

MEDICAL AND ECONOMICAL SIGNIFICANCE OF MILLIPEDES

Monika Jung¹⁾, Barbara Nieradko-Iwanicka²⁾, Grzegorz Kania²⁾

¹⁾ Chair of Pharmacology and Biology, Medical University in Lublin, Poland

²⁾ Hygiene and Epidemiology Department, Medical University in Lublin, Poland

ABSTRACT

Millipedes as saprophagous epigeic fauna members are involved in the decomposition of organic debris. Diplopod's activity improve circulation of nutrients, elements and accelerate environmental reclamation. Beyond mentioned above Diplopoda can provoke sanitary and epidemiological threats especially during swarming or seasonal migrations activities. Considering following aspects: contact with defensive secretions, pseudo-parasitic activity that imitates helminthiasis or passive transmission of pathogens, we can discuss direct or distant effects on human health. Education and proper prophylactic remain neglected. Variety of substances from millipedes are checked for their medical action: direct body extracts and active chemical components from defensive secretion. Millipedes may serve as source for zoo-pharmacology and zoo-indication development. This issue review shows that both pros and cons of the Diplopod's influences should be considered. Literature was carried out from middle of 20th century article till recent papers. The nowadays literature were found in Google Scholar, PubMed, Medline search base.

Keywords: Millipedes, Diplopoda, secretions, medical importance, antibiotic and antifungal activity.

ARTICLE INFO

PolHypRes 2023 Vol. 82 Issue 1 pp. 115 – 125

ISSN: 1734-7009 **eISSN:** 2084-0535

DOI: 10.2478/phr-2023-0007

Pages: 11, figures: 1, tables: 0

page www of the periodical: www.phr.net.pl

Review article

Submission date: 15.01.2022 r.

Acceptance for print: 24.01.2023 r.

Publisher

Polish Hyperbaric Medicine and Technology Society

INTRODUCTION

Both importance in nature and species diversity rank millipedes just after Insecta and Arachnida [1,2,3]. Diplopoda are the numerous taxon of Myriapoda, that cover approximately 12.000 described species gathered in 145 families and 16 orders [4,5,6,7,8]. Millipedes as epigeic litter dwelling decomposers, produce significant amounts of feces that are distributed through different levels of the soil profile. Excrements enrich the soil with more easily and quickly absorbed compounds [9,10].

Most millipedes display a nocturnal lifestyle, their body color oscillate between shades of brown, beige or black, that help to hide. Other species may exhibit an aposematic bright coloration of the body, or even glow [11,12,13,14]. When stressed, millipedes hide the abdominal sensitive part of the body, head and legs through curling up into a ball or spiral fashion. Escaping by jumping away was also observed [15]. Penicillata entangle predator with thicket of bristles that cover their body [16]. These adaptations were caused by the poor motility that limits escape possibility within the dense litter. Millipedes, in the course of antipredatory and anti parasitic behavior excrete defensive secretions [4,17,18,19]. Tropical millipedes *Rhinocricus* sp. can spray their product for an approximately half of meter distance [20]. Diplopoda, due to the lack of structures and organs for active injection of secretions, belong to the group of poisonous animals [21].

Human habitats are commonly visited by millipedes especially during rainy period. Native and accidentally introduced foreign species should be considered as future synanthropic taxon or pest particularly in case of population gradation within migrating population [22,23].

AIM

The goal of this review was the evaluation of Diplopod's medical influences: threats of human health and future millipedes application prospects.

METHODS

The English- language articles review was extracted from Google Scholar, PubMed, Medline search base. Doctorate thesis bibliography, written by first author of this review, served as source, hence this article includes unique information, native published in polish.

DISCUSSION

DEFENSIVE SECRETIONS

Millipedes when disturbed expel defensive secretions. Representatives of the orders Callipodida Glomerida, Julida, Polydesmida Polyzoniida Spirobolida, Spirostreptida have repugnatorial glands [2,11]. The openings of the defensive glands for the most millipedes are located laterally on both sides, starting from the 5th-6th body segment on the metasomite or at the end of the paranotal projections (Polydesmida). Ozopores are distributed along the body segments dorsally for Pentazonia species [24,25,26]. Members of Chordeumatida, Sphaerotheriida, Polyxenida, Siphoniulida and Glomeridesmida lack repugnatorial glands [27]. Considering the structure of the gland and the composition of the defense secretion, Shear [27] after Eisner et al. [17] had listed three groups of defense glands: Glomerida, Julida and Polydesmida. Ilić et al. [28] complemented the list with fourth type: Colobognatha. Physical properties (color and odor), characterize taxon (Fig.1). Composition of defensive secretions can be sophisticated and same group of chemicals appear in different taxa, while others are restricted to specific order or species [25,27].

List of organic components include: quinazolinone alkaloids (glomerine, homoglomerine) (Glomerida); heterocyclic molecules substituted with nitrogen (for Polyzoniida); terpenes (pinen and limonene) (for Polyzoniida, Siphonophorida); benzoquinones, hydroquinones and their derivatives (for Julida, Spirobolida, Spirostreptida); phenols (for Callipodida, Julida, Polydesmida) [29-37]. Cyanogenic compound such as mandelonitrile; benzoyl cyanide are noted for all millipedes but few [38, 39]. Well documented in the literature, and thus extremely diverse in terms of chemical composition of defense substances, is the Juliformia superorder [19,27,37,40,41].



Fig. 1. *Blaniulus guttulatus* (Fabricius, 1798). Blind Julida member, discharging the characteristic pigmented defensive secretion. Photo Monika Jung.

MEDICAL IMPORTANCE, SHORT AND LONG TERM EFFECTS ALTERNATIONS

Reports on the threat of human health associated with defensive secretions concern on the possibility of local dermal and systemic malfunction or toxicosis [42,43]. Blackening erythematous lesions but also paresthesia was described by Neto et al. [44] on emergency room case report of 23-year-old female in Bangu, Rio de Janeiro. Odors emitted by the millipede *Oxidus gracilis* (C.L.Koch, 1847) (Polydesmida) cause headaches after prolonged exposure [45]. Julida; *Cylindroiulus caeruleocinctus* (Wood, 1864), *Cylindroiulus latestriatus* (Curtis, 1845), *Choneiulus palmatus* (Němec, 1895), *Ommatoiulus sabulosus* (Linnaeus 1758), *Leptoiulus proximus* (Němec, 1896) secrete para-benzoquinone known for their genotoxic effects. Methyl-1,4 benzoquinone present in the secretions of the Brazilian Spirobolida species *Rhinocricus padbergi* Verhof, 1938 and *Floridobolus penneri* Causey, 1957 leads to carcinogenesis [20, 32]. Benzoquinones ejected by tropical species when reach ocular area leads to lacrimation, periorbital oedema, conjunctivitis, keratitis followed by corneal ulceration. The oily and adhesive secretion, causes contact erythema and blisters [46].

The most common event is accidental rupture of a millipede hidden in clothing, footwear or leaf litter and provoke lesions; erythematous, purpuric, cyanotic [44,47]. Painful, burning sensation, dermatitis and necrosis of the limb and proximities appear and clinical picture depends upon chemical composition of the secretion and the time of exposure [42,48].

Prolonged contact with the skin and accompanying mucous membranes provoke recovery difficulties of the tissue with possible gangrene progression. Pathogenesis includes: superficial dermal injury, burns of the skin followed by blisters with discoloration that undergoes secondary infection. Browning, blackening and unusual skin abbreviations mask inflammation that can persists even for a few months [46,47,49-51].

MEDICAL ISSUES AND DISEASES RELATED

Incidental ingestion of millipedes leads to noninvasive colonization of human alimentary tract. Numerous *Nopoiulus kochii* (Gervais, 1847) were seen in excrement and vomited material [52]. Mowlavi et al. [53] reported *Brachyiulus lusitanus* Verhoeff, 1898 being free living inhabitant of gastrointestinal tract. According to Kielczewski [54] *Polydesmus complanatus* (Linnaeus, 1761) and *Julus terrestris* Linnaeus, 1758 provoked ailments similar to helminthosis. Stojałowska [55] after Brumpt [56] and Pawłowski [57] have listed *Cylindroiulus teutonicus* Pocock, 1900 *Julus terrestris* Linnaeus, 1758. and *P. complanatus* as pseudoparasites. Unpalatable taste is being masked by fruits and vegetables occasionally inhabited by foraging millipedes *Blaniulus guttulatus* (Fabricius, 1798). There is a possibility of oral administration of millipede's eggs or immature stages from drinking water [55].

Epidemiological issue is proposed also for tapeworm transmission. Vakarenko and Korniyushin [58] described larval cysticeroid: *Sobolevitaenia verulamii* (Cestoda, Cyclophyllidae), carried by *Glomeris connexa* C.

L. Koch, 1847 (Glomerida). Adult tapeworm is bird's (Turdinae) dwelling parasite. Diplopoda: *Fontaria virginensis* (Gray, 1832) (Polydesmida) and *Julus* sp. serve as intermediate hosts for larvae of the flat worm *Hymenolepis diminuta* (Cestoda, Cyclophyllidae). Humans may be the definitive host for this species, not to the mention the possibility of auto-invasion hence multiple infections [59,60].

Arthropods as vectors extend microorganisms passively or actively. The possible transmission of infectious diseases is being considered and relevant in case of swarm population gradation of migrating millipedes. Reports of passive transmission bacteria to humans by millipedes goes as follows: *C. caeruleocinctus* transfer: *Klebsiella ozaenae*, *Citrobacter freundii*, *Serratia marcescens* and *Pantoea agglomerans*, *Raoultella planticola*, *Salmonella arizonae*, *Xanthomonas maltophilia*. *O. sabulosus* transmits: *Klebsiella pneumoniae*, *Klebsiella oxytoca*, *Enterobacter sakazakii*, *Citrobacter freundii*, *Pantoea agglomerans*, *Hafnia alvei* and *Raoultella planticola* [20,61-66].

APPLICATIONS FOR MILLIPEDE'S SECRETIONS IN MEDICINE AND ZOO PHARMACOLOGY

Chemical secretions of millipedes, are well documented in the literature for their repellent or sedative and toxic properties [27,67,68]. Arachnids and vertebrates consuming millipedes *Glomeris marginata* (Villers, 1789) were poisoned by glomerin and homoglomerin [69]. Para-cresol is defensive allomone that deter ants [70, 71]. Mosquitoes *Aedes aegypti* fed less frequently during laboratory trial for blood samples with benzoquinones from *Orthoporus dorsovittatus* (Verhoeff, 1938) [67]. The long time effect of benzoquinones as preventive method against lone star tick, *Amblyomma americanum* was checked in laboratory tests by Carroll et al. [68].

Primates use millipedes directly to control blood-feeding arthropods. Rubbing living millipedes on the fur was observed for wedge capped capuchin monkey *Cebus olivaceus* [72]. Mixing defensive secretion with onion, garlic was noted by Weldon et al. [73] for capuchin monkey *Cebus apella* and by Zito et al. [74] in case of black lemur *Eulemur macaco* and owl monkeys *Aotus* spp.

Millipedes are usually spit out or vomited due to their repulsive smell and taste [75]. However secretion-resistant vertebrate enrich their diet with millipedes, with no symptoms of poisoning, predatory controls the population of millipedes in turn [55,73,76]. Diplopoda might be edible after some preparation and are used for their anti-pathogenic properties. A decoction from *Tachypodoiulus niger* (Leach, 1814) is used by the inhabitants of Mizoram in India against tuberculosis [77]. The Bobo tribe folks of Burkina Faso consume millipedes to improve the caloric content of the meal but also due the potential antimalarial effect [78]. As an element of prophylaxis against intestinal parasites the Red-fronted lemurs *Eulemur rufifrons*, eat millipedes after squeezing and preliminary chewing of arthropods which liberates defensive secretions [79]. Prey rolling behavior was noted for Coati *Nasua* spp. [73]. Representatives of the tropical batrachofauna choose millipedes to supplement their diet with valuable poisonous substances accumulated in body in order to deter predators. Alkaloids found in dermal

glands of poisonous frogs were derived from ingested ants and millipedes [80,81]. Spiropyrolizidine oximes found in the skin of *Dendrobates pumilio* living in Panama, are derived from chemical secretions from consumed millipedes *Rhinotus purpureus* (Pocock, 1894) and beetles [81-82]. Arthropoda are source of compounds of pharmacological properties, this therapeutic effect is used both in folk and conventional medicine [40,83,84]. Research are being carried out on the pharmacological use of batrachotoxin e.g. epibatidine alkaloid from skin glands of poisonous dendrobatidae *Epipedobates tricolor* [80]. Chemical is known as mediator of non-opioid receptors thus no addictive, analgesics but toxic in low doses. Nowadays investigations are being concerned on synthetic analogs with safe and selective therapeutic effect [85]. There are research on Glomeridae and Julidae secretion as antioxidant and suppressor of nerve degeneration factor [86-87].

MILLIPEDE'S SECRETIONS AND THEIR IMPACT ON HEALTH CARE

Diplopoda dwell ruderal places, heaps and spots of decomposition with composting materia [88-93]. Suspected function of secretions is support of survival in environments inhabited by variety of organisms including pathogens [94]. There are reports on Diplopod's antifungal and antibacterial characteristic detected for hemolymph or body extract [95-97].

Fungicidal and bactericidal activity of 1-octen-3-ol, from *Niponia nodulosa* Verhoeff, 1931 (Polydesmida) secretion was described by Omura et al [98]. Laboratory trials reveal inhibitory effect of *Pachyiulus hungaricus* (Karsch, 1881) (Julida) secretion on variety bacteria; *Aeromonas hydrophila*, *Pseudomonas aeruginosa*, *Escherichia coli*, *Xanthomonas arboricola*, *Listeria monocytogenes*, *Staphylococcus aureus* and *Bacillus subtilis* and fungi; *Aspergillus flavus*, *Aspergillus niger*, *Fusarium subglutinans*, *Fusarium semitectum*, *Fusarium equiseti* and *Gliocladium roseum* [87,99]. In vitro trials over *Megaphyllum unilineatum* (C. L. Koch, 1838) (Julida) defensive secretion reveal fungicidal action on *Fusarium* sp. [87]

Both p-cresol and phenol which are present in Julida secretions, effectively inhibit the development of fungal spores and mycelia. Phenol (carbolic acid) was formerly used for disinfection in health facilities. Phenolic derivatives form cave predator *Apfelbeckia insculpta* (C.L. Koch, 1867) (Callipodida) play antifungal and antibacterial activity [27, 100].

Benzoquinones and derivatives secreted by Julida, Spirobolida, and Spirostreptida exhibit toxicity against fungi, and bacteria and nematodes [83,87,99,101]. Antibiotic action against Gram-positive bacteria (including methicillin-resistant *S. aureus*) was described by Glukhova et al. [102]. In vitro research carried by teams; Stanković et al. [99], Pesewu et al. [101] proved that millipede extract (benzoquinones) were destructive for methicillin resistant *S. aureus* strain. Development of *S. aureus* was inhibited after organic acid and alkaloid defensive secretion compounds of *Tachypodoiulus* sp. [103].

Defensive secretion as mixture of benzoquinonoids from of *Rhinocricus* sp. possess fungicidal properties and bactericidal activity for both Gram positive, negative pathogens including *S. aureus* [104]. Diplopoda should be considered as a source of

multi complexes molecular systems of potential toxicity on pathogenic microorganisms.

Medical issue are also disorders regarded to human progressive industrial activity and environmental contamination. Trace element toxicosis which are achieved in low doses [105-107]. Millipedes can survive in contaminated area. The cooperation of edaphic organisms enable the creation of favorable conditions and habitats for development of pioneer species population, facilitating the process of secondary succession [88,108,111]. Environmental monitoring is based on zoo indicators and millipedes serve for habitat evaluation. Pedobionts succession and sarcophagus activity increase decomposition processes, therefore Diplopoda may help with reclamation of polluted area [88,108,109,111-120].

CONCLUSIONS

Diplopoda are frequently seen in human habitats and considering as a nuisance during swarming behavior and massive migrations. Although the most toxic secretions were described for tropical taxa and more severe clinical cases are noted from equatorial regions, native and alien species shifts their geographic distribution. Children are the most exposed group as result of their natural vigor and curiosity along with poor education of prophylaxis. Directly exposed to defensive section and the most frequently injured are bare body extremities or face and eyes. To identify chemicals and asses the threat the millipede sample should be delivered. Although most symptoms pass after a few days, dermal discoloration changes may persist and wounded skin is a root for secondary infection. There is a risk of passive transport of pathogenic bacteria to human settlements. Occupationally exposed are field and forestry workers, common inhabitants at leisure and work places. Education means of preventive measure and risk factors assessment are being neglected.

Millipedes are potential source for pharmacology development e.g. drug improvement. This progressive field of science is a great promise due to a growing demand and new applications being discovered. Diplopoda are crucial for proper microelement turn over hence play role in secondary succession and monitoring of environment reclamation. Ecotoxicology may help with prognosis of domestic area pollution and influence on health condition of inhabitants. Proper millipede's population control and prophylaxis measures could limit threats both direct and indirect for human health which escalate especially in synanthropization context.

FORMATTING OF FUNDING SOURCES

This research did not receive any specific grant from funding agencies in the public, commercial, or not-for-profit sectors.

CONFLICT OF INTEREST

The authors declared no conflict of interest.

REFERENCES

- Shelley RM. A revised, annotated, family-level classification of the Diplopoda. *Arthropoda Sel. Moscow*. 2002. 11: 187-207.
- David JF. Diplopoda- Ecology. in: Minelli A. (ed.), *Treatise on Zoology-Anatomy, Taxonomy, Biology. The Myriapoda*, Brill, Leiden-Boston, 2015, pp. 303-327.
- Enghoff H, Golovatch SI, Short M, Stoev P, Wesener T. Diplopoda- taxonomic overview. in: Minelli A. (ed.), *Treatise on Zoology-Anatomy, Taxonomy, Biology. The Myriapoda*, Brill, Leiden-Boston, 2015, pp. 363-453.
- Sierwald P, Bond JE. Current Status of the Myriapod Class Diplopoda (Millipedes): Taxonomic Diversity and Phylogeny. *Annu. Rev. Entomol.* 2007. 52: 401-420. <https://doi.org/10.1146/annurev.ento.52.111805.090210>.
- Golovatch SI, Kime RD. Millipede (Diplopoda) distributions: A review. *Soil Org.* 2009. 81: 565-597. <https://soil-organisms.org/index.php/SO/article/view/210>.
- Shear W. Class Diplopoda de Blainville Gervais, 1844, in: Zhang Z-Q. (ed.), *Animal biodiversity: An outline of higher-level classification and survey of taxonomic richness*, Zootaxa. 2011, pp. 159-164. <https://doi.org/10.11646/zootaxa.3148.1.32>.
- Brewer MS, Sierwald P, Bond JE. Millipede taxonomy after 250 years: classification and taxonomic practices in a mega-diverse yet understudied arthropod group. *PLoS One*. 2012. e37240. <https://doi.org/10.1371/journal.pone.0037240>.
- Sierwald P, Spelda J. MilliBase, VI, 2020. <http://www.millibase.org>, doi:10.14284/370.
- Kime RD, Golovatch SI. Trends in the ecological strategies and evolution of millipedes (Diplopoda). *Biol. J. Linn. Soc.* 2000. 69: 333-349. <https://doi.org/10.1111/j.1095-8312.2000.tb01209.x>.
- David JF. The role of litter-feeding macroarthropods in decomposition processes: a reappraisal of common views. *Soil Biol. Biochem.* Elsevier. 2014. 76: 109-118. <https://doi.org/10.1016/j.soilbio.2014.05.009>.
- Blower J.G. Millipedes, Keys and notes for the identification of the species, in: Kermack DM, Barnes RSK. (eds.), *Synopses of the British fauna*. Brill E.J., Buckhuys W., London, 1985, pp.1-242.
- Marek PE, Moore W. Discovery of a glowing millipede in California and the gradual evolution of bioluminescence in Diplopoda. *Proc. Natl. Acad. Sci. U.S.A.* 2015. 112: 6419-6424. <https://doi.org/10.1073/pnas.1500014112>.
- Srisonchai R, Enghoff H, Likhitrakarn N, Panha S. Four colorful new species of dragon millipedes, genus *Desmoxytes* Chamberlin, 1923, from northern Thailand (Diplopoda: Polydesmida: Paradoxosomatidae). *Zootaxa*. 2016. 4170: 93-113. doi: 10.11646/zootaxa.4170.1.4. PMID: 27701275.
- Means JC, Hennen DA, Marek PE. A revision of the *minor* species group in the millipede genus *Nannaria* Chamberlin, 1918 (Diplopoda, Polydesmida, Xystodesmidae), *Zookeys*. 2021. 1030: 1-180. <https://doi.org/10.3897/zookeys.1030.62544>.
- Evans MEG, Blower JG. A jumping millipede, *Nature*. 1973. 246: 427-428. <https://doi.org/10.1038/246427a0>.
- Eisner T, Eisner M, Deyrup M. Millipede defense: use of detachable bristles to entangle ants. *Proc. Natl. Acad. Sci. U.S.A.* 1996. 93: 10848-10851. <https://doi.org/10.1073/pnas.93.20.10848>.
- Eisner T, Alsop D, Hicks K, Meinwald J. Defensive secretions of millipedes. in: Bettini S. (ed.), *Handbook of Experimental Pharmacology Arthropod Venoms*. Springer-Verlag, Berlin, 1978. pp. 41-72.
- Deml R, Huth A. Benzoquinones and hydroquinones in defensive secretions of tropical millipedes. *Sci. Nat.* 2000. 87: 80-82. <https://doi.org/10.1007/s10886-017-0832-1>.
- Kania G, Kowalski R, Pietraś R. Defensive secretions in millipede species of the order Julida (Diplopoda). *Acta Soc. Zool. Bohemicae*. 2016. 80: 17-20.
- Kania G. The medical significance of millipedes (Diplopoda) – human consequences of their chemical secretions and transmission of pathogenic bacteria. in: Buczek A, Błaszczak Cz. (eds.), *Arthropods in urban and suburban environments*. Koliber, Lublin. 2017, pp. 169-186.
- Prokopowicz D. Rośliny trujące i zwierzęta jadowite. Wyd. Ekonomia i Środowisko, Białystok. 1999 pp. 207.
- Voigtländer K. Mass occurrences and swarming behaviour of millipedes (Diplopoda: Julidae) in Eastern Germany. *Peckiana*. 2005. 4: 181-187.
- Stoev P, Korsós Z. *Oxidus gracilis* (C.L. Koch, 1847)(Diplopoda, Polydesmida, Paradoxosomatidae) -14.2., in: Roques A, Lees D. (eds.), *Factsheets for 80 representative alien species*. BioRisk. 2010. 4: pp. 855-1021.
- Minelli A, Golovatch SI. Myriapods. in: Levin SA. (ed.), *Encyclopedia of Biodiversity*. Academic Press, Waltham, MA. 2001, pp. 421-432. <https://doi.org/10.1016/B978-0-12-384719-5.00208-2>.
- Makarov SE, Curčić BP, Tesević VV, Jadranin MB, Vujisić LV, Curčić SB, Mandić BM, Sekulić TLj, Mitic BM. Defensive secretions in three species of polydesmids (Diplopoda, Polydesmida, Polydesmidae), *J. Chem. Ecol.* 2010. 36: 978-982. <https://doi.org/10.1007/s10886-010-9847-6>.
- Geoffroy J. Subphylum Myriapoda, Class Diplopoda, fourth ed., in: James H, Thorp DCh, (eds.), *Thorp and Covich's Freshwater Invertebrates: Ecology and General Biology*. Elsevier/Academic Press, Amsterdam. 2015.
- Shear W. The chemical defenses of millipedes (diplopoda): Biochemistry, physiology and ecology. *Biochem. Syst. Ecol.* 2015. 61: 78-117. <https://doi.org/10.1016/j.bse.2015.04.033>.
- Ilić B, Unković N, Ćirić A, Glamočlija J, Ljaljević Grbić M, Raspoćnić G, Bodner M, Vukojević J, Makarov S. Phenol-based millipede defence: antimicrobial activity of secretions from the Balkan endemic millipede *Apfelbeckia insculpta* (L. Koch, 1867) (Diplopoda: Callipodida). *Sci. Nat.* 2019. 106: 1-37. <https://doi.org/10.1007/s00114-019-1631-z>.
- Shear WA, Jones TH, Wesener T. Glomerin and homoglomerin from the North American pill millipede *Onomeris sinuata* (Loomis, 1943) (Diplopoda, Pentazonia, Glomeridae). *Int. J. Myriap.* 2011. 94: 1-10. <https://doi.org/10.3897/ijm.4.1105>.
- Kuwahara Y, Mori N, Tanabe T. Detection of a neotropical frog alkaloid spiropyrrolizidine 236 from a Japanese polyzoniid millipede *Kiusiozonium okai* as a major defense component together with polyzonimine and nitropolyzonimine. *Jpn. J. Environ. Ent. Zool.* 2007. 18: 91-96.
- Wood W, Hanke F, Kubo I, Carroll J, Crews P. Buzonamine, a new alkaloid from the defensive secretion of the millipede, *Buzonium crassipes*. *Biochem. Syst. Ecol.* 2000. 28: 305-312. [https://doi.org/10.1016/S0305-1978\(99\)00068-X](https://doi.org/10.1016/S0305-1978(99)00068-X).
- Arab A, Zacarin GG, Fontanetti CS, Camargo-Mathias MI, Dos Santos MG, Cabrera AC. Composition of the defensive secretion of the neotropical millipede *Rhinocricus padbergi* Verhoeff 1938 (Diplopoda: Spirobolida: Rhinocricidae). *Entomotropica*. 2003. 18: 79-82.
- Wu X, Buden DW, Attygalle AB. Hydroquinones from defensive secretion of a giant Pacific millipede, *Acladocricus setigerus* (Diplopoda: Spirobolida), *Chemoecology*. 2007. 17: 131-138. <https://doi.org/10.1007/s00049-007-0372-1>.
- Bodner M, Raspoćnić G. Millipedes that smell like bugs: (E)-Alkenals in the defensive secretion of the julid diplopod *Allajulus dicentrus*. *J. Chem. Ecol.* 2012. 38: 547-556. <https://doi.org/10.1007/s10886-012-0127-5>.
- Sekulić TLj, Vujisić LJ, Ćurčić BPM, Mandić BM, Antić DŽ, Trifunović SS, Godevac DM, Vajs EV, Tomić VT, Makarov S.E. Quinones and non-quinones from the defensive secretion of *Unciger transsilvanicus* (Verhoeff, 1899) (Diplopoda, Julida, Julidae), from Serbia. *Arch. Biol. Sci. Belgr.* 2014. 66: 385-390. <https://doi.org/10.2298/ABS1401385S>.
- Shimizu N, Kuwahara Y, Yakamaru R, Tanabe T. n-Hexyl laurate and fourteen related fatty-acid esters: new secretory compounds from the julid millipede, *Anaulaciulus* sp. *J. Chem. Ecol.* 2012. 38: 23-28. <https://doi.org/10.1007/s10886-012-0063-4>.
- Bodner M, Vagalinski B, Makarov SE, Antić DŽ, Vujisić LV, Leis HJ, Raspoćnić G. "Quinone Millipedes" Reconsidered: Evidence for a Mosaic-Like Taxonomic Distribution of Phenol-Based Secretions across the Julidae. *J. Chem. Ecol.* 2016. 42: 249-258. <https://doi.org/10.1007/s10886-016-0680-4>.
- Kuwahara Y, Shimizu N, Tanabe T. Release of hydrogen cyanide via a post-secretion Schotten-Baumann reaction in defensive fluids of Polydesmoid millipedes. *J. Chem. Ecol.* 2011. 37: 232-238. <https://doi.org/10.1007/s10886-011-9920-9>.
- Ishida Y, Kuwahara Y, Dadashpour M. A sacrificial millipede altruistically protects its swarm using a drone blood enzyme, mandelonitrile oxidase. *Sci. Rep.* 2016. 6: 1-10. <https://doi.org/10.1038/srep26998>.
- Ilić B, Dimkić I, Unković N, Ljaljević Grbić M, Vukojević J, Vujisić L, Tešević V, Stanković S, Makarov S, Lučić L. Millipedes vs. pathogens: defensive secretions of some julids (Diplopoda: Julida) as potential antimicrobial agents. *J. Appl. Entomol.* 2018. 142: 775-791. <https://doi.org/10.1111/jen.12526>.

41. Rodriguez J, Jones T.H, Sierwal P, Marek P, Shear W, Brewer M, Kocot K, Bond J. Step-wise evolution of complex chemical defenses in millipedes: a phylogenomic approach. *Sci. Rep.* 2018. 8: <https://doi.org/10.1038/s41598-018-19996-6>.
42. Haddad V.Jr., Cardoso JL, Lupi O, Tying SK. Tropical dermatology: venomous arthropods and human skin: Part II. Diplopoda Chilopoda, and Arachnida. *J. Am. Acad. Dermatol.* 2012. 67: 1-9. <https://doi.org/10.1016/j.jaad.2012.05.028>.
43. Lofgran T, Warrington SJ. Millipede Envenomation. StatPearls, Treasure Island (FL): StatPearls Publishing, VI, 2022. <https://www.ncbi.nlm.nih.gov/books/NBK557454/>
44. Neto A, Fred SH, Filho B, Martins G. Skin lesions simulating blue toe syndrome caused by prolonged contact with a millipede. *Rev. Soc. Bras. Med. Trop.* 2014. 47: 257-258. <http://dx.doi.org/10.1590/0037-8682-0212-2013>.
45. Taira J, Nakamura K, Higa Y. Identification of secretory compounds from the millipede, *Oxidus gracilis* C.L. Koch (Polydesmida: Paradoxosomatidae) and their variation in different habitats. *Appl. Ent. Zool.* 2003. 38: 401- 404. <https://doi.org/10.1303/aez.2003.401>.
46. De Capitani EM, Vieira RJ, Bucarechi F, Fernandes LCR, Toledo AS, Camargo AC. Human accidents involving *Rhinocricus* spp., a common millipede genus observed in urban areas of Brazil. *Clin. Toxicol.* 2011. 49: 187-190. <https://doi.org/10.3109/15563650.2011.560855>.
47. Lima CA., Cardoso JL, Magela A, Oliveira FG, Talhari S, Haddad V.Jr. Exogenous pigmentation in toes feigning ischemia of the extremities: a diagnostic challenge brought by arthropods of the Diplopoda Class ("millipedes"). *An Bras Dermatol.* 2010. 85: 391-392. <https://doi.org/10.1590/s0365-05962010000300018>.
48. Lacy FA, Elston DM. What's Eating You? Millipede Burns, *Cutis.* 2019. 103: 195-196.
49. Shpall S, Frieden I. Mahogany discoloration of the skin due to the defensive secretion of a millipede, *Pediatr. Dermatol.* 1991. 8: 25-27. <https://doi.org/10.1111/j.1525-1470.1991.tb00834.x>.
50. Haddad V. Jr., Cardoso JLC. Accidents provoked by millipede with dermatological manifestations: report of two cases. *An. Bras. Dermatol.* 2000. 75: 471-474. <https://doi.org/10.1016/j.abd.2019.10.003>.
51. Pennini SN, Rebello PFB, Guerra MGV, Talhari S. Millipede accident with unusual dermatological lesion, *An. Bras. Dermatol.* 2019. 94: 765-767. <https://doi.org/10.1016/j.abd.2019.10.003>.
52. Ertek M, Aslan I, Yazgi H, Torun HC, Ayyildiz A, Tasyaran MA. Infestation of the human intestine by the millipede, *Nopoiulus kochii*. *Med. Vet. Entomol.* 2004. 18: 306-307. <https://doi.org/10.1111/j.0269-283X.2004.00507.x>.
53. Mowlavi G, Naddaf SR, Rezaeian M, Najafi N, Lucio-Forster A, Bowman DD. Apparent pseudoparasitism of the alimentary canal of a 5-year-old child by the millipede *Brachyiulus lusitanus* (Diplopoda: Julidae). *Parasite.* 2009. 16: 161-163. <https://doi.org/10.1051/parasite/2009162161>.
54. Kielczewski B, Gromada Wijs (Myriapoda). w: Żółtowski Z. (red.), *Arachnoentomologia lekarska*. Wydawnictwo Lekarskie PZWL, Warszawa. 1976; ss. 90-93.
55. Stojalowska W, Krocionogi (Diplopoda) Polski. Wyd. 1. (first ed.), Państwowe Wydawnictwo Naukowe, Warszawa. 1961.
56. Brumpt E. Précis de Parasitologie, sixth ed. Masson ET Cie Editeurs, Paris. 1949.
57. Pawłowski E. Wytyczne do parazytologii człowieka. Wyd. 1. Moskwa- Leningrad. 1948
58. Vakarenko EG, Kornysushin VV. Description of a cysticercoid of *Soboleviaenia verulamii* (Cestoda, Cyclophyllidae) from its intermediate host *Glomeris connexa* (Diplopoda, Glomeridae). *Vest Zool.* 2002. 36: 61-64.
59. Beaver PC, Jung RC, Cupp EW. Cyclophyllidean Tapeworms. in: *Clinical Parasitology*, Lea and Febiger. Philadelphia, 1984, pp. 505-543.
60. Kadłubowski R, Kurnatowska A. Zarys parazytologii lekarskiej. Wyd. VII (seventh ed.), Wydawnictwo Lekarskie PZWL, Warszawa, 1999.
61. Jarosz J, Kania G. The question of whether gut microflora of the millipede *Ommatoiulus sabulosus* could function as a threshold to food infections. *Pedobiologia.* 2000. 44: 705-708. [https://doi.org/10.1078/S0031-4056\(04\)70083-9](https://doi.org/10.1078/S0031-4056(04)70083-9).
62. Byzov BA. Intestinal microbiota of millipedes. in: König H, Varma A. (eds.), *Soil Biology. Intestinal microorganisms of termites and other Invertebrates*, Springer-Verlag, Berlin, Heidelberg. 2006. 6: pp. 90-114.
63. Kania G. The economic and medical significance of millipedes (Diplopoda) with emphasis on *Ommatoiulus sabulosus*. in: Tajovský K, Schlagamersky J, Pizl V. (eds.), *Contributions to Soil Zoology in Central Europe III*. Ceske Budejovice. 2009. pp. 73-77.
64. Kania G, Kłapeć T. Seasonal activity of millipedes (Diplopoda) – their economic and medical significance. *Ann. Agric. Environ. Med.* 2012. 19: 646-650.
65. Dutkiewicz J, Mackiewicz B, Lemieszek MK, Golec M. *Pantoea agglomerans*: a mysterious bacterium of evil and good. Part III, Deleterious effects: infections of humans, animals and plants. *Ann. Agric. Environ. Med.* 2016. 23: 197-205. <https://doi.org/10.5604/12321966.1203878>.
66. Alagesan P. Millipedes as host for microbes—a review, *Int. J. Microbiol. Res.* 2017. 8: 19-24. <https://doi.org/10.5829/idosi.ijmr.2017.19.24>.
67. Weldon PJ, Aldrich JR, Klun JA, Oliver JE, Deboun M. Benzoquinones from millipedes deter mosquitoes and elicit self-anointing in capuchin monkeys (*Cebus* sp.). *Sci. Nat.* 2003. 90: 301-304. <https://doi.org/10.1007/s00114-003-0427-2>.
68. Carroll JF, Kramer M, Weldon PJ, Robbins RG. Anointing chemicals and ectoparasites: effects of benzoquinones from millipedes on the lone star tick, *Amblyomma americanum*. *J. Chem. Ecol.* 2005. 31: 63-75. <https://doi.org/10.1007/s10886-005-0974-4>.
69. Alagesan P. Millipedes: Diversity, distribution and ecology. in: Chakravarthy AK, Sridhara S. (eds.), *Arthropod Diversity and Conservation in the Tropics and Sub-tropics*, Springer. 2016. pp. 119-137.
70. Shear W, Jones T. Miras H. A possible phylogenetic signal in milliped chemical defenses: the polydesmidan millipede *Leonardesmus injucundus* Shelley & Shear secretes p-cresol and lacks a cyanogenic apparatus (Diplopoda, Polydesmida, Nearctodesmidae). *Biochem. Syst. Ecol.* 2007. 35: 838-842. <https://doi.org/10.1016/j.bse.2007.01.005>.
71. Sekulić TLJ., Vujić LJ., Čurčić BPM, Mandić BM, Antić DŽ, Trifunović SS, Godevac DM, Vajs EV, Tomić VT, Makarov SE. Quinones and non-quinones from the defensive secretion of *Unciger transsilvanicus* (Verhoeff, 1899) (Diplopoda, Julida, Julidae), from Serbia. *Arch. Biol. Sci.* 2014. 66: 385-390. <https://doi.org/10.2298/ABS1401385S>.
72. Valderrama X, Robinson JG, Attygalle AB, Eisner T. Seasonal anointment with millipedes in a wild primate: a chemical defense against insects? *J. Chem. Ecol.* 2000. 26: 2781-2790. <https://doi.org/10.1023/A:1026489826714>.
73. Weldon P, Cranmore C, Chatfield J. Prey-rolling behavior of coatis (*Nasua* sp.) is elicited by benzoquinones from millipedes. *Sci. Nat.* 2006. 93: 14-16. <https://doi.org/10.1007/s00114-005-0064-z>.
74. Zito M, Evans S, Weldon PJ. Owl monkeys (*Aotus* spp.) self-anoint with plants and millipedes. *Folia Primatol.* 2003. 74: 159-161. <https://doi.org/10.1159/000070649>.
75. Olivieri N, Capone F, Puopolo M, Santucci D, Allea E. Response of CD-1 mice to the chemical defence of a common arthropod (*Ommatoiulus sabulosus*). *Physiol. Behav.* 2001. 74: 305-311. [https://doi.org/10.1016/s0031-9384\(01\)00563-7](https://doi.org/10.1016/s0031-9384(01)00563-7).
76. Jung M, Kania G. Wrogowie naturalni krocionogów (Diplopoda) - drapieżne kręgowce. (Natural enemies of millipedes (Diplopoda), Vertebrate predators. in: Maciąg M, Maciąg K. (red.), *Makroorganizmy-przegląd wybranych badań*, Lublin 2018, Wydawnictwo Naukowe Tygiel, Lublin. 2018. pp. 31-45.
77. Chinlampaing M, Singh RK, Shukla AC. Ethnobiological diversity of northeast India: empirical learning with traditional knowledge holders of Mizoram and Arunachal Pradesh. *Ind. J. Trad. Knowledge.* 2013. 12: 18-30. <http://nopr.niscares.in/handle/123456789/15342>
78. Enghoff H, Mann N, Tchibozo S, List M, Schwarzing B, Schoefberger W, Schwarzing C, Paoletti MG. Millipedes as food for humans: their nutritional and possibly antimalarial value—a first report, eCAM, Hindawi Publishing Corporation Evidence-Based Complementary and Alternative Medicine, IX, 2014. <https://doi.org/10.1155/2014/651768>.
79. Peckre LR, Defolie Ch, Kappeler PM, Fichtel C. Potential self-medication using millipede secretions in red-fronted lemurs: combining anointment and ingestion for a joint action against gastrointestinal parasites? *Primates.* 2018. 59: 483-494. <https://doi.org/10.1007/s10329-018-0674-7>.
80. Daly JW, Garraffo HM, Jain P, Spande TF, Snelling RR, Jaramillo C, Rand S. Arthropod–Frog connection: Decahydroquinoline and Pyrrolizidine alkaloids common to microsymbiotic myrmicine ants and dendrobatid frogs. *J. Chem. Ecol.* 2000. 26: 73-85. <https://doi.org/10.1023/A:1005437427326>.
81. Clark VC, Raxworthy ChR, Rakotomalala V, Sierwald P, Fisher BL. Convergent evolution of chemical defense in poison frogs and arthropod prey between Madagascar and the Neotropics. *Proc. Natl. Acad. Sci. USA* 2005. 102: 11617-11622. <https://doi.org/10.1073/pnas.0503502102>.
82. Saporito RA, Donnelly MA, Hoffman R.L., Garraffo HM, Daly JW. A siphonotid millipede (Rhinetus) as the source of spiropyrrolizidine oximes of dendrobatid frogs. *J. Chem. Ecol.* 2003. 29: 2781-2786. DOI: 10.1023/b:joe.000008065.28364.a0
83. Dandawate PR, Vyas AC, Padhye SB, Singh MW, Baruah JB. Perspectives on medicinal properties of benzoquinone compounds. *Mini-Rev. Med. Chem.* 2010. 10: 436-454. DOI: 10.2174/138955710791330909
84. Meyer-Rochow VB. Therapeutic arthropods and other, largely terrestrial, folk-medicinally important invertebrates: a comparative survey and review. *J. Ethnobiol. Ethnomed.* 2017. 13: 1-31. DOI: 10.1186/s13002-017-0136-0

85. Salehi B, Sestito S, Rapposelli S, Peron G, Calina D, Sharifi-Rad M, Sharopov F, Martins N, Sharifi-Rad J. Epibatidine: A Promising Natural Alkaloid in Health, Biomolecules. 2018. 23: 10. <https://doi.org/10.3390/biom9010006>
86. Ambarish CN, Sridhar KR. Pill-millipedes (Arthrosphaera: Sphaerotheriidae) of the Western Ghats: a future source of antioxidants? Int. J. Trop. Insect Sci. 2015. 35: 117-124. DOI: <https://doi.org/10.1017/S1742758415000132>
87. Ilić B, Unković N, Knežević A, Savković Ž, Ljaljević Grbić M, Vukojević J, Jovanović Z, Makarov S, Lučić L. Multifaceted activity of millipede secretions: Antioxidant, antineurodegenerative, and anti-Fusarium effects of the defensivesecretions of *Pachyiulus hungaricus* (Karsch, 1881) and *Megaphyllium unilineatum* (C. L. Koch, 1838) (Diplopoda: Julida), PLoS One. 2019. 14:e0209999. DOI: 10.1371/journal.pone.0209999
88. Voigtländer K, Balkenhol B. Studies on millipede assemblages (Myriapoda, Diplopoda) as influenced by habitat qualities of afforested mine sites. Nor. J. Entomol. 2006. 53: 345-360.
89. Nogarol LR, Fontanetti CS. Acute and subchronic exposure of diplopods to substrate containing sewage mud: tissular responses of the midgut, Micron. 2010. 41: 239-246. <https://doi.org/10.1016/j.micron.2009.10.009>
90. Steel H, Bert W. Biodiversity of compost mesofauna and its potential as an indicator of the composting potential status, Dynamic soil, Dynamic plant. 2012. 2: 45-50. <https://biblio.ugent.be/publication/1996013/file/2007101>
91. Braschler B, Gilgado JD, Zwahlen V, Rusterholz HP, Buchholz S, Baur B. Ground-dwelling invertebrate diversity in domestic gardens along a rural-urban gradient: Landscape characteristics are more important than garden characteristics. PLoS One. 2020. 15: e0240061. doi: 10.1371/journal.pone.0240061
92. Gilgado JD. Hidden in plain sight: six millipede species (Myriapoda: Diplopoda) new for the fauna of Switzerland. Rev. Suisse Zool. 2020. 127: 249-259. DOI: 10.35929/RSZ.0019
93. Haľková B, Drabová M, Mock A. An annotated checklist of millipede fauna from Slovakia, with ecological and biogeographic characteristics. Biodivers. Data J. 2021. 9: e71495. <https://doi.org/10.3897/BDJ.9.e71495>
94. Anitha C, Basil-Rose M.R. Characterization of naturally occurring agglutinin from the midgut of the rusty millipede *Trigoniulus corallines*. J. Global Trends Pharm. Sci. 2018. 9: 4968-4977.
95. Roncadori R.W, Duffey SS, Blum MS. Antifungal activity of defensive secretions of certain millipedes. Mycologia, 1985. 72: 185-191. <https://doi.org/10.2307/3793067>
96. Xyländer W. Antibacterial substances and characteristics of the hemolymph of chilopoda and diplopoda (Myriapoda, Arthropoda). Soil Organisms. 2009. 81: 413-429. <https://www.soil-organisms.org/index.php/SO/article/view/37>
97. Billah MK, Kwang D, Adofo C, Olu-Taiwo MA, Pesewu GA. Antibacterial activities of millipede extracts against selected bacterial pathogens. Journal of Microbiology and Antimicrobial Agents. 2015. 1: 3-35.
98. Omura H, Kuwahara Y, Tanabe T. 1-octen-3-ol together with geosmin: new secretion compounds from a polydesmid millipede, *Niponia nodulosa*. J. Chem. Ecol. 2002. 28: 2601-2612. DOI: 10.1023/a:1021400606217
99. Stanković S, Dimkić I, Vujisić L, Pavković-Lučić S, Jovanović Z, Stević T. Chemical defense in a millipede: evaluation and characterization of antimicrobial activity of the defensive secretion from *Pachyiulus hungaricus* (Karsch, 1881) (Diplopoda, Julida, Julidae), PLoS ONE. 2016. 11: e0167249. <https://doi.org/10.1371/journal.pone.0167249> PMID: 27907048
100. Ilić B, Unković N, Čirić A, Glamočlija J, Ljaljević Grbić M, Raspotnić G, Bodner M, Vukojević J, Makarov S. Phenol-based millipede defence: antimicrobial activity of secretions from the Balkan endemic millipede *Apfelbeckia insculpta* (L. Koch, 1867) (Diplopoda: Callipodidae). Sci. Nat. 2019. 106: 37. DOI: 10.1007/s00114-019-1631-z
101. Pesewu GA, Maxwell KB, Christian A, Douglas K, Olu-Taiwo AM, Osei-Djarbeng S. Antibacterial Activities of Millipedes Extracts against Methicillin-Resistant *Staphylococcus aureus* (MRSA). Am. Int. J. Contemp. Res. 2015. 2: 1-13.
102. [102] Glukhova AA, Karabanova AA, Yakushev AV, Semeniyuk II, Boykova YV, Malkina ND, Efimenko TA, Ivankova TD, Terekhova LP, Efremenkova OV. Antibiotic Activity of Actinobacteria from the Digestive Tract of Millipede *Nedyopus dawydoffiae* (Diplopoda), *Antibiotics*. 2018. 7: 94. <https://doi.org/10.3390/antibiotics7040094>
103. Ngbolua KTN, Ngunde-te-Ngunde S, Tshidibi DJ, Lengbiye ME, Mpiana PT, Ekutsu EG, Munene JJMM, Gbolo ZB, Bongo NG, Nzemu G. Anti-Sickling and Antibacterial Activities of Extracts from a Congolese Diplopod (*Tachypodoiulus* sp., Arthropoda). J Adv Bot Zool. 2014. 1: 1-5. <https://doi.org/10.5281/zenodo.913641>
104. Tacoronte Morales JE, Cruel Sigüenza J, Canchingre Bone ME, Bernal Villavicencio C, Cabrera Pedros MT. Antimicrobial activity of defensive secretions of terrestrial invertebrates (Diplopoda, Spirobolida, *Rhinocricus*) from the insular neotropics. in: Proceedings of the 2nd International Electronic Conference on Antibiotics—Drugs for Superbugs: Antibiotic Discovery, Modes of Action And Mechanisms of Resistance, Med. Sci. Forum. 2022. <https://doi.org/10.3390/eca2022-12729>.
105. Walker CH, Hopkin SP, Sibly RM, Peakall DB. w: Migula P (red.), Środowisko. Podstawy ekotoksykologii. Polskie Wydawnictwo Naukowe, Warszawa. 2002. 373 pp.
106. Hobbelen PHF, van den Brink PJ, Hobbelen JF, van Gesten CAM. Effects of heavy metals on the structure and functioning of detritivore communities in a contaminated floodplain area, Soil Biol. Biochem. 2006. 38: 1596-1607. <https://doi.org/10.1016/j.soilbio.2005.11.010>
107. Draszawka-Bolzan B, Cyranik E. Circuit heavy metals in nature, World Scientific News. 2014. 4: 32-38.
108. Berg MP, Hemerik L. Secondary succession of terrestrial isopod, centipede and millipede communities in grasslands under restoration, Biol. Fertil. Soils. 2004. 40: 163-170. DOI: 10.1007/s00374-004-0765-z
109. Grelle C, Fabre M, Lepretre A, Descamps M. Myriapod and isopod communities in soils contaminated by heavy metals in northern France, Eur. J. Soil Sci. 2000. 51: 425-433. <https://doi.org/10.1046/j.1365-2389.2000.00317.x>
110. Nahmani J, Lavelle P. Effects of heavy metal pollution on soil macrofauna in a grassland of Northern France, Eur. J. Soil Biol. 2002. 38: 297-300. [https://doi.org/10.1016/S1164-5563\(02\)01169-X](https://doi.org/10.1016/S1164-5563(02)01169-X)
111. Tajovský K. Colonization of colliery spoil heaps by millipedes (Diplopoda) and terrestrial Isopods (Oniscidea) in the Sokolov Region. Czech Republic. Restor. Ecol. 2001. 9: 365-369. DOI: 10.1046/j.1526-100X.2001.94005.x
112. Dunger W, Voigtländer K. Soil fauna (Lumbricidae, Collembola, Diplopoda and Chilopoda) as indicators of soil eco-subsystem development in post-mining sites of eastern Germany—a review. Soil Org. 2009. 81: 1–51. <https://soil-organisms.org/index.php/SO/article/view/184>
113. Godoy JAP, Fontanetti CS. Diplopods as bioindicators of soils: analysis of midgut of individuals maintained in substrate containing sewage sludge. Water Air Soil Pollut. 2010. 210: 389-398. DOI: 10.1007/s11270-009-0261-z
114. Kania G, Lechowski J. Bioaccumulation of some elements in the millipede *Glomeris hexasticha* (Brandt, 1833) (Diplopoda, Glomerida). J. Elem. 2014. 19: 155-164. DOI: 10.5601/jelem.2014.19.1.595
115. Kania G. Znaczenie krocionogów (*Diplopoda*) w ekotoksykologii. Czy Diplopoda są bioindykatorami środowiska?, w: Maciąg K, Danielewska A. (red.), Wybrane zagadnienia z zakresu nauk biologicznych i weterynaryjnych. Wydawnictwo Naukowe TYGIEL Sp. z o. o., Lublin. 2020. 88-98. <http://bc.wydawnictwo-tygiel.pl/publikacja/2962C5AA-CEDA-7383-E336-20DB1A4FB554>
116. Mekonen S. Soil Fauna as Webmasters, Engineers and Bioindicators in Ecosystems: Implications for Conservation Ecology and Sustainable Agriculture. Am. J. Life Sci. 2019. 7: 17-26. doi: 10.11648/j.ajls.20190701.14
117. Nakamura K, Taira J. Distribution of elements in the millipede, *Oxidus gracilis* C. L.Koch (Polydesmida: Paradoxosomatidae) and the relation to environmental habitats. Bio-Metals, 2005. 18: 651-658. DOI: 10.1007/s10534-005-4575-z
118. da Silva Souza T, Christoforetti CA, Bozzatto V, Fontanetti CS. The use of diplopods in soil ecotoxicology—a review, Ecotoxicol. Environ. Saf. 2014. 103: 68-73. doi: 10.1016/j.ecoenv.2013.10.025.
119. Tracz H. Diplopoda, Chilopoda i Isopoda w waloryzacji ekosystemów leśnych Leśnego Kompleksu Promocyjnego „Lasy Spalsko-Rogowskie”, Studia i Materiały Centrum Edukacji Przyrodniczo Leśnej w Rogowie. 2013. 35: 36-47. <https://cepl.sggw.edu.pl/studia-i-materialy-cepl-31-40/>
120. Tuf IH, Tufová J. Proposal of ecological classification of centipede, millipede and terrestrial isopods faunas for evaluation of habitat quality in Czech Republic, Čas. Slez. Muz. Opava, 2008. 57: 37-44. <http://ekologie.upol.cz/ad/tuf/pdf/papers/Tuf+Tufova2008>

Barbara Nieradko-Iwanicka

Zakład Higieny i Epidemiologii Uniwersytet Medyczny w Lublinie
 ul Chodźki 7, 20-093 Lublin
 e-mail: barbara.nieradko-iwanicka@umlub.pl

