frozen soil. Nitrogen mineralization was expected to occur because decomposition of organic matter continued in frozen soil as indicated by the CO₂ production and CO₂ accumulation in soil (see later). NO₃⁻ concentration

remained low during the mid-winter. It is possible that part of the accumulated NH₄⁺ is nitrified but NO₃⁻ is then used rapidly in denitrification. Therefore, the availability of NO₃⁻ is an important factor limiting denitrification rate in frozen soil (Öquist et al., 2007).

4.2. N₂O and CO₂ emissions

The bare plots had higher mean N₂O emissions and lower soil temperature compared to plots with natural snow cover supporting studies from forest soils by Groffman et al. (2006). Many field and laboratory studies have shown high emissions only during soil thawing (e.g. Christensen and Tiedje, 1990; Teepe et al., 2000; Regina et al., 2004; Syväsalö et al., 2006) but there are only few field (Kammann et al., 1998; Röver et al., 1998; Teepe et al., 2000) and laboratory studies (Teepe et al., 2001; Koponen et al., 2004, 2006a) that have reported high N₂O emission pulses during soil freezing. In our study, enhanced N₂O emissions were seen not only during soil thawing in the spring but also during soil freezing in the autumn. The freezing related N₂O pulse occurred only from the bare plots with lower soil temperature.

Enhanced denitrification in bare plots may be related to more severe frost causing the death of some microbes and therefore release of microbially available organic carbon which stimulates respiration and depletion of O₂ (Christensen and Tiedje, 1990; Mørkved et al., 2006) thus providing favourable conditions for denitrification. Enhanced emissions could also result from changes in the microbial community. However, there are some contradictory results on whether freezing and thawing can change the microbial community structure in soil (Koponen et al., 2006b; Sharma et al., 2006), but evidently thawing increases microbial activity in soil as seen here from the increased CO₂ production.

Thawing related N₂O emission pulse was seen on bare and snow plots. In bare plots during thawing there was first an N₂O emission pulse simultaneously with a rapid decrease in the N₂O concentration in soil. This first pulse is a result of release of trapped gas from the soil (Goodroad and Keeney, 1984; Teepe et al., 2001). The second N₂O pulse occurred in April when the snow was melting but there was still frost deeper in the soil. The high soil moisture probably favoured denitrification and N₂O production after soil thawing (Goodroad and Keeney, 1984). The higher N₂O emissions from bare plots in the early summer may result from higher availability of NO₃⁻ because all vegetation had died and did not consume NO₃⁻.

The removal of snow altered the soil physical–biological conditions and this affected the CO₂ emissions after thawing. The CO₂ emissions in the autumn and during the winter period until thawing were similar in the plots under snow cover and the bare plots. During thaw, enhanced CO₂ emissions were first seen from the bare soil when the topsoil (2–3 cm) was thawing and then a few weeks later after snowmelt from the soils under snow. In late May and early June the CO₂ emissions from the soil which had

been under snow were higher than those from the bare soil. This could be a result from higher availability of plant derived easily available carbon (root material and root exudates) and higher respiration in soil with snow than in soil without snow—on the bare plots almost all plants had died as a result of a severe frost.

4.3. N₂O and CO₂ concentrations in soil

During the coldest months the emissions of N₂O and CO₂ were low from all plots. This was a result of an ice barrier in the top of the soil limiting the gas diffusion from the soil (van Bochove et al., 2001). The early winter in December and January was mild and there were several rain showers in periods when the top soil was already frozen, which promoted formation of an ice layer. The difference in the accumulation of N₂O and CO₂ in the soil indicates that the decrease in temperature resulting from the snow removal favours more the production of N₂O than that of CO₂ (the conclusion on the production can be done from the gas accumulation because both of the gases were efficiently blocked in soil with only minor release to the atmosphere). Koponen et al. (2004) showed from laboratory experiments that the low temperature conditions favour more the production of N₂O (denitrification) than the microbial respiration. Denitrifying microbes can be active even at temperatures as low as –4 °C (Röver et al., 1998; Öquist et al., 2004; Schmidt and Lipson, 2004; Phillips, 2007) resulting from the unfrozen liquid contained in soil microsites (Teepe et al., 2001; Sparrman et al., 2004).

Concentrations of N₂O and CO₂ in frozen soil were high, but close to those reported by van Bochove et al. (2001). Most of the N₂O was accumulated in the bare plots within 2 weeks after the soil freezing. Mean temperature at 5 cm depth was –0.88 °C during that period. N₂O concentrations increased only slowly after that period until mid-March. When soil temperature dropped down to –15 °C, there was a decrease in the N₂O concentration. This can be a result of lower N₂O production in soil with less unfrozen water and physical release of N₂O via cracks in the soil. Concentration of CO₂ had similar trend as that of N₂O. We do not know why concentration of N₂O and CO₂ decreased in late winter when the frost was still developing. There was no evidence of gas release to the atmosphere during that period. The gases were released from soil just after the frost was disappearing. It has to be noted that in soil with snow there was a continuous accumulation of N₂O and CO₂ gases until thawing even the concentrations in soil were lower than in bare soil. Under snow the soil temperature was at the range where even 30% of soil water can remain unfrozen (Edwards and Cresser, 1992) allowing microbial activity.

4.4. CH₄ fluxes and concentrations

There were no changes in the CH₄ fluxes after removal of snow in contrast to the results of Groffman et al. (2006) or Borken et al. (2006). However, the CH₄ uptake rates in our study were

low when compared to the forest soils. Therefore, the possible minor effect of the snow removal on the low fluxes here could be difficult to verify. Methane concentration in soil was below the atmospheric CH₄ concentration also during the winter indicating CH₄ oxidation even during the coldest period of the year.

5. Conclusions

The results here show how the possible reduction in snow cover thickness with global warming would affect the greenhouse gas dynamics of boreal agricultural soil, for example, the timing and magnitude of the high emissions can be changed. It is important that the thinner snow pack with lower soil temperatures can enhance the N₂O production during the cold periods of the year. The results stress the importance of the N₂O production at low soil temperatures, also below 0 °C. Furthermore, the N₂O production in winter does not correlate well with the N₂O emissions as a result of the low gas diffusion rate in frozen soil. Therefore, the production of N₂O during winter cannot be estimated only by measuring N₂O release to the atmosphere.

6. Acknowledgments

This work was funded by Maj- and Tor Nessling Foundation and Niemi Foundation. Pekka Issakainen from Agrifood Research Finland is thanked for the assistance in the field. Dr. Narasinha Shurpali is acknowledged for commenting the manuscript.

References

- Borken, W., Davidson, E. A., Savage, K., Sundquist, E. T. and Steudler, P. 2006. Effect of summer throughfall exclusion, summer drought, and winter snow cover on methane fluxes in a temperate forest soil. *Soil Biol. Biochem.* **38**, 1388–1395.
- Christensen, S. and Tiedje, J. M. 1990. Brief and vigorous N₂O production by soil at spring thaw. *J. Soil Sci.* **41**, 1–4.
- Drebs, A., Norlund, A., Karlsson, P., Helminen, J. and Rissanen, P. 2002. *Climatological Statistics of Finland 1971–2000*, No. 2002:1, Finnish Meteorological Institute, Edita Prima Oy, Helsinki, 99 p.
- Edwards, A. C. and Cresser, M. S. 1992. Freezing and its effects on chemical and biological properties of soil. *Adv. Soil Sci.*, **18**, 59–79.
- Fawcett, J. K. and Scott, J. E. 1960. A Rapid and precise method for the determination of urea. *J. Clin. Path.* **13**, 156–159.
- Gandahl, R. 1957. Bestämning av tjälgräns i mark med enkel typ av tjälgränsmätare. *Grundförbättring* **10**, 7–19. (In Swedish)
- Goodroad, L. L. and Keeney, D. R. 1984. Nitrous oxide emission from soils during thawing. *Can. J. Soil Sci.* **64**, 187–194.
- Groffman, P. M., Hardy, J. P., Scott, N., Fitzhugh, R. D., Driscoll, C. T., and co-authors. 2006. Snow depth, soil freezing and fluxes of carbon dioxide, nitrous oxide and methane in a northern hardwood forest. *Global Change Biol.* **12**, 1748–1760.
- Holtan-Hartwig, L., Dörsch, P. and Bakken, L. R. 2002. Low temperature control of soil denitrifying communities: kinetics of N₂O production and reduction. *Soil Biol. Biochem.* **34**, 1797–1806.
- Houghton, J. T., Ding, Y., Griggs, D. J., Noguer, M., van der Linden, P. J., and co-authors. 2001. *Climate Change 2001: The Scientific Basis. Contribution of Working Group I to the Third Assessment Report of the Intergovernmental Panel on Climate Change*. Cambridge University Press, Cambridge. 881 p.
- Kammann, C., Gründhage, L., Müller, C., Jacobi, S. and Jäger, H. J. 1998. Seasonal variability and mitigation options for N₂O emissions from differently managed grasslands. *Environ. Pollut.* **102 S1**, 179–186.
- Kammann, C., Gründhage, L. and Jäger, H. J. 2001. A new sampling technique to monitor concentrations of CH₄, N₂O and CO₂ in air at well-defined depths in soils with varied water potential. *Eur. J. Soil Sci.* **52**, 297–303.
- Koponen, H. T., Flöjt, L. and Martikainen, P. J. 2004. Nitrous oxide emissions from agricultural soils at low temperatures: a laboratory microcosm study. *Soil Biol. Biochem.* **36**, 757–766.
- Koponen, H. T., Escude Duran, C., Maljanen, M., Hytönen, J. and Martikainen, P. J. 2006a. Temperature responses of NO and N₂O emissions from boreal organic soil. *Soil Biol. Biochem.* **38**, 1779–1787.
- Koponen, H. T., Jaakkola, T., Keinänen-Toivola, M. M., Kaipainen, S., Tuomainen, J., and co-authors. 2006b. Microbial communities, biomass and activities in soils as affected by freeze thaw cycles. *Soil Biol. Biochem.* **38**, 1861–1871.
- Maljanen, M., Liikanen, A., Silvola, J. and Martikainen, P. J. 2003a. Nitrous oxide emissions from boreal organic soil under different land-use. *Soil Biol. Biochem.* **35**, 689–700.
- Maljanen, M., Liikanen, A., Silvola, J. and Martikainen, P. J. 2003b. Measuring N₂O emissions from organic soils with closed chamber or gas gradient methods. *Eur. J. Soil Sci.* **54**, 625–631.
- Mørkved, P. T., Dörsch, P., Henriksen, T. M. and Bakken, L. R. 2006. N₂O emissions and product ratios of nitrification and denitrification as affected by freezing and thawing. *Soil Biol. Biochem.* **38**, 3411–3420.
- Öquist, M. G., Nilsson, M., Sörensson, F., Kasimir-Klemetsson, Å., Persson, T., and co-authors. 2004. Nitrous oxide production in a forest soil at low temperatures-processes and environmental controls. *FEMS Microbiol. Ecol.* **46**, 371–378.
- Öquist, M. G., Petrone, K., Nilsson, M., Persson, and Klemetsson, L. 2007. Nitrification controls N₂O production rates in frozen boreal forest soil. *Soil Biol. Biochem.* **39**, 1809–1811.
- Phillips, R. L. 2007. Organic agriculture and nitrous oxide emissions at sub-zero temperatures. *J. Environ. Qual.* **36**, 23–30.
- Regina, K., Syväsalö, E., Hannukkala, A. and Esala, M. 2004. Fluxes of N₂O from farmed peat soils in Finland. *Eur. J. Soil Sci.* **55**, 591–599.
- Röver, M., Heinemeyer, O. and Kaiser, E. A. 1998. Microbial induced nitrous oxide emissions from an arable soil during winter. *Soil Biol. Biochem.* **30**, 1859–1865.
- Schmidt, S. K. and Lipson, D. A. 2004. Microbial growth under the snow: Implications for nutrient and allelochemical availability in temperate soils. *Plant Soil* **259**, 1–7.
- Schürmann, A., Mohn, J. and Bachofen, R. 2002. N₂O emissions from snow-covered soils in the Swiss Alps. *Tellus* **54B**, 134–142.
- Sharma, S., Szele, Z., Schilling, R., Munch, J. C. and Schlöter, M. 2006. Influence of freeze-thaw stress on the structure and function of microbial communities and denitrifying populations in soil. *Appl. Environ. Microb.* **72**, 2184–2154.

- Sommerfeld, R. A., Mosier, A. R. and Musselman, R. C. 1993. CO₂, CH₄ and N₂O flux through a Wyoming snowpack and implications for global budgets. *Nature* **361**, 140–142.
- Sparrman, T., Öquist, M., Klemetsson, L., Schleucher, J. and Nilsson, M. 2004. Quantifying unfrozen water in frozen soil by high-field ²H NMR. *Environ. Sci. & Technol.* **38**, 5420–5425.
- Syväsalo, E., Regina, K., Turtola, E., Lemola, R. and Esala, M. 2006. Fluxes of nitrous oxide and methane, and nitrogen leaching from organically and conventionally cultivated sandy soil in western Finland. *Agricult. Ecosys. Environ.* **113**, 342–348.
- Teepe, R., Brumme, R. and Beese, F. 2000. Nitrous oxide from frozen soils under agricultural, fallow and forest. *Soil Biol. Biochem.* **32**, 1807–1810.
- Teepe, R., Brumme, R. and Beese, F. 2001. Nitrous oxide emissions from soil during freezing and thawing periods. *Soil Biol. Biochem.* **33**, 1269–1275.
- van Bochove, E., Jones, H. G., Bertrand, N. and Prévost, D. 2000. Winter fluxes of greenhouse gases from snow-covered agricultural soil: intra-annual and interannual variations. *Global Biogeochem. Cy.* **14**, 113–125.
- van Bochove, E., Thériault, G. and Rochette, P. 2001. Thick ice layers in snow and frozen soil affecting gas emissions from agricultural soils during winter. *J. Geophys. Res.* **106**, 23061–23071.
- Virkajärvi, P. and Järvenranta, K. 2001. Leaf dynamics of timothy and meadow fescue under Nordic conditions. *Grass Forage Sci.* **56**, 294–304.